

BUDAPEST TREATY ON THE INTERNATIONAL
RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

Mogen International N.V.
Einsteinweg 97
2333 CB LEIDEN
Nederland

name and address of depositor

RECEIPT IN THE CASE OF AN ORIGINAL DEPOSIT
issued pursuant to Rule 7.1 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified at the bottom of this page

jc511 U.S. PTO

09/258031



02/25/99

I. IDENTIFICATION OF THE MICROORGANISM

Identification reference given by the
DEPOSITOR:

E. coli DH5 alpha strain / the plasmid
pMOG800

Accession number given by the
INTERNATIONAL DEPOSITARY AUTHORITY:

CBS 414.93

II. SCIENTIFIC DESCRIPTION AND/OR PROPOSED TAXONOMIC DESIGNATION

The microorganism identified under I above was accompanied by:



a scientific description



a proposed taxonomic designation

(mark with a cross where applicable)

III. RECEIPT AND ACCEPTANCE

This International Depositary accepts the microorganism identified under I above, which was
received by it on Thursday, 12 August 1993 (date of the original deposit)¹

IV. RECEIPT OF REQUEST FOR CONVERSION

The microorganism identified under I above was received by this International Depositary
Authority on not applicable (date of the original deposit) and a
request to convert the original deposit to a deposit under the Budapest Treaty was received by
it on not applicable (date of receipt of request for conversion)

V. INTERNATIONAL DEPOSITARY AUTHORITY

Name: Centraalbureau voor Schimmelcultures

Address: Oosterstraat 1
P.O. Box 273
3740 AG BAARN
The Netherlands

Signature(s) of person(s) having the power to
represent the International Depositary
Authority or of authorized official(s):

drs F.M. van Asma

Date: Friday, 13 August 1993

¹ Where Rule 6.4(d) applies, such date is the date on which the status of international
depository authority was acquired.

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FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

Mogen International N.V.
Einsteinweg 97
2333 CB LEIDEN
Nederland

*name and address of the party to whom the
viability statement is issued*

VIABILITY STATEMENT


issued pursuant to Rule 10.2 by the
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I. DEPOSITOR	II. IDENTIFICATION OF THE MICROORGANISM
<p>Name: Mogen International N.V.</p> <p>Address: Einsteinweg 97 2333 CB LEIDEN Nederland</p>	<p>Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY:</p> <p>CBS 414.93</p> <p>Date of the deposit or of the transfer:¹</p> <p>Thursday, 12 August 1993</p>
III. VIABILITY STATEMENT	
<p>The viability of the microorganism identified under II above was tested on Friday, 13 August 1993². On that date, the said microorganism was</p> <p><input checked="" type="checkbox"/>³ viable</p> <p><input type="checkbox"/>³ no longer viable</p>	

¹ Indicate the date of the original deposit or, where a new deposit or a transfer has been made,
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IV. CONDITIONS UNDER WHICH THE VIABILITY HAS BEEN PERFORMED ⁴	
V. INTERNATIONAL DEPOSITARY AUTHORITY	
Name: Centraalbureau voor Schimmelcultures	Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):
Address: Oosterstraat 1 P.O. Box 273 3740 AG BAARN The Netherlands	 Date: Friday, 13 August 1993
	drs F.M. van Asma

⁴ Fill in if the information has been requested and if the results of the test were negative.



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(21) International Application Number: PCT/EP97/04923 (22) International Filing Date: 4 September 1997 (04.09.97) (30) Priority Data: 96202466.7 4 September 1996 (04.09.96) EP (34) Countries for which the regional or international application was filed: NL et al. 97200831.2 19 March 1997 (19.03.97) EP (34) Countries for which the regional or international application was filed: NL et al. (71) Applicant (for all designated States except US): MOGEN INTERNATIONAL N.V. [NL/NL]; Einsteinweg 97, NL-2333 CB Leiden (NL). (72) Inventors; and (75) Inventors/Applicants (for US only): STUIVER, Maarten, Hendrik [NL/NL]; Groenhoevelaan 71, NL-2343 BR Oegstgeest (NL). CUSTERS, Jérôme, Hubertus, Henricus, Victor [NL/NL]; Nieuwstraat 25D, NL-2312 KA Leiden (NL). SELA-BUURLAGE, Marianne, Beatrix [NL/IL]; Kefar, 108 Bilu (IL). MELCHERS, Leo, Sjoerd [NL/NL]; Wilhelmina Bladergroenweg 45, NL-2331 BZ Leiden (NL). VAN DEVENTER-TROOST, Johanna, Pietermella, Els [NL/NL]; Justus van Schoonhovenstraat 49, NL-2613	NX Delft (NL). LAGEWEG, Wessel [NL/NL]; Speet 33, NL-1141 BX Monnickendam (NL). PONSTEIN, Anne, Silene [NL/NL]; Meerforel 5, NL-2318 MR Leiden (NL). (74) Agent: VAN WEZENBEEK, Bart; Mogen International N.V., Einsteinweg 97, NL-2333 CB Leiden (NL). (81) Designated States: AL, AU, BA, BB, BG, BR, CA, CN, CU, CZ, EE, GE, HU, IL, IS, JP, KP, KR, LC, LK, LR, LT, LV, MD, MG, MK, MN, MX, NO, NZ, PL, RO, SD, SG, SI, SK, SL, TR, TT, UA, UG, US, UZ, VN, YU, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published Without international search report and to be republished upon receipt of that report.	
(54) Title: ANTIFUNGAL PROTEINS, DNA CODING THEREFORE, AND HOSTS INCORPORATING SAME		
(57) Abstract		
<p>The present invention provides an isolated protein obtainable from a plant source which has antifungal activity, specifically anti-<i>Phytophthora</i> activity and/or anti-<i>Pythium</i> activity and a molecular weight of about 55–65 kDa as judged by SDS PAGE-electrophoresis, an isolated DNA sequence comprising an open reading frame capable of encoding a protein according to the invention, preferably characterised in that it comprises an open reading frame which is capable of encoding a protein depicted in SEQ ID NO. 16, SEQ ID NO. 57, SEQ ID NO. 70, SEQ ID NO. 72 or SEQ ID NO. 74 or mutants thereof, and DNA capable of hybridising therewith under stringent conditions. The invention further comprises plants incorporating chimeric DNA capable of encoding a protein according to the invention, and wherein the protein is expressed. Also shown is the carbohydrate and preferably hexose oxidating activity of said protein. Also methods are provided for combating fungi, especially <i>Phytophthora</i> and <i>Pythium</i> species, using a protein or a host cell capable of producing the protein.</p>		
<p>FR/fungal/resistance/hexose/oxidase/Ms59/WL64/Phytophthora/Pythium/</p> <p>DO NOT REMOVE FROM LIBRARY</p>		

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ANTIFUNGAL PROTEINS, DNA CODING THEREFORE, AND HOSTS
INCORPORATING SAME

FIELD OF THE INVENTION

5 The present invention relates to new oxidases, which can act as antifungal proteins, DNA coding therefor and hosts incorporating the DNA, as well as methods of combating fungal pathogens by causing said fungal pathogens to be contacted with said protein or proteins. The invention further relates to plants, incorporating and expressing
10 DNA coding for antifungal proteins, and to plants which as a result thereof show reduced susceptibility to fungal pathogens.

BACKGROUND ART

Fungal diseases of crop plants have been one of the principal
15 causes of crop losses throughout the history of crop cultivation. The growing of crops as monocultures encourages the proliferation of virulent races of fungal pathogens and wherever a new variety of crop plant becomes grown on a wide scale of the risks of a virulent strain of a pathogen evolving to attack that crop increase drastically. The
20 occurrence of disease is significantly worsened by the international transport of pathogen-carrying plant materials, which can bring together plants with pathogens against which they have had no opportunity to evolve resistance. Thus by man's intervention the natural balance between host and pathogen has been disturbed with
25 disastrous effect on a number of occasions. Catastrophic losses and even famines such as occurred in Ireland during the 19th century, caused by the potato blight fungus (*Phytophthora infestans*) have resulted from such activities. Fungal disease can also make it completely impossible to grow certain crops in large areas, as was the
30 case when *Fusarium* wilt wiped out tomato growing in large areas of the Eastern USA or the downy mildew (*Plasmopara viticola*) fungus devastated vine growing in parts of Europe. Outbreaks of fungal disease can also have a severe effect on the environment as happened when almost the entire English Elm (*Ulmus procera*) population was
35 destroyed by Dutch Elm Disease (*Ceratocystis ulmi*). In addition the losses which may be caused during the growing of crops fungal disease may contribute to further post harvest losses. Various soft rots such as *Botrytis cinerea* are particularly problematic in soft fruit, for

Phytophthora infestans belongs to the group of fungi referred to as Oomycetes. *Phytophthora infestans* infects various members of Solanaceae, such as potato, tomato and some ornamentals. It causes late blight of potatoes and tomatoes affecting all parts except roots. Geographically, the fungus is widely distributed, and it can be found in all potato-producing countries. Economically late blight in potatoes is of major importance, as infection early in the season can severely reduce crop yield. Currently the disease is controlled by spraying chemical fungicides (dithiocarbamates, such as mancozeb, manec and zineb) regularly. Both from an environmental and economical point of view, biological control of diseases caused by *Phytophthora infestans* could have advantages over the use of chemical fungicides.

Pythium also belongs to the group of fungi referred to as Oomycetes. The genus *Pythium* differs from the related genus *Phytophthora* by forming relatively undifferentiated sporangia. Geographically, this fungus is widely distributed on all continents. The first main type of disease caused by *Pythium* species is damping-off, due to sudden and fast developing attacks on young seedlings in the field or in nurseries. *Pythium* species cause a second type of disease which is root necrosis and causes a general slowing of plant growth (for example wheat and maize) and loss of yield. The main losses caused by *Pythium* in Europe are to field crops such as sugarbeet. In principle, losses tend to be all-or-nothing. Similarly, nursery sowings of ornamentals and forest trees may be completely destroyed. (For a review on Oomycetes, vide: European Handbook of Plant Diseases, ed. by I.M. Smith et al., 1988, Blackwell Scientific Publications, Ch.8)

Another fungus is *Botrytis*, especially *B. cinerea*, belonging to the group of Fungi Imperfecti, which causes gray mold blight or bud and flower blight, which is common on soft ripe fruits after harvesting, but it can also occur before harvest. It can also affect various vegetables such as lettuce, beans and tomato. Other species of *Botrytis* are common on flowers, such as lilies, gladiolus and tulips.

A protein with antifungal activity, isolated from TMV-induced tobacco leaves, which is capable of causing lysis of germinating spores and hyphal tips of *Phytophthora infestans* and which causes the hyphae to grow at a reduced rate, was disclosed in WO91/18984 A1. This protein has an apparent molecular weight of about 24 kDa and was named

AP24. Comparison of its complete amino acid sequence, as deduced from the nucleic acid sequence of the AP24 gene, with proteins known from databases revealed that the protein was an osmotin-like protein.

Despite initial success in combating fungal pathogens, such as *Phytophthora infestans*, and the genetic engineering of plants capable of producing these antifungal proteins with activity against this fungal pathogen there remains a need to identify and isolate other proteins with antifungal activity against this fungus.

10

SUMMARY OF THE INVENTION

The present invention provides an isolated protein obtainable from a plant source which has antifungal activity, most effectively directed to Oomycetes, and preferably to *Phytophthora* and/or *Pythium* and a molecular weight of about 55-65 kDa as judged by SDS PAGE-electrophoresis. Preferred proteins are those that are obtainable from sunflower or lettuce plants. Even more preferred proteins are obtainable from sunflower or lettuce leaves induced with sodium salicylate. A still more preferred isolated protein is characterised in that it is selected from the group of proteins having the amino acid sequence selected from the group comprising of the amino acid sequences depicted in SEQ ID NO's: 1, 2 or 6, 16, 20, 49, 50, 51, 58, 71, 73 or 75 as well as muteins thereof which have antifungal and especially anti-*Phytophthora* and/or anti-*Pythium* activity. A still further preferred protein according to the invention is one characterised in that it comprises a protein that comprises the amino acid sequence as represented by SEQ ID NO's: 16, 20, 58, 71, 73 or 75 or by a part of said sequence like represented in SEQ ID NO: 6.

The invention also provides a new enzyme, the enzymatic activity being oxidation of carbohydrates.

The invention also embraces an isolated DNA sequence comprising an open reading frame capable of encoding a protein according to the invention, preferably characterised in that the open reading frame is capable of encoding a protein according to the invention, and DNA capable of hybridising therewith under stringent conditions.

The invention also provides a chimeric DNA sequence according to the invention further comprising a transcriptional initiation region and, optionally, a transcriptional termination region, so linked to said open reading frame as to enable the DNA to be transcribed in a

living host cell when present therein, thereby producing RNA which comprises said open reading frame. A preferred chimeric DNA sequence according to the invention is one, wherein the RNA comprising said open reading frame is capable of being translated into protein in said
5 host cell, when present therein, thereby producing said protein. Especially preferred are DNA sequences comprising a sequence as depicted in SEQ ID NO's: 15, 19, 57, 70, 72 or 74.

The invention also embraces a chimeric DNA sequence comprising a DNA sequence according to the invention, which may be selected from
10 replicons, such as bacterial cloning plasmids and vectors, such as a bacterial expression vector, a (non-integrative) plant viral vector, a Ti-plasmid vector of *Agrobacterium*, such as a binary vector, and the like, as well as a host cell comprising a replicon or vector according to the invention, and which is capable of maintaining said replicon
15 once present therein. Preferred according to that embodiment is a host cell which is a plant cell, said vector being a non-integrative viral vector.

The invention further provides a host cell stably incorporating in its genome a chimeric DNA sequence according to the invention, such
20 as a plant cell, as well as multicellular hosts comprising such cells, or essentially consisting of such cells, such as plants. Especially preferred are plants characterised in that the chimeric DNA according to the invention is expressed in at least a number of the plant's cells causing the said antifungal protein to be produced therein.

25 According to yet another embodiment of the invention a method for producing a protein with carbohydrate oxidase activity is provided, characterised in that a host cell according to the invention is grown under conditions allowing the said protein to be produced by said host cell, optionally followed by the step of recovering the
30 protein from the host cells.

Another part of the invention is directed to the antifungal use of a protein which has carbohydrate oxidase activity.

The invention provides also for the use of a protein according to the invention for retarding the growth of fungi, preferably
35 Oomycetes and more preferably *Phytophthora* and *Pythium*. According to yet another embodiment, retarding the growth of the fungi is on or in the neighbourhood of the plant by applying a microorganism capable of producing the protein or by harvesting the protein from a microbial

host and applying the protein in an agrochemical formulation.

The invention also provides a method for obtaining plants with reduced susceptibility to fungi, especially *Phytophthora* and/or *Pythium*, comprising the steps of

- 5 (a) introducing into ancestor cells which are susceptible of regeneration into a whole plant,
 - a chimeric DNA sequence comprising an open reading frame capable of encoding a protein according to claim 1, said open reading frame being operatively linked to a transcriptional and translational region
 - 10 and, optionally, a transcriptional termination region, allowing the said protein to be produced in a plant cell that is susceptible to infection by said fungus, and
 - a chimeric DNA sequence capable of encoding a plant selectable marker allowing selection of transformed ancestor cells when said
 - 15 selectable marker is present therein, and
- (b) regenerating said ancestor cells into plants under conditions favouring ancestor cells which have the said selectable marker, and
- (c) identifying a plant which produces a protein according to claim 1, thereby reducing the susceptibility of said plant to infection by said
- 20 fungus.

Preferred according to the invention is a method characterised in that step (a) is performed using an *Agrobacterium tumefaciens* strain capable of T-DNA transfer to plant cells and which harbours the said chimeric DNA cloned into binary vector pMOG800; another preferred

25 method is when step (b) is performed in the presence of an antibiotic favouring cells which have a neomycin phosphotransferase.

The invention further provides an antifungal composition comprising a protein according to the invention and a suitable carrier.

- 30 An antibody, capable of reacting with an N-terminal fragment of a protein according to the invention, preferably to the peptide represented by SEQ ID NO's: 6, 16, 20, 58, 71, 73 or 75 is also provided. The antibody is suitably used to detect expression levels of chimeric DNA according to the invention in host cells and
- 35 multicellular hosts, preferably plants, capable of producing a protein according to the invention.

The invention also provides a nucleic acid sequence obtainable from a gene encoding a protein according to the invention, said nucleic acid

sequence having tissue-specific transcriptional regulatory activity in a plant. The invention specifically provides a nucleic acid sequence obtainable from the region upstream of the translational initiation site of said gene, preferably at least 500 nucleotides immediately upstream of the translational initiation site of said gene.

DESCRIPTION OF THE FIGURES

Figure 1: SDS-PAGE (12.5%) of the different purification steps of MS59 sunflower protein. Mw= molecular weight markers; 1= crude sunflower protein extract after gel filtration (G25); 2= protein fraction bound to cation exchange chromatography (S-sepharose); 3= pool of active fractions after cation exchange chromatography (Mono S); 4= flow through from hydrophobic interaction chromatography (phenyl superose); 5= active fractions after gel filtration.

Figure 2: SDS-PAGE (12.5%) of different fractions (number 6 to 16) of the gelfiltration (SD75) column. Fraction 10 to 15 was tested in 3 dilutions for growth inhibition on *Phytophthora infestans* (PANEL A) and on *Pythium ultimum* (PANEL B)

Figure 3: SDS-PAGE (12.5%) of fractions eluted from nine gel slices (lane 1 to 9) of a native PAGE in which a MS59 containing SD75 fraction (SD75 fraction 13) was separated. Right panel: SDS-PAGE (12.5%) with SD75 fraction 13 (L) and two fractions of elution experiment fraction 2 (with MS59) and fraction 5 (with a ~30 kD protein). Bottom panel: growth inhibition of *Phytophthora infestans* tested with elution fraction 1 to 6, with 5 μ l and 1 μ l added per well.

Figure 4: Microscopical analysis of an in vitro fungal inhibition assay 24 hours after addition of *Phytophthora infestans* zoospores to PDA medium. Left panel: control incubation, only MES buffer was added. Right panel: *E. coli*-produced MS59 in MES buffer was added to the incubation.

Figure 5: Microscopical analysis of an *in vitro* fungal inhibition assay 24 hours after addition of *Pythium ultimum* hyphal fragments to PDA medium. Left panel: control incubation, only MES buffer was added. Right panel: *E. coli*-produced MS59 in MES buffer was added to the incubation.

Figure 6: (A). SD 75 gelfiltration profile of WL64. WL64 eluates at fractions 13, 14, 15. Molecular weight markers are indicated above the arrows at the top of the plot. X-axis: fraction number. Y-axis: A280.
10 (B). Coomassie stained 12.5% SDS-PAGE gel of fractions 11-17 of the SD 75 gelfiltration profile. Molecular weight markers are indicated on the right and are in kDa. The protein bands that correlate with antifungal activity are indicated between the arrows.
(C). *In vitro* antifungal assay. Ten microlitres of the respective
15 fractions (500 μ l total) were used to screen the growth inhibition of *Rhizoctonia solani* hyphal fragments.

Figure 7: Coomassie stained 12.5% SDS-PAGE gel of the purification of WL64. Lane 1, lettuce extract; lane 2, HIC peak; lane 3, Source S
20 peak; lane 4, Mono S peak; lane 5, SD 75 peak; lane 6, Mono P peak. Molecular weight markers are indicated on both sides of the figure and are in kDa.

Figure 8: (A). Lineweaver-Burk plot of MS59 (open diamonds), WL64 (closed circles), and GOX (open squares) oxidase activities with
25 glucose as substrate. Amounts of protein per assay were 17, 29, and 45 ng for MS59, WL64 and GOX respectively.

(B). Lineweaver-Burk plot of MS59 (open diamonds), WL64 (closed circles), and GOX (open squares) oxidase activities with fungal cell
30 walls as substrate. Amounts of protein per assay were 17, 29, and 225 ng for MS59, WL64 and GOX respectively.

Figure 9: Substrate specificity for the oxidase activities of MS59 (dotted bars), WL64 (diagonal striped bars), and GOX (filled bars).

Figure 10: Alignment of the proteins of the invention MS59, WL64 and the two homologues from *A. thaliana* At26 (SEQ ID NO: 71) and At27 (SEQ ID NO: 75) (with the known berberine bridge enzymes (EcBBE and PsBBE). Conserved changes are denoted in gray, while areas of identity (3 of the 6 amino acids identical) are given in black.

DETAILED DESCRIPTION OF THE INVENTION

The antifungal effect of the protein(s) of the invention has been demonstrated in *in vitro* assays for the following fungi;

10 *Phytophthora infestans*, *Phytophthora cactorum*, *Phytophthora nicotiana*, *Phytophthora megasperma*, *Pythium ultimum*, *Pythium sylvaticum*, *Pythium violae*, *Pythium paroecandrum*, *Rhizoctonia solani*, *Tanatephorus cucumeris*, *Helicobasidium purpureum*, *Sclerotium cepivorum*, *Pichia pastoris* and *Botrytis cinerea* for purposes of illustration. It will be

15 clear, that the use of the protein(s) of the invention, or DNA encoding therefore, for use in a process of combating fungi is not limited to the mentioned fungi. There is no reason to assume that the protein(s) according to the invention do not possess antifungal activity against a far broader range of fungi than those tested here,

20 especially in the class of Oomycetes.

Although the invention is illustrated in detail for transgenic tomato, tobacco, carrot, potato and *Brassica napus* plants, it should be understood that any plant species that is subject to some form of

25 fungal attack, especially from the fungi mentioned above, may be provided with one or more plant expressible gene constructs, which when expressed overproduce the protein(s) of the invention in said plant in order to decrease the rate of infectivity and/or the effects of such attack. The invention can even be practiced in plant species

30 that are presently not amenable for transformation, as the amenability of such species is just a matter of time and because transformation as such is of no relevance for the principles underlying the invention. Hence, plants for the purpose of this description shall include angiosperms as well as gymnosperms, monocotyledonous as well as

35 dicotyledonous plants, be they for feed, food or industrial processing purposes; included are plants used for any agricultural or horticultural purpose including forestry and flower culture, as well as home gardening or indoor gardening, or other decorative purposes.

The protein according to the present invention may be obtained by isolating it from any suitable plant source material containing it. A particularly suitable source comprises leaves of the sunflower (*Helianthus*) and leaves of lettuce (*Lactuca sativa* cv. Lollo bionda).

5 The presence of antifungal proteins according to the invention in plant source material can readily be determined for any plant species by making plant extracts from those species and testing those extracts for the presence of antifungal activity using *in vitro* antifungal assays as described herein, further fractionating the obtained samples
10 by any suitable protein fractionation technique in conjunction with the *in vitro* assay until an antifungal fraction is obtained which comprises an approximately 55-65 kDa protein, internally denoted as MS59 or its homologue WL64, which in isolated form shows antifungal activity. Especially, fractions may be tested for antifungal activity
15 on Oomycetes, for example, *Phytophthora* or *Pythium ultimum* and the like, or other fungi, such as the Basidiomycetes, Ascomycetes, Zygomycetes or other classes or subclasses.

Alternatively, antifungal proteins according to the invention may be obtained by cloning DNA comprising an open reading frame
20 capable of encoding said protein, or the precursor thereof, linking said open reading frame to a transcriptional, and optionally a translational initiation and transcriptional termination region, inserting said DNA into a suitable host cell and allowing said host cell to produce said protein. Subsequently, the protein may be
25 recovered from said host cells, preferably after secretion of the protein into the culture medium by said host cells. Alternatively, said host cells may be used directly in a process of combating fungal pathogens according to the invention as a pesticidal acceptable composition.

30 Host cells suitable for use in a process of obtaining a protein according to the invention may be selected from prokaryotic microbial hosts, such as bacteria e.g. *Agrobacterium*, *Bacillus*, *Cyanobacteria*, *E.coli*, *Pseudomonas*, and the like, as well as eukaryotic hosts including yeasts, e.g. *Saccharomyces cerevisiae*, fungi, e.g.
35 *Trichoderma* and plant cells, including protoplasts.

In a method of retarding the growth of the fungi on or in the neighbourhood of the plant leaves, host cells may suitably be selected from any species routinely used as biological fungicides.

Also the proteins can be produced by microorganisms, harvested and applied in a agrochemical formulation.

The word protein means a sequence of amino acids connected through peptide bonds. Polypeptides or peptides are also considered to be proteins. Muteins of the protein of the invention are proteins that are obtained from the proteins depicted in the sequence listing by replacing, adding and/or deleting one or more amino acids, while still retaining their antifungal activity. Such muteins can readily be made by protein engineering in vivo, e.g. by changing the open reading frame capable of encoding the antifungal protein such that the amino acid sequence is thereby affected. As long as the changes in the amino acid sequences do not altogether abolish the antifungal activity such muteins are embraced in the present invention.

The present invention provides a chimeric DNA sequence which comprises an open reading frame capable of encoding a protein according to the invention. The expression chimeric DNA sequence shall mean to comprise any DNA sequence which comprises DNA sequences not naturally found in nature. For instance, chimeric DNA shall mean to comprise DNA comprising the said open reading frame in a non-natural location of the plant genome, notwithstanding the fact that said plant genome normally contains a copy of the said open reading frame in its natural chromosomal location. Similarly, the said open reading frame may be incorporated in the plant genome wherein it is not naturally found, or in a replicon or vector where it is not naturally found, such as a bacterial plasmid or a viral vector. Chimeric DNA shall not be limited to DNA molecules which are replicable in a host, but shall also mean to comprise DNA capable of being ligated into a replicon, for instance by virtue of specific adaptor sequences, physically linked to the open reading frame according to the invention. The open reading frame may or may not be linked to its natural upstream and downstream regulatory elements.

The open reading frame may be derived from a genomic library. In this latter it may contain one or more introns separating the exons making up the open reading frame that encodes a protein according to the invention. The open reading frame may also be encoded by one uninterrupted exon, or by a cDNA to the mRNA encoding a protein according to the invention. Open reading frames according to the invention also comprise those in which one or more introns have been

artificially removed or added. Each of these variants is embraced by the present invention.

Also part of the invention are chimeric DNA sequences coding for an antifungal protein which comprise one or more of the EST-sequences shown in SEQ ID NO's: 21 to 48. As can be derived from the sequence listings these EST's for which no function was hitherto known share a considerable homology with the DNA sequence coding for the proteins isolated from *Helianthus* and *Lactuca*.

Another part of the invention is formed by the intrinsic activity of the proteins of the invention. They have been found to be carbohydrate oxidases, capable of oxidating a large number of different mono- and di-saccharides. The substrate specificity resembles the specificity of the enzyme hexose oxidase (EC 1.1.3.5), also known as D-hexose: oxygen 1-oxidoreductase. They have also been shown able to oxidise a purified mixture of fungal (*Rhizoctonia*-derived) cell wall components. It is believed that this oxidative capacity confers the antifungal properties to the proteins. In literature there is one example of an antifungal oxidase, the glucose oxidase from the fungus *Aspergillus* (WO 95/14784). The proteins of this invention, however, show a broader substrate spectrum like hexose oxidase and have a lower K_m for the substrate.

From homology searches it has been found that some parts of the amino acid sequence of the proteins of the invention are more conserved and are related to sequences commonly found in oxidases. The highest homology has been found with reticuline oxidase, which enzyme is known from the family of *Papaveraceae* (Facchini, P.J. et al., *Plant Physiol.* 112, 1669-1677, 1996).

In order to be capable of being expressed in a host cell a chimeric DNA according to the invention will usually be provided with regulatory elements enabling it to be recognised by the biochemical machinery of the host and allowing for the open reading frame to be transcribed and/or translated in the host. It will usually comprise a transcriptional initiation region which may be suitably derived from any gene capable of being expressed in the host cell of choice, as well as a translational initiation region for ribosome recognition and attachment. In eukaryotic cells, an expression cassette usually comprises in addition a transcriptional termination region located downstream of said open reading frame, allowing transcription to

terminate and polyadenylation of the primary transcript to occur. In addition, the codon usage may be adapted to accepted codon usage of the host of choice. The principles governing the expression of a chimeric DNA construct in a chosen host cell are commonly understood
5 by those of ordinary skill in the art and the construction of expressible chimeric DNA constructs is now routine for any sort of host cell, be it prokaryotic or eukaryotic.

In order for the open reading frame to be maintained in a host cell it will usually be provided in the form of a replicon comprising
10 said open reading frame according to the invention linked to DNA which is recognised and replicated by the chosen host cell. Accordingly, the selection of the replicon is determined largely by the host cell of choice. Such principles as govern the selection of suitable replicons for a particular chosen host are well within the realm of the ordinary
15 skilled person in the art.

A special type of replicon is one capable of transferring itself, or a part thereof, to another host cell, such as a plant cell, thereby co-transferring the open reading frame according to the invention to said plant cell. Replicons with such capability are
20 herein referred to as vectors. An example of such vector is a Ti-plasmid vector which, when present in a suitable host, such as *Agrobacterium tumefaciens*, is capable of transferring part of itself, the so-called T-region, to a plant cell. Different types of Ti-plasmid vectors (vide: EP 0 116 718 B1) are now routinely being used to
25 transfer chimeric DNA sequences into plant cells, or protoplasts, from which new plants may be generated which stably incorporate said chimeric DNA in their genomes. A particularly preferred form of Ti-plasmid vectors are the so-called binary vectors as claimed in (EP 0
120 516 B1 and US 4,940,838). Other suitable vectors, which may be
30 used to introduce DNA according to the invention into a plant host, may be selected from the viral vectors, e.g. non-integrative plant viral vectors, such as derivable from the double stranded plant viruses (e.g. CaMV) and single stranded viruses, gemini viruses and the like. The use of such vectors may be advantageous, particularly
35 when it is difficult to stably transform the plant host. Such may be the case with woody species, especially trees and vines.

The expression "host cells incorporating a chimeric DNA sequence according to the invention in their genome" shall mean to comprise

cells, as well as multicellular organisms comprising such cells, or essentially consisting of such cells, which stably incorporate said chimeric DNA into their genome thereby maintaining the chimeric DNA, and preferably transmitting a copy of such chimeric DNA to progeny
5 cells, be it through mitosis or meiosis. According to a preferred embodiment of the invention plants are provided, which essentially consist of cells which incorporate one or more copies of said chimeric DNA into their genome, and which are capable of transmitting a copy or copies to their progeny, preferably in a Mendelian fashion. By virtue
10 of the transcription and translation of the chimeric DNA according to the invention in some or all of the plant's cells, those cells that produce the antifungal protein will show enhanced resistance to fungal infections, especially to *Phytophthora* infections. Although the principles as indicated above govern transcription of DNA in plant
15 cells are not always understood, the creation of chimeric DNA capable of being expressed in substantially a constitutive fashion, that is, in substantially most cell types of the plant and substantially without serious temporal and/or developmental restrictions, is now routine. Transcription initiation regions routinely in use for that
20 purpose are promoters obtainable from the cauliflower mosaic virus, notably the 35S RNA and 19S RNA transcript promoters and the so-called T-DNA promoters of *Agrobacterium tumefaciens*, in particular to be mentioned are the nopaline synthase promoter, octopine synthase promoter (as disclosed in EP 0 122 791 B1) and the mannopine synthase
25 promoter. In addition plant promoters may be used, which may be substantially constitutive, such as the rice actin gene promoter, or e.g. organ-specific, such as the root-specific promoter.

Alternatively, pathogen-inducible promoters may be used such as the
PRP1 promoter (also named *gst1* promoter) obtainable from potato
30 (Martini N. et al. (1993), Mol. Gen. Genet. 263, 179-186). The choice of the promoter is not essential, although it must be said that constitutive high-level promoters are slightly preferred. It is further known that duplication of certain elements, so-called enhancers, may considerably enhance the expression level of the DNA
35 under its regime (vide for instance: Kay R. et al. (1987), Science 236, 1299-1302: the duplication of the sequence between -343 and -90 of the CaMV 35S promoter increases the activity of that promoter). In addition to the 35S promoter, singly or doubly enhanced, examples of

high-level promoters are the light-inducible ribulose biphosphate carboxylase small subunit (*rbcSSU*) promoter and the chlorophyll a/b binding protein (*Cab*) promoter. Also envisaged by the present invention are hybrid promoters, which comprise elements of different promoter regions physically linked. A well known example thereof is the so-called CaMV enhanced mannopine synthase promoter (US Patent 5,106,739), which comprises elements of the mannopine synthase promoter linked to the CaMV enhancer.

As regards the necessity of a transcriptional terminator region, it is generally believed that such a region enhances the reliability as well as the efficiency of transcription in plant cells. Use thereof is therefore strongly preferred in the context of the present invention.

As regards the applicability of the invention in different plant species, it has to be mentioned that one particular embodiment of the invention is merely illustrated with transgenic tomato and tobacco plants as an example, the actual applicability being in fact not limited to these plant species. Any plant species that is subject to some form of fungal attack, in particular by Oomycetes such as *Phytophthora infestans*, may be treated with proteins according to the invention, or preferably, be provided with a chimeric DNA sequence according to the invention, allowing the protein to be produced in some or all of the plant's cells.

Although some of the embodiments of the invention may not be practicable at present, e.g. because some plant species are as yet recalcitrant to genetic transformation, the practicing of the invention in such plant species is merely a matter of time and not a matter of principle, because the amenability to genetic transformation as such is of no relevance to the underlying embodiment of the invention.

Transformation of plant species is now routine for an impressive number of plant species, including both the *Dicotyledoneae* as well as the *Monocotyledoneae*. In principle any transformation method may be used to introduce chimeric DNA according to the invention into a suitable ancestor cell, as long as the cells are capable of being regenerated into whole plants. Methods may suitably be selected from the calcium/polyethylene glycol method for protoplasts (Krens, F.A. et al., 1982, *Nature* 296, 72-74; Negrutiu I. et al, June 1987, *Plant Mol.*

Biol. 8, 363-373), electroporation of protoplasts (Shillito R.D. et al., 1985 Bio/Technol. 3, 1099-1102), microinjection into plant material (Crossway A. et al., 1986, Mol. Gen. Genet. 202, 179-185), (DNA or RNA-coated) particle bombardment of various plant material
5 (Klein T.M. et al., 1987, Nature 327, 70), infection with (non-integrative) viruses and the like. A preferred method according to the invention comprises *Agrobacterium*-mediated DNA transfer. Especially preferred is the use of the so-called binary vector technology as disclosed in EP A 120 516 and U.S. Patent 4,940,838).

10 Tomato transformation is preferably done essentially as described by Van Roekel et al. (Van Roekel, J.S.C., Damm, B., Melchers, L.S., Hoekema, A. (1993). Factors influencing transformation frequency of tomato (*Lycopersicon esculentum*). Plant Cell Reports, 12, 644-647). Potato transformation is preferably done essentially as described by
15 Hoekema et al. (Hoekema, A., Huisman, M.J., Molendijk, L., van den Elzen, P.J.M., and Cornelissen, B.J.C. (1989). The genetic engineering of two commercial potato cultivars for resistance to potato virus X. Bio/Technology 7, 273-278).

Generally, after transformation plant cells or cell groupings are
20 selected for the presence of one or more markers which are encoded by plant expressible genes co-transferred with the nucleic acid sequence encoding the protein according to the invention, whereafter the transformed material is regenerated into a whole plant.

Although considered somewhat more recalcitrant towards genetic
25 transformation, monocotyledonous plants are amenable to transformation and fertile transgenic plants can be regenerated from transformed cells or embryos, or other plant material. Presently, preferred
methods for transformation of monocots are microprojectile bombardment
of embryos, explants or suspension cells, and direct DNA uptake or
30 electroporation (Shimamoto, et al, 1989, Nature 338, 274-276).

Transgenic maize plants have been obtained by introducing the *Streptomyces hygrosopicus* bar-gene, which encodes phosphinothricin acetyltransferase (an enzyme which inactivates the herbicide phosphinothricin), into embryogenic cells of a maize suspension
35 culture by microprojectile bombardment (Gordon-Kamm, 1990, Plant Cell, 2, 603-618). The introduction of genetic material into aleurone protoplasts of other monocot crops such as wheat and barley has been reported (Lee, 1989, Plant Mol. Biol. 13, 21-30). Wheat plants have

been regenerated from embryogenic suspension culture by selecting only the aged compact and nodular embryogenic callus tissues for the establishment of the embryogenic suspension cultures (Vasil, 1990 Bio/Technol. 8, 429-434). The combination with transformation systems
5 for these crops enables the application of the present invention to monocots.

Monocotyledonous plants, including commercially important crops such as rice and corn are also amenable to DNA transfer by *Agrobacterium* strains (vide WO 94/00977; EP 0 159 418 B1; Gould J,
10 Michael D, Hasegawa O, Ulian EC, Peterson G, Smith RH, (1991) Plant. Physiol. 95, 426-434).

Following DNA transfer and regeneration, putatively transformed plants may be evaluated, for instance using Southern analysis, for the presence of the chimeric DNA according to the invention, copy number
15 and/or genomic organization. In addition, or alternatively, expression levels of the newly introduced DNA may be undertaken, using Northern and/or Western analysis, techniques well known to persons having ordinary skill in the art. After the initial analysis, which is optional, transformed plants showing the desired copy number and
20 expression level of the newly introduced chimeric DNA according to the invention may be tested for resistance levels against a pathogen susceptible to the protein according to the invention, such as *Phytophthora infestans*. Alternatively, the selected plants may be subjected to another round of transformation, for instance to
25 introduce further genes, such as genes encoding chitinases, glucanases, osmotins, magainins or the like, in order to enhance resistance levels, or broaden the resistance to other fungi found not to be susceptible to the protein according to the invention in an in vitro assay as described herein.

30 Other evaluations may include the testing of fungal resistance under field conditions, checking fertility, yield, and other characteristics. Such testing is now routinely performed by persons having ordinary skill in the art.

Following such evaluations, the transformed plants may be grown
35 directly, but usually they may be used as parental lines in the breeding of new varieties or in the creation of hybrids and the like.

Many plant proteins exhibit antifungal effects, some however do not do so as such, but yield a significant synergistic antifungal

effect if used in combination with other plant proteins. In European Patent Application 440 304 A1 it was disclosed that simultaneous relative over-expression of a plant expressible glucanase gene in conjunction with a basic chitinase from tobacco in transgenic plants results in a higher level of resistance to fungi than in plants expressing a plant expressible class-I chitinase alone.

Both chitinases, glucanases, osmotins, magainins and the new antifungal protein according to the invention accumulate in infected plant tissues upon an incompatible pathogen-plant interaction. From this observation and the fact that several proteins are found to synergise each others antifungal effects, we envision, that the antifungal protein according to the invention may be suitably used in conjunction with other proteins that are associated with pathogen resistance.

Examples of proteins that may be used in combination with the proteins according to the invention include, but are not limited to, β -1,3-glucanases and chitinases which are obtainable from barley (Swegle M. et al., 1989, Plant Mol. Biol. 12, 403-412; Balance G.M. et al., 1976, Can. J. Plant Sci. 56, 459-466 ; Hoj P.B. et al., 1988, FEBS Lett. 230, 67-71; Hoj P.B. et al., 1989, Plant Mol. Biol. 13, 31-42 1989), bean (Boller T. et al., 1983, Planta 157, 22-31; Broglie K.E. et al. 1986, Proc. Natl. Acad. Sci. USA 83, 6820-6824; Vögeli U. et al., 1988 Planta 174, 364-372); Mauch F. & Staehelin L.A., 1989, Plant Cell 1, 447-457); cucumber (Motraux J.P. & Boller T. (1986), Physiol. Mol. Plant Pathol. 28, 161-169); leek (Spanu P. et al., 1989, Planta 177, 447-455); maize (Nasser W. et al., 1988, Plant Mol. Biol. 11, 529-538), oat (Fink W. et al., 1988, Plant Physiol. 88, 270-275), pea (Mauch F. et al. 1984, Plant Physiol. 76, 607-611; Mauch F. et al., 1988, Plant Physiol. 87, 325-333), poplar (Parsons, T.J. et al, 1989, Proc. Natl. Acad. Sci. USA 86, 7895-7899), potato (Gaynor J.J. 1988, Nucl. Acids Res. 16, 5210; Kombrink E. et al. 1988, Proc. Natl. Acad. Sci. USA 85, 782-786; Laflamme D. and Roxby R., 1989, Plant Mol. Biol. 13, 249-250), tobacco (e.g. Legrand M. et al. 1987, Proc. Natl. Acad. Sci. USA 84, 6750-6754; Shinshi H. et al. 1987, Proc. Natl. Acad. Sci. USA 84, 89-93), tomato (Joosten M.H.A. & De Wit P.J.G.M. 1989, Plant Physiol. 89, 945-951), wheat (Molano J. et al., 1979, J. Biol. Chem. 254, 4901-4907), and the like.

To obtain transgenic plants capable of constitutively expressing more than one chimeric gene, a number of alternatives are available including the following:

- A. The use of DNA, e.g. a T-DNA on a binary plasmid, with a number of modified genes physically coupled to a selectable marker gene. The advantage of this method is that the chimeric genes are physically coupled and therefore migrate as a single Mendelian locus.
- B. Cross-pollination of transgenic plants each already capable of expressing one or more chimeric genes, preferably coupled to a selectable marker gene, with pollen from a transgenic plant which contains one or more chimeric genes coupled to another selectable marker. Afterwards the seed, which is obtained by this crossing, maybe selected on the basis of the presence of the two selectable markers, or on the basis of the presence of the chimeric genes themselves. The plants obtained from the selected seeds can afterwards be used for further crossing. In principle the chimeric genes are not on a single locus and the genes may therefore segregate as independent loci.
- C. The use of a number of a plurality chimeric DNA molecules, e.g. plasmids, each having one or more chimeric genes and a selectable marker. If the frequency of co-transformation is high, then selection on the basis of only one marker is sufficient. In other cases, the selection on the basis of more than one marker is preferred.
- D. Consecutive transformation of transgenic plants already containing a first, second, (etc), chimeric gene with new chimeric DNA, optionally comprising a selectable marker gene. As in method B, the chimeric genes are in principle not on a single locus and the chimeric genes may therefore segregate as independent loci.
- E. Combinations of the above mentioned strategies.

The actual strategy may depend on several considerations as maybe easily determined such as the purpose of the parental lines (direct growing, use in a breeding programme, use to produce hybrids) but is not critical with respect to the described invention.

In this context it should be emphasised that plants already containing chimeric DNA capable of encoding antifungal proteins may form a suitable genetic background for introducing chimeric DNA according to the invention, for instance in order to enhance resistance levels, or broaden the resistance. The cloning of other genes corresponding to proteins that can suitably be used in

combination with DNA, and the obtention of transgenic plants, capable of relatively over-expressing same, as well as the assessment of their effect on pathogen resistance in *planta*, is now within the scope of the ordinary skilled person in the art.

5 The obtention of transgenic plants capable of expressing, or relatively over-expressing, proteins according to the invention is a preferred method for counteracting the damages caused by fungi, such as Oomycetes like *Phytophthora infestans*, as will be clear from the above description. However, the invention is not limited thereto. The
10 invention clearly envisions also the use of the proteins according to the invention as such, preferably in the form of a fungicidal composition. Fungicidal composition include those in which the protein is formulated as such, but also in the form of host cells, such as bacterial cells, capable of producing the protein thereby causing the
15 pathogen to be contacted with the protein. Suitable host cells may for instance be selected from harmless bacteria and fungi, preferably those that are capable of colonising roots and/or leaves of plants. Example of bacterial hosts that may be used in a method according to the invention are strains of *Agrobacterium*, *Arthrobacter*,
20 *Azospyrillum*, *Pseudomonas*, *Rhizobacterium*, and the like, optionally after having been made suitable for that purpose.

 Compositions containing antifungal proteins according to the invention may comprise in addition thereto, osmotin-like proteins as defined in WO91/18984. Independently, the invention provides
25 antifungal compositions which further comprise inhibitory agents such as classical fungal antibiotics, SAFPs and chemical fungicides such as polyoxines, nikkomycines, carboxymides, aromatic carbohydrates, carboxines, morpholines, inhibitors of sterol biosynthesis,
30 organophosphorus compounds, enzymes such as glucanases, chitinases, lysozymes and the like. Either *per se*, or in combination with other active constituents, the antifungal protein of the invention should be applied in concentrations between 1 ng/ml and 1 mg/ml, preferably between 2 ng/ml and 0.1 mg/ml, within pH boundaries of 3.0 and 9.0. In general it is desired to use buffered preparations, e.g. phosphate
35 buffers between 1mM and 1M, preferably between 10 mM and 100mM, in particular between 15 and 50 mM, whereby in case of low buffer concentrations it is desired to add a salt to increase ionic strength, preferably NaCl in concentrations between 1 mM and 1M, preferably 10

mM and 100 mM.

Plants, or parts thereof, which relatively over-express a protein according to the invention, including plant varieties, with improved resistance against fungal diseases, especially diseases caused by Oomycetes like *Phytophthora* and *Pythium* may be grown in the field, in the greenhouse, or at home or elsewhere. Plants or edible parts thereof may be used for animal feed or human consumption, or may be processed for food, feed or other purposes in any form of agriculture or industry. Agriculture shall mean to include horticulture, arboriculture, flower culture, and the like. Industries which may benefit from plant material according to the invention include but are not limited to the pharmaceutical industry, the paper and pulp manufacturing industry, sugar manufacturing industry, feed and food industry, enzyme manufacturers and the like.

The advantages of the plants, or parts thereof, according to the invention are the decreased need for fungicide treatment, thus lowering costs of material, labour, and environmental pollution, or prolonging shelf-life of products (e.g. fruit, seed, and the like) of such plants. Plants for the purpose of this invention shall mean multicellular organisms capable of photosynthesis, and subject to some form of fungal disease. They shall at least include angiosperms as well as gymnosperms, monocotyledonous as well as dicotyledonous plants.

The phrase "plants which relatively over-express a protein" shall mean plants which contain cells expressing a transgene-encoded protein which is either not naturally present in said plant, or if it is present by virtue of an endogenous gene encoding an identical protein, not in the same quantity, or not in the same cells, compartments of cells, tissues or organs of the plant. It is known for instance that proteins which normally accumulate intracellularly may be targeted to the apoplastic space.

According to another aspect of the invention the regulatory region of a plant gene coding for the antifungal protein of the invention may be used to express other heterologous sequences under the control thereof. The use of a regulatory element of at least 1000 bp directly upstream of the gene coding region is sufficient for obtaining expression of any heterologous sequence.

Heterologous sequences in this respect means gene regions not naturally associated to said regulatory region, and they comprise both different gene coding regions, as well as antisense gene regions. Heterologous coding sequences that may be advantageously expressed in the vascular tissue comprise those coding for antipathogenic proteins, e.g. insecticidal, bactericidal, fungicidal, and nematocidal proteins. In such a strategy it may prove exceptionally advantageous to select a protein with activity against a pathogen or pest which has a preference for phloem as source of nutrients (e.g. aphids), or as entrance to invade the plant. Examples are extensin, lectin, or lipoxidase against aphids (See WO93/04177). Assuming that the regulatory region according to the invention is active in xylem, antifungal proteins may be expressed under the control of said regulatory region to combat *Fusarium*, *Verticillium*, *Alternaria* and *Ceratocystus* species.

The use of the regulatory region according to the invention may also be used advantageously to regulate or control phloem transport processes. Numerous other applications will readily occur to those of skill in the art.

The expression of part of (part of) an endogenous gene in the antisense orientation (such as disclosed in EP 0 233 399 A), can effectively down-regulate expression of said endogenous gene, with interesting applications. Moreover, the gene encoding the antifungal protein according to the invention itself may be down-regulated using the antisense approach which may help establishing the nature and function of the protein. The regions responsible for tissue-specific expression may be unravelled further using the GUS-marker in a way analogous to the way illustrated herein.

The following state of the art may be taken into consideration, especially as illustrating the general level of skill in the art to which this invention pertains.

EP-A 392 225 A2; EP-A 440 304 A1; EP-A 460 753 A2; WO90/07001 A1; US Patent 4,940,840.

Yet another part of the invention is directed at the production of a novel oxidative enzyme, capable of oxidising carbohydrates even at low concentrations due to its low K_m . Most specifically hexoses are the substrate of the enzymatic activity although also other sugars are

affected to some lesser extent. The nzymes can be isolated from the sources in which they naturally occur (according to the method described in this invention) or they can be isolated from plants or other organisms transformed with an expressible gene encoding the protein. These oxidases can be used in industrial processes for the oxidation of carbohydrates, such as glucose, mannose, galactose, cellobiose, maltose and lactose.

Evaluation of transgenic plants

Subsequently transformed plants are evaluated for the presence of the desired properties and/or the extent to which the desired properties are expressed. A first evaluation may include the level of expression of the newly introduced genes, the level of fungal resistance of the transformed plants, stable heritability of the desired properties, field trials and the like.

Secondly, if desirable, the transformed plants can be crossbred with other varieties, for instance varieties of higher commercial value or varieties in which other desired characteristics have already been introduced, or used for the creation of hybrid seeds, or be subject to another round of transformation and the like.

Synergy

The combination of one of the antifungal protein according to the instant invention and other antifungal proteins of plant or microbial source are predicted to show a drastic synergistic antifungal effect. Similar synergistic antifungal effects were shown if combinations of antifungal CBPs or Chi-V are combined with either β -1,3-glucanases or chitinases from other plant origins. Apparently, the synergizing effect of combinations of pathogen induced proteins is a more general phenomenon that has important consequences for the engineering of fungal resistant plants.

Plants, or parts thereof of commercial interest, with improved resistance against phytopathogenic fungi can be grown in the field or in greenhouses, and subsequently be used for animal feed, direct consumption by humans, for prolonged storage, used in food- or other industrial processing, and the like. The advantages of the plants, or parts thereof, according to the invention are the decreased need for

fungicide treatment, thus lowering costs of material, labour, and environmental pollution, or prolonged shelf-life of products (e.g. fruit, seed, and the like) of such plants.

5

EXPERIMENTAL PART

Standard methods for the isolation, manipulation and amplification of DNA, as well as suitable vectors for replication of recombinant DNA, suitable bacterium strains, selection markers, media and the like are described for instance in Maniatis et al., molecular cloning: A Laboratory Manual 2nd. edition (1989) Cold Spring Harbor Laboratory Press; DNA Cloning: Volumes I and II (D.N. Glover ed. 1985); and in: From Genes To Clones (E.-L. Winnacker ed. 1987).

In vitro antifungal assay

15 All fungi were cultured on potato dextrose agar (Difco) at 25°C, except *Botrytis cinerea* and *Phoma lingam* which were grown on oat meal agar (Difco) at 25°C. *Phytophthora infestans* was grown on rye agar at 18°C in the dark (Caten and Jinks, 1968). *Botrytis cinerea* and *Phoma lingam* were cultivated under UV. Spores of sporulating fungi were
20 harvested by flooding the agar plates with water. The spore concentration was adjusted to 10,000 sp/mL. In the case of *Rhizoctonia solani* and *Pythium ultimum* liquid shake cultures were grown in potato dextrose broth at 25°C. To prepare inoculum from these shake cultures, mycelium was harvested and vortexed for 1 minute. After passage
25 through a fine sieve, inoculum density was adjusted to 2500 - 5000 fragments, of 1 to 3 cells each, per mL.

In case of sporulating fungi, all were tested both with and without pregerminating the spores before application of the protein samples. In case of non-sporulating fungi, hyphal fragments were used.

30 The antifungal activity was monitored during purification in a microtiter plate assay using the fungi *Phytophthora infestans* and *Pythium ultimum* according to Woloshuk et al., 1991 or using other fungi in a similar way. In each well of a 24-well microtiter dish 250 µl potato dextrose agar (PDA) was pipetted. Fungal spores in the case
35 of e.g. *Phytophthora infestans* and hyphal fragments in the case of e.g. *Pythium ultimum* were suspended in water and 400-600 spores or 200 fragments in 50 µl were added to the wells. Subsequently 100 µl filter sterilized (0.22 µm filter) protein solution (in 50 mM MES, pH 6.0)

was added. Microtiter dishes were wrapped with Parafilm and incubated at room temperature. At several timepoints after the initiation of incubation the fungus was monitored microscopically for effects of the added protein. After 2-3 days the mycelium of the growing fungus in the wells was stained with lactophenol cotton blue and the extent of growth was estimated.

GI: growth inhibition; a scale of 0 - 4 is used, 0 = no visible inhibition, 1 = weak inhibition (0 to 30%) inhibition, 2 = moderate (30 to 60%) inhibition, 3 = strong (60 to 90%) inhibition, 4 = very strong (100%) inhibition.

EXAMPLE 1

Purification of an antifungal protein MS59 from sunflower induced with salicylic acid

Leaves of 7 to 8 weeks old sunflower (*Helianthus annuus* cv. zebulon) plants were sprayed daily for 5 times with 10 mM sodium salicylate. After 3 hours the plants were extensively rinsed with water to remove the sodium salicylate. Three days after the final spray, leaves (400 gram) were harvested into liquid nitrogen and homogenized at 4°C in 500 ml 0.5 M NaOAc pH5.2, and 4 gram active carbon, using a Waring blender. The homogenate was filtered over four layers of cheese cloth and subsequently the filtrate was centrifuged for 50 minutes at 20,000 g at 4°C and desalted by passage through a Sephadex G25 column (medium course; Pharmacia), length 60 cm, diameter 11.5 cm, equilibrated in 40 mM NaOAc pH5.2. The desalted protein solution was stored overnight at 4°C and subsequently centrifuged for 45 minutes at 20,000 g at 4°C. The supernatant was passed through a S-sephadex (Fast-flow, Pharmacia) column, length 5 cm, diameter 5 cm, which was equilibrated with 40 mM NaOAc pH 5.2. The column was washed with the above mentioned buffer (flow rate 400 to 500 ml/hr) until the OD₂₈₀ dropped to zero. The bound proteins were eluted using 400 mM NaCl in 200 ml of the above mentioned buffer.

After dialysis against 50 mM MES pH 6.0 the eluate was analyzed for antifungal activity. Antifungal activity was monitored in a microtiter plate assay using the fungus *Phytophthora infestans* and *Pythium ultimum*. See above for details concerning *in vitro* assaying. Subsequently, cationexchange chromatography was reapplied whereby the eluate was passed through an FPLC Mono-S HR 5/5 (Pharmacia) and eluted

with a linear gradient from 0 to 400 mM NaCl. All fractions were analyzed by electrophoresis (Laemmli (1970), Nature 227:680-685) using a 12.5% polyacrylamide gel in the presence of sodium dodecyl sulphate (SDS), using prestained molecular weight markers (15-105 kDa) as reference. Additionally, of all fractions antifungal activity towards *Phytophthora infestans* and *Pythium ultimum* was monitored. Antifungal activity eluted from the column between 45-60 mM NaCl and in all active fractions a 59 kD band was visible. Fractions containing the antifungal activity were pooled and dialysed to 1 M ammonium sulphate in 50 mM potassium phosphate, pH 7. The pool was subjected to hydrophobic interaction chromatography, whereby the sample was applied to an FPLC Phenyl Superose HR 5/5 (Pharmacia) equilibrated in the same buffer and eluted with a linear decreasing gradient from 1 to 0 M ammonium sulphate in 50 mM potassium phosphate, pH 7. As above again all fractions were analyzed on SDS-PAGE and monitored for antifungal activity. Also the pool of proteins not capable of binding to this column (Flow Through, FT) was thus analyzed at the conditions chosen here. Antifungal activity was present most abundantly in the FT and secondly also in the fractions eluting between 0.76 and 0.45 M ammonium sulphate. In both cases a 59 kD protein was visible on SDS-PAGE. FT and the gradient fractions were separately dialysed to 50 mM MES, 0.2 M NaCl and separately chromatographed on a FPLC Superdex 75 HR 10/30 column (Pharmacia) equilibrated to the same buffer. Proteins elute from this column according to their molecular size. In both cases again the presence of a 59 kD protein coincided with antifungal activity towards *Phytophthora infestans* and *Pythium ultimum* as judged from SDS-PAGE and in vitro antifungal assays. The 59 kD protein present in the FT of the hydrophobic interaction column was most abundant and termed MS59 and its purification is visualized in Figure 1. Results of its separation over the gelfiltration column and subsequent analysis both on SDS-PAGE and on *Phytophthora infestans* is shown in Figure 2. Several characteristics (antifungal activity, chromatographical properties, molecular mass) of the gradient protein and MS59 indicate that the two proteins are very similar.

To characterize MS59 further its amino acid sequence was partially determined. Therefore, MS59 was separated in the presence of 0.1 mM thioglycolate in the upper reservoir buffer and SDS on a 12.5% polyacrylamide gel, which was prerun for 2 hours at 50 V with 0.05 mM

glutathione in the upper reservoir buffer. The gel was stained with 5% (w/v) Serva Blue G in 45% (v/v) methanol and 10% acetic acid for 30 minutes and destained in 20% (v/v) acetic acid for 30 minutes and the 59 kDa band was cut out and sequenced using Edman degradation on an Applied Biosystems 477A protein sequencer according to the protocol provided by the manufacturer. N-terminal amino acid sequencing of MS59 revealed that the N-terminus was blocked. To obtain internal sequences, MS59 was digested with trypsin. Trypsin cleaves protein at arginine and lysine residues. The digestion products were separated on a reversed-phase column and analyzed by Edman degradation. Two tryptic fragments were sequenced: Pep1 and Pep2. Of Pep1 25 amino acid residues were identified: S-I-N-V-D-I-E-Q-E-T-A-W-V-Q-A-G-A-T-L-G-E-V-Y-Y-R (SEQ ID NO: 1). The amino acid sequence is given using the one-letter code. Of Pep2 a further 25 amino acid residues were identified: D-P-S-F-P-I-T-G-E-V-Y-T-P-G-(?)-S-S-F-P-T-V-L-Q-N-Y (SEQ ID NO: 2). The amino acid residue between brackets could not be identified unambiguously.

20

EXAMPLE 2

Elution of antifungal protein from native PAGE and subsequent testing

It is obvious from Figures 1 and 2 that MS59 is not completely pure. To further ensure that indeed the 59 kDa protein is responsible for the observed antifungal activity, the fraction containing the peak amount of 59 kDa was electrophoresed on a native gel, using the same system as described above however without SDS and without boiling the samples before loading. The gel lane was sliced in 0.5 cm horizontal pieces and each piece was eluted individually for 48 hours in 50 mM Mes, pH 6. After centrifugation the resulting supernatant was analyzed both on SDS-PAGE and *in vitro* for antifungal activity. Results are shown in Figure 3. Only in those fractions containing MS59, was antifungal activity observed against *Phytophthora infestans* and *Pythium ultimum*.

EXAMPLE 3

In vitro antifungal assays on non-Oocymetes

In vitro fungal assays were performed as described in the general experimental part. As positive control *Phytophthora infestans* was tested. The peak of MS59 is located in fraction 4. Results are shown in Table 1.

Table 1. Antifungal effects of MS59 containing fractions from Mono-S, pH 6

10	fungus	spore stage *)	1	2	3	4	5	6	7/8
15	Fusarium oxysporum	spore	0	0	0	0	0	0	0
		germl.	2	2	2	2	3	3.5	3.5
	Fusarium solani	spore	0	0	0	0	0	0	0
	Phytophthora infestans	spore	0	2	2	4	3.5	2	1
	Phytophthora nicotianae	hyph	0	1	2	4	4	2	1
	Phytophthora cactorum	hyph	0	0	2	4	4	1	1
20	Pythium ultimum	hyph	0	0	0	4	4	0	0
	Pythium sylvaticum	hyph	0	0	0	2	1	0	0
	Pythium paroecandrum	hyph	0	0	0	2	2	0	0

*) spore = no pregermination, germl = germination until the germtube is 3-5 times the length of the spore, hyph. = hyphal fragments were used as starting inoculum.

GI: growth inhibition; a scale of 0 - 4 is used, 0 = no visible growth inhibition, 1 = weak (0 to 30%) inhibition, 2 = moderate (30 to 60%) inhibition, 3 = strong (60 to 90%) inhibition, 4 = very strong (100%) inhibition.

As can be seen *Phytophthora* and *Pythium* spp., appeared very sensitive to MS59.

EXAMPLE 4

Purification of an antifungal protein WL64 from lettuce
induced with salicylic acid

Leaves of 7 to 8 weeks old lettuce (*Lactuca sativa* cv. Lollo
5 bionda) plants were sprayed daily with 10 mM salicylate for 4 days.
After two hours the plants were extensively rinsed with water to
remove the sodium salicylate. On day 5, the leaves were harvested into
liquid nitrogen and stored at -80°C until further use.

Lettuce leaves were thawed and homogenized at 4°C in 0.5M NaOAc
10 pH 5.2, 0.1% β -mercaptoethanol (lettuce : buffer = 1: 1.5 (w/v)), and
10 grams active carbon per kg leaves, using a Waring blender. The
homogenate was centrifuged for 60 minutes at 9,000 g at 4°C. The
supernatant was subsequently filtered over 10 layers of cheese cloth.
The filtrate was brought to 40% saturation with ammonium sulphate and
15 centrifuged for 30 minutes at 9,000 g. The resulting supernatant,
containing 85% of protein and >95% of antifungal activity relative to
the crude homogenate, was subjected to hydrophobic interaction
chromatography.

The supernatant was filtered over a paper filter and applied to
20 a phenyl-sepharose 6FF High sub column (Pharmacia, 100 ml bed volume
in a Pharmacia XK 50/20 column) pre-equilibrated with 40% (1.45M)
ammonium sulphate in 50mM potassium-phosphate buffer, pH 6.0 (referred
to as buffer A) at a flow rate of 10 ml/min or less. The column was
washed with at least 10 column volumes of buffer A after which bound
25 protein was eluted with a decreasing salt gradient from 100% buffer A
to 20% buffer A (50mM KPi pH 6.0 as buffer B) over a period of 40 min
at a flow rate of 10 ml/min, followed by a linear decreasing gradient
from 20% A to 0% A (=100% B) over a period of 30 min at the same flow
rate. The column was washed for another 45 min with buffer B, after
30 which the elution was completed. One-minute fractions were collected
(10 ml/fraction). Fractions 40-75 (called the HIC-peak) contained
antifungal activity.

The pooled fractions were concentrated (using a stirred flow
cell and a YM 30 kDa membrane (Amicon)) and subsequently 15 times
35 diluted with 25mM sodium acetate, pH 4.5. This solution was applied to
a pre-packed Source S column (16/20, Pharmacia) with a flow rate of 10
ml/min. After washing of the column with 5 column volumes of said
buffer, protein was eluted from the column with an increasing NaCl

gradient (0-0.4M NaCl in 25mM NaOAc, pH 4.5) over a 60 min period, 2.5 ml/min, 1 min fractions. Fractions were collected in 250µl 1M potassium phosphate, pH 7.0, in order to neutralize the relatively acidic NaOAc buffer. The fractions containing antifungal activity
5 (fractions 25-45 (0.2-0.3M NaCl)) were pooled and are referred to as the Source S-peak.

The Source S-peak was concentrated and buffer exchanged to 25mM NaOAc, pH 4.5, resulting in a fraction of about 10 ml, and subjected to cation-exchange chromatography using a Mono S column (5/5,
10 Pharmacia). The column is eluted with the following NaCl gradients (NaCl in 25mM NaOAc, pH 4.5): 0-5 min, 0-0.1M NaCl; 5-20 min, 0.1-0.16M NaCl; 20-21 min, 0.16-0.25M NaCl; 21-31 min, 0.25M NaCl; 31-32 min, 0.25-1.0M NaCl, followed by 1.0M NaCl for 10 min after which the elution is completed. The antifungal activity eluted from the column
15 during the 0.25M NaCl step (usually fractions 22-30; the Mono-S peak). Flow-rate 1 ml/min, 1ml fractions, collected in 100µl 1M potassium phosphate, pH 7.0.

The Mono S-peak was concentrated to about 0.5-1.0 ml and subjected to gelfiltration chromatography (Superdex 75, 10/30,
20 Pharmacia), with 200mM NaCl in 50mM potassium phosphate, pH 7.0 as the running buffer. The sample volume was 200 µl; flow rate 0.5 ml/min; 0.5 ml/fraction. The antifungal activity elutes from the column at the position of the 66 kDa marker. Comparison of the active fractions (SD 75 peak) with the protein pattern on SDS-PAGE reveals a 64 kDa protein
25 as the most likely candidate for the lettuce-antifungal protein (Figs. 6A-C). This protein was named WL64.

The SD 75-peak was buffer-exchanged to pH 9.5 for chromatophocusing on a Mono P column (Pharmacia) according to the
manufacturers instructions. All activity was found in the flow-through
30 of the column (even in the case when the column was equilibrated to pH 11.0) although there was some separation (3 overlapping peaks in flow-through). The flow-rate was 0.5 ml/min; 0.5 ml/fraction. The fractions containing the anti-fungal activity were pooled and buffer-exchanged to 50mM MES, pH 6.0. Coomassie staining of the highest purified
35 protein fraction after SDS-PAGE revealed about 6 protein bands of which two bands of 64 kDa and 55 kDa, were the most prominent ones (Fig. 7). The estimated relative amounts of both proteins in the final fraction was 1/6-1/8 for the 64 kDa protein and 1/2-1/3 for the 55 kDa

protein. Although on gel it is shown that this column clearly contributes to the purification of the 64 kDa protein, the specific activity, as well as the recovery of the protein in the pooled fractions dropped considerably (see table 2).

5 A representative purification procedure is summarized in table 2.

Table 2. Purification of WL 64

Sample or Column	Protein (mg)	Activity (GI-units)	Spec. act. (GI-u/mg)	Purifica- tion (x-fold)	Recovery (%)
Lettuce (1.54 kg)					
Extract	685				
(NH ₄) ₂ SO ₄ sup	584	101250	173	1	100
HIC	174	44000	253	1.46	43
Source S	38.7	32400	837	4.84	32
Mono S	2.3	8960	3896	22.5	8.8
SD-75	0.452	8200	18142	105	8.1
Mono P	0.137	1752	12788	74	1.7

10 The activity is represented as growth inhibition units (GI-units). Four GI units represent the amount of protein that results in a growth inhibition of 100% in the *in vitro* assay as described in the general part of the Examples.

15

EXAMPLE 5

Elution of WL64 from native PAGE and subsequent testing

Since WL64 was not completely pure, it was further investigated whether or not the 64 kDa protein was indeed responsible for the observed antifungal activity. The Mono P fraction containing the peak amount of antifungal activity was submitted to electrophoresis on a native 10% polyacrylamide gel under acidic conditions, in the absence of SDS and β -mercaptoethanol and without boiling. Two adjacent gel lanes were sliced in 0.3 cm horizontal pieces. One part was used

directly in the antifungal assay, the other part was subjected to SDS-PAGE under denaturing conditions. Growth inhibition clearly correlated to the 64 kDa protein and not to the 55 kDa protein.

5

EXAMPLE 6**Glycosylation of WL64**

WL64, as well as the 55kDa protein are glycosylated as illustrated by binding to concanavalin A and by the DIG-Glycan-detection kit (Boehringer). Both proteins were not sensitive to glycopeptidase-F treatment, indicating that the glycosylation is probably O-linked.

EXAMPLE 7**Amino acid sequencing of WL64**

For N-terminal amino acid sequencing an amount of 21 µg of purified protein (representing about 4µg WL64) was separated on a 7.5% polyacrylamide gel and was subsequently blotted onto PVDF membrane. The membrane was stained with 0.1% Serva Blue G in 45% methanol, 10% acetic acid for 5 minutes at room temperature and destained with 45% methanol, 10% acetic acid. The 64 kDa band was cut out and sequenced using Edman degradation on an Applied Biosystems 477A protein sequencer according to the protocol provided by the manufacturer.

For internal protein sequencing 105 µg of purified protein (representing about 20µg WL64) was separated on a 7.5% SDS-polyacrylamide gel. The gel was stained with 0.2% Serva Blue G in 20% methanol, 0.5% acetic acid for 20 min at room temperature and destained with 30% methanol at room temperature for about 1 hour. The 64 kDa band was cut out and the protein was subsequently digested with trypsin. The digestion products were separated on a reverse phase column and analyzed by Edman degradation.

Besides the N-terminal sequence (SEQ ID NO: 49), two tryptic fragments were sequenced (SEQ ID NO: 50 and SEQ ID NO: 51).

SEQ ID NO: 49: Thr-Ser-Thr-Ser-Ile-Ile-Asp-Arg-Phe-Thr-Gln-(Cys/Ser)-Leu-Asn-Asn-Arg-Ala-Asp-Pro-(Ser)-(Phe)-

SEQ ID NO: 50: (Ser)-Ile-(???) -Val-(Ser)-Ile-Glu-Asp-Glu-Thr-Ala-(Trp)-Val-Gln-Ala-Gly-Ala-Thr-Leu-Gly-Glu-Val-Tyr-(Tyr)-

SEQ ID NO: 51: Ala-Asp-Pro-Ser-Phe-Pro-Leu-Ser-Gly-Gln-Leu-Tyr-Thr-Pro-

The amino acid residues between brackets could not be identified unambiguously.

EXAMPLE 8

5 Anti-fungal activity of MS59 and WL64

Based on the sequence homology between MS59 and WL64, both proteins appear to be very related to each other. This might also be the case for their anti-fungal activity, as well as for their specific activities towards the respective fungi. This hypothesis was tested
10 and the results are summarized in table 3.

Table 3. Anti-fungal activity of MS59 and WL64 ¹

Pathogen	Amount of WL64 needed for complete inhibition (GI 4) (ngram per assay)	Amount of MS59 needed for complete inhibition (GI 4) (ngram per assay)
<i>Phytophthora</i> <i>infestans</i>	10	5
<i>Pythium ultimum</i>	10	5
<i>Rhizoctonia solani</i>	20	10
<i>Tanatephorus</i> <i>cucumeris</i>	20	n.t.
<i>Helicobasidium</i> <i>purpureum</i>	15	7.5
<i>Sclerotium cepivorum</i>	40	20
<i>Pichia pastoris</i>	n.t.	5
<i>Botrytis cinerea</i>	200	10

n.t. = not tested

15 ¹ Antifungal assays were carried out as described in the general experimental part.

Note that the amounts of protein were estimated by means of Coomassie staining on SDS-PAGE gels, meaning that the amounts of protein depicted here are indicative, rather than absolut .

EXAMPLE 9

Oxidase activities

A 50ml culture of *Rhizoctonia solani* in potato dextrose broth was extensively sonicated on ice and subsequently centrifuged at 3,000 g for 20 minutes at 4°C. The resulting supernatant was then centrifuged at 25,000 g for 1 hour. The pellet was washed twice with demineralized water and resuspended in 1 ml water containing 1.0% Triton X-100. In this way a fungal cell wall suspension was obtained.

Oxidase activity was measured utilizing the reagent 4-amino-antipyrine (4-AAP), based on Gallo, 1981 (Gallo, Methods in Enzymology, 71:665-668, 1981). A reaction volume of 500µl contained 50mM potassium phosphate buffer pH 7.0, 25µM FAD, 10mM NaN₃, 0.01% Triton X-100, 6mM 2,4,6,tribromohydroxybenzoic acid, 2mM 4-AAP, and 10 units horseradish peroxidase. Hydrogenperoxide production was measured at 510nm. Known amounts of hydrogen peroxide were included for calibration.

WL64, as well as MS59, performed oxidase activities using the fungal cell wall suspension as substrate. Different substances were subsequently tested as possible substrates, a.o. some carbohydrates and amino acids (see example 10). Glucose, and other carbohydrates were found to serve as substrate for the oxidase activity of both MS59 and WL64.

Since MS59 and WL64 displayed carbohydrate and especially glucose oxidase activity it was investigated whether the fungal cell wall suspension could serve as a substrate for glucose oxidase (GOX) from *Aspergillus niger* (Sigma, G 2133). This was indeed the case. Kinetic studies showed that MS59, WL64 and GOX display Michaelis-Menten kinetics when glucose is used as substrate, as illustrated by means of a Lineweaver-Burk plot (Fig 8A). The K_m values for MS59 and WL64 were more than one order of magnitude lower than that for GOX: 19.5µM and 23.3µM for WL64 and MS59 respectively and 359µM for GOX. This means that the affinity for glucose is much higher for MS59 and WL64 than that of GOX. The V_{max} values were, however, comparable being 5.7, 16.8, and 9.7µmol H₂O₂/min/mg protein for WL64, MS59 and GOX, respectively.

Kinetic studies using the fungal cell wall suspension as substrate showed Michaelis-Menten kinetics for both MS59 and WL64, but

not for GOX as shown in Fig. 8B. The K_m values for MS59 and WL64 were 4.7 μ l and 24.3 μ l respectively, using the suspension described above. The V_{max} values were 22.0 and 11.2 μ mol H_2O_2 /min/mg protein for respectively MS59 and WL64. Since GOX with fungal cell walls as substrate does not show a linear relationship in a Lineweaver-Burk plot, the K_m and V_{max} could not be extrapolated from the plot. The kinetic data are summarized in table 4.

Table 4. Kinetic parameters of MS59, WL64 and glucose oxidase

Enzyme	Glucose		Fungal Cell Wall Suspension	
	K_m (μ M)	V_{max} (μ mol H_2O_2 /min /mg)	K_m (μ l)	V_{max} (μ mol H_2O_2 /min/mg)
MS59	23.3	16.8	4.7	22.0
WL64	19.5	5.7	24.3	11.2
GOX	359	9.7		

EXAMPLE 10

Substrate specificities

Different substances were tested as possible substrates. Among sucrose, sorbitol, fructose, c.m. cellulose, β -alanine, aspartic acid, chitine, cellulose, glutamate, glycine-glycine, laminarin, and glucose, only the latter served as substrate for at least WL64. Concentrations of the various substrates varied between 5mM and 50mM. It was further investigated whether glucose was the only substrate for MS59 and WL64 or that other carbohydrates could also be oxidized. The enzyme assays were performed as described in Example 9, the substrate concentrations being 50mM. GOX was shown to oxidase glucose exclusively (Fig. 9). Same figure shows that MS59 and WL64 display a much broader substrate specificity, ranging from C_4 -sugars to di- and polysaccharides. The highest (and almost equal activities) were obtained with D-glucose, D-mannose, D-galactose, cellobiose, maltose, and lactose (Fig. 9). This range of substrates resembles the range found to be converted by hexose oxidase (EC. 1.1.3.5).

EXAMPLE 11**Identification and characterization of genes homologous to the deduced MS59 nucleotide sequence**

Based on the amino acid sequences of pep1 (a.a. 12 to 22 of SEQ ID NO: 1) and pep2 (a.a. 2 to 12 of SEQ ID NO: 2), primers were designated for PCR. Genomic DNA was isolated from sunflower cv. Zebulon and PCR primers 4 (5' AAC TTC TCC IAG IGT IGC ICC IGC TTG IAC CCA3', SEQ ID NO: 3) and 5 (5' GAT CCI TCT TTC CCI ATT ACT GGI GAG GTT TA3', SEQ ID NO: 4) were used to amplify a 354 bp DNA fragment from the sunflower genome with PCR. PCR products corresponding to this fragment size were cloned (SEQ ID NO: 5). Sequence analysis of the product revealed the presence of an uninterrupted Open Reading Frame (ORF) (SEQ ID NO: 6) of which the first and last stretch of amino acids corresponded with the amino acid sequences of SEQ ID NO: 1 and SEQ ID NO: 2. Several clones sequenced contained point mutations, varying from 1 to 4 in this PCR fragment. All but one of these mutations were silent mutations (nucleotide nr 57 T to C, nucleotide nr 63 C to A, nucleotide nr 225 A to G) which therefore did not alter amino acid sequences encoded. One clone however did contain a point mutation (nucleotide nr 203 G to A) which altered the amino acid sequence at amino acid 68 from Arg to Lys.

A southern blot of sunflower genomic DNA, probed with SEQ ID NO: 5 indicated the existence of multiple homologous sequences in the genome. Using SphI, 6 bands were detected, EcoRV 5 bands, SpeI 3 bands and NdeI 4 bands. With other enzymes 3-4 bands were previously discerned. This analysis suggests the existence of 3 genes with (partial) homology to the ms59 sequences.

New PCR primers were developed based on the non-variable areas between the original PCR primer sequences. Primers: for 3' RACE: 5' CAG GCA GCT GTG GTT TGT GGC 3' (SEQ ID NO: 7), for 5' RACE: 5' GTC CAC AAT GAA GAA GGG TTG 3' (SEQ ID NO: 8) and for nested 3' RACE: 5' ACG TAG ATA TCG AAC AAG AAA CCG C 3' (SEQ ID NO: 9).

35

Poly(A) containing RNA was isolated from sunflower leaf material that was induced by spraying 5 times with a 10 mM sodium salicylate solution. cDNA was prepared and 5' and 3' RACE PCR reactions were

performed as described in the instructions of the Marathon™ kit (Clontech laboratories, Inc., Palo Alto, CA). Partial cDNA clones were isolated by 5' and 3' RACE PCR reactions. Sequence analysis confirmed the identity of the partial cDNA clones.

- 5 Again new PCR and nested PCR primers were developed based on newly obtained sequence information from cloned 5' and 3' RACE PCR products. Primers for 5'RACE: 5' CTG GGG AAG CCC GTG TAG TAA AGC 3' (SEQ ID NO:11), 5' CGG GAA GTT GCA GAA GAT TGG GTT G 3' (SEQ ID NO:13), for nested 5'RACE: 5' GAG CAA GAG AAG AAG GAG AC 3' (SEQ ID NO:14), for 3' RACE: 5' GCT TTA CTA CAC GGG CTT CCC CAG 3' (SEQ ID NO: 10), and for nested 3' RACE: 5' GGT ACT CCA ACC ACG GCG CTC 3' (SEQ ID NO:12). Four partial cDNA clones were isolated which together encode all of the Open Reading Frame including a putative signal peptide followed by an approximately 59 kDa protein, and 5' and 3' UTR's (untranslated regions) (SEQ ID NO: 15). A full length cDNA clone of 1784 bp, of which the ORF (pos. 21 to pos. 1608) encodes 529 amino acid residues (SEQ ID NO:16), could be assembled out of these four partial cDNA clones and the PCR fragment mentioned above (SEQ ID NO: 5).
- 15 The amino-terminal signal sequence (Von Heijne et al., 1983 and Von Heijne, 1985) is not likely fully presented within the first 19 amino acid residues. A prediction of the putative cleavage site was made.
- 20

- The amino acid sequence of this cDNA clone was used in a BLAST homology search. This sequence revealed high homology to the Berberine Bridge Enzymes (BBE) from Californian poppy (*Eschscholtzia californica*) (Dittrich and Kutchan, 1991, Proc. Natl. Acad. Sci. USA 88, 9969-9973) and Opium poppy (*Papaver somniferum*) (Facchini et al., 1996, Plant Physiol. 112, 1669-1677).
- 25

- BLAST screening of Expressed Sequence Tag (dbEST) databases with the amino acid sequence as shown in SEQ ID NO: 16 revealed homologues of the MS59 protein in *Arabidopsis thaliana* (SEQ ID NO: 21 to SEQ ID NO: 47) and rice (SEQ ID NO: 48).
- 30

- The EST sequences are listed in the sense orientation considering the orientation of homology to MS59. Sequences of the EST clones were altered by inserting one or two extra unknown nucleotides (N or NN) at frameshift positions in order to obtain one single translation frame with homology to MS59.
- 35

EXAMPLE 12**Isolation of the gene encoding WL64 and determination of th
nucleotid sequence**

Based on the amino acid sequence of the amino-terminus of the
5 WL64 protein (SEQ ID NO: 49, Thr-Ser-Thr-Ser-Ile-Ile-Asp-Arg-Phe-Thr-
Gln-(Cys/Ser)-Leu-Asn-Asn-Arg-Ala-Asp-Pro-(Ser)-(Phe)-) a primer (a.a.
1 to 11 of SEQ ID NO: 49) was developed for PCR.
The N-terminal amino acid sequence (SEQ ID NO: 49) revealed high
homology to the corresponding portion of the MS59 protein (amino acid
10 residues 20 to 39 in SEQ ID NO: 16).
cDNA was prepared from Poly(A) containing RNA that was isolated from
lettuce (*Lactuca sativa* cv. Lollo bionda) leaves that were induced by
spraying 5 times with a 10 mM sodium salicylate solution. PCR primers
FR-WL64-142 (5'ACT TCT ACT TCT ATT ATT GAT AGG TTT ACT CA3', SEQ ID
15 NO: 52) and MS59 primer 4 (5'AAC TTC TCC IAG IGT IGC ICC IGC TTG IAC
CCA3', SEQ ID NO: 3) were used to amplify a 405 bp fragment from the
lettuce cDNA pool. PCR products corresponding to this PCR fragment
were cloned and sequenced (SEQ ID NO: 53) and revealed an
uninterrupted open reading frame (SEQ ID NO: 54).

Table 5. EST sequences showing homology to MS59.

Frameshifts were introduced for optimal aligning of the EST's with the MS59 sequence. In the columns with frameshift 1 and frameshift 2 the position of the frameshift and the shift (frame--->frame) are listed.

5 The (-) mark means, no frameshift present.

SEQ ID NO:	EST name	GenBank accession	Frame (1, 2, 3)	Frameshift 1	Frameshift 2
21	ATTS5925	F19886	2	-	-
22	ATTS0345	Z17771	2	202, 2 ----> 1	-
23	ATTS5268	F14356	2	298, 2 ----> 3	-
24	TC13883	-	1	177, 1 ----> 2	-
25	TC11550	-	2	-	-
26	P_16053	R84094	1	-	-
27	P_22214	N97049	2	310, 2 ----> 3	-
28	P_16873	R90518	3	317, 3 ----> 2	-
29	ATTS2532	Z30784	3	188, 3 ----> 1	312, 1 ----> 2
30	TC11456	-	3	-	-
31	P_8818	T45555	1	98, 1 ----> 3	-
32	P_21340	N96011	1	-	-
33	P_22585	W43206	2	367, 2 ----> 1	-
34	Q_ATTS2533	Z30785	2	-	-
35	P_17333	H76902	2	-	-
36	P_9615	T46352	1	-	-
37	Q_ATTS2959	Z33920	2	-	-
38	P_2730	T20722	1	-	-
39	TC9870	-	2	-	-
40	P_14876	H36354	2	241, 2 ----> 1	-
41	P_21353	N96040	1	89, 1 ----> 2	-
42, 43	Q_ATTS3343	Z34583	1	-	-
44	Q_ATTS4954	F14032	2	139, 2 ----> 1	-
45	Q_ATTS1606	Z26512	2	-	-
46	P_7866	T44603	1	222, 1 ----> 3	-
47	AA0410042	24308	2	421, 2 ----> 1	-
48	RICS2381A	D40415	3	-	-

New PCR primers were developed based on the sequence of SEQ ID NO: 53 that is located between the original PCR primers. Primers for 5' RACE: 5' CAC GTT TAT GGA GCG TAA GTT GAA C3' (SEQ ID NO: 55) and for 3' RACE: 5' CAC CCT TCA CAC ATT CAA GCA GC3' (SEQ ID NO: 56) were synthesized and used in 5' and 3' RACE PCR reactions, performed as described in the instructions of the Marathon™ cDNA amplification kit (Clontech laboratories, Inc., Palo Alto, CA). Two partial cDNA clones were amplified by 5' and 3' RACE reactions. Sequence analysis confirmed the identity of the partial cDNA clones which together encode all of the open reading frame including a putative signal peptide and 5' and 3' UTR's (untranslated regions). A full length cDNA clone of 1981 bp (SEQ ID NO: 57) was assembled of which the ORF (pos. 7 to pos. 1629) encodes 540 amino acid residues (SEQ ID NO: 58). The amino terminal signal sequence is represented by the first 27 amino acid residues.

15

EXAMPLE 13

Characterization and isolation of Berberine Bridge Enzyme genes from *Papaver somniferum* and *Eschscholtzia californica*

Genomic DNA was prepared from leaves of full grown Californian poppy (*Eschscholtzia californica*) and Opium poppy (*Papaver somniferum* cv Marianne) plants.

Primers were designed for the Californian poppy gene (EcBBE) at the start of the mature protein (5' GGT AAT GAT CTC CTT TCT TGT TTG ACC 3', SEQ ID NO: 59) and at the stop codon introducing a Not I restriction site just downstream of the TAG stop codon (5' AGA GCG GCC GCT ATA TTA CAA CTT CTC CAC CAT CAC TCC TC 3', SEQ ID NO: 60).

For the Opium poppy gene (PsBBE) primers were designed in a similar way at the start of the presumed mature protein (5' GGT GAT GTT AAT GAT AAT CTC CTC 3', SEQ ID NO: 61) and at the TAG stop codon introducing a Not I restriction site (5' AGA GCG GCC GCT ACA ATT CCT TCA ACA TGT AAA TTT CCT C 3', SEQ ID NO: 62).

These primers were used to amplify the mature portion of both the BBE genes.

The PCR products were digested with Not I and ligated into vector pET32a (Novagen, Madison, WI) digested with *EcoR* V and Not I. The correct insertion of the fragment was confirmed using restriction enzyme analysis and DNA sequencing.

EXAMPLE 14

**Characterization and isolation of MS59 homologu s fr m
Arabidopsis thaliana**

In our blast screening we identified 26 EST's with homology to MS59. One EST was found in Rice and the remaining 25 were all found in *A. thaliana*. Homologous EST's were found over the entire length of the MS59 sequence. Analysis of the *Arabidopsis* expressed sequence tags revealed that there are 3 EST's with high homology at the 5' end of the protein (SEQ ID NO: 21, SEQ ID NO: 39 and SEQ ID NO: 40) of which SEQ ID NO: 39 and SEQ ID NO: 40 are overlapping sequences. The 3' part of MS59 showed homology to 7 EST sequences (SEQ ID NO: 24, 27, 32, 34, 41, 43 and 45) of which SEQ ID NO: 24 is overlapping with SEQ ID NO: 43 and SEQ ID NO: 32 is overlapping with SEQ ID NO: 45.

Primers were designed, located at the start of the presumed mature part (possible cleavage sites were predicted according to consensus sequences described by Von Heijne et al., 1983 and Von Heijne, 1985) of the two different EST's homologous with the 5' part of MS59 (SEQ ID NO: 16).

The EST sequence represented by SEQ ID NO: 21 possibly missed the first three amino acid residues of the predicted mature part when compared to the MS59 amino acid sequence (SEQ ID NO:16) and, therefore, A.a. residues 20 to 22 of SEQ ID NO:16 were introduced by including 9 nucleotides at the 5' end of the primer.

Primer located 5' in SEQ ID NO: 21, adding residues 20 to 22 of MS59 (SEQ ID NO: 16): 5' ACT TCC CGT AGA AAC TCG GAG ACT TTC ACA CAA TGC 3' (SEQ ID NO: 63).

Primer located behind the predicted cleavage site of SEQ ID NO: 39 and SEQ ID NO: 40: 5' TCC ATC CAA GAT CAA TTC ATA AAC TGT GTC (SEQ ID NO: 64).

Primers were also made located around the stopcodon of the five different EST's homologous with the 3' part of the MS59 a.a. sequence (SEQ ID NO: 16) and introducing a Not I restriction site for cloning in the pET32a *E. coli* expression vector.

Primer located in SEQ ID NO: 24 and SEQ ID NO: 43, 5' AGA GCG GCC GCT TTC ATG AAC CTA GCT TCT AGT AGG 3' (SEQ ID NO: 65). Primer in SEQ ID NO 27, 5' AGA GCG GCC GCG AAA TGG CCC CCC TTT TAA AAC GGG G 3' (SEQ ID

NO: 66). Primer in SEQ ID NO:32 and SEQ ID NO: 41, 5' AGA GCG GCC GCA
AAT GAT ATC TTC AGG TAA CTT TGT TCA C (SEQ ID NO: 67). Primer in SEQ
ID NO: 34, 5' AGA GCG GCC GCA TAA TCA AAT AAA TAC ACT TAT GGT AAC ACA
G (SEQ ID NO: 68) and the primer in SEQ ID NO: 45, 5' AGA GCG GCC GCT
5 GGT TTT GTA TTG AGG ACT CAA AAC AG 3' (SEQ ID NO: 69).

All possible combinations of the 5' primers with the 3' primers were
used in a PCR on genomic DNA isolated from *Arabidopsis thaliana* cv
Columbia. In a PCR with the primers SEQ ID NO: 63 and SEQ ID NO: 68 an
10 approximately 1800 bp band was amplified. This band was cloned and
identity of the PCR product was confirmed by DNA sequencing. The
cloned PCR product of 1757 bp (SEQ ID NO: 70) contained an intron from
position 570 to position 801, the open reading frame of SEQ ID NO: 70
consists of 508 amino acid residues (SEQ ID NO: 71).

15 Total RNA was isolated from *Arabidopsis thaliana* Col-0 from 12 days
old sterile etiolated seedlings grown in the dark on Murashige and
Skoog agar, from 12 days old sterile seedlings grown in liquid
Murashige and Skoog medium with a 16 hour photoperiod and from leaves,
stems, flowers and siliques from full grown plants (Newman et al.,
20 1994 Plant Physiol. 106: 1241-1255). The RNA from the different
developmental stages was pooled. Poly(A)⁺ RNA was isolated using the
Poly(A) Quick® mRNA Isolation kit (Stratagene, La Jolla, CA) and cDNA
was prepared using the Marathon™ cDNA Amplification Kit (Clontech
Laboratories Inc., Palo Alto, CA).

25 PCR reactions were set up with the cDNA pool with different
combinations of 5' primers and 3' primers. A PCR product was amplified
with the primer combination SEQ ID NO: 63 and SEQ ID NO: 68 of
approximately 1600 bp. The PCR product was cloned in the *EcoR* V and
30 *Not* I restriction sites of the bacterial expression vector pET32a
(Novagen, Madison, WI). The sequence of the PCR product was determined
and revealed an uninterrupted open reading frame of 1527 bp (SEQ ID
NO: 72) representing a protein of 508 amino acid residues (SEQ ID NO:
73).

35 A second cDNA clone of about 1600 bp was amplified with the primer
combination SEQ ID NO: 64 and SEQ ID NO: 65. This cDNA clone was also
ligated into the *EcoR* V and *Not* I restriction sites of pET32a

(Novagen, Madison, WI). This cDNA PCR clone was also characterized by DNA sequencing and consisted of an uninterrupted open reading frame of 1530 bp (SEQ ID NO: 74) encoding 509 amino acid residues (SEQ ID NO: 75).

5

EXAMPLE 15

Expression of MS59, the Berberine Bridge Enzymes from *Papaver somniferum* and *Eschscholtzia californica* and two homologous proteins from *Arabidopsis thaliana* cv Columbia in *E. coli*

10

A PCR fragment containing the presumed mature portion of MS59 was introduced in vector pET32c (Novagen, Madison, WI), and the correct insertion of the fragment is confirmed using DNA sequencing. Then, the plasmid was introduced into *E. coli* AD494 (DE3) pLyss (Novagen, Madison, WI). Small scale cultures (2 ml) of several colonies were then started of which half is induced by the addition of IPTG to 1mM final concentration. Total extracts from *E. coli* were run on SDS gels and analyzed by Coomassie Brilliant Blue staining. Several clones exhibited strong overexpression of the MS59 protein. A clone which had strong overexpression was selected for a large scale culture. Five hundred ml of LB supplemented with 0.4 mM glucose was inoculated with a culture of this *E. coli* and grown to an optical density of 0.5-0.7. Then, IPTG was added to a final concentration of 1 mM and protein production allowed for 3 hours at 30°C. A large proportion of the MS59 protein was found in the insoluble protein fraction, a small amount appeared soluble. The resulting insoluble protein preparation contained mainly MS59 protein. This preparation is used for raising antibodies (Example 17). The soluble fraction was used in an *in vitro* assay to test whether the MS59 protein still exhibited antifungal activity.

20
25
30

The pET32a plasmids containing the open reading frames of the four MS59/WL64 homologues were introduced into *E. coli* AD494(DE3)pLyss (Novagen, Madison, WI). Small scale cultures (25 ml) of several independent clones were grown to an optical density of 0.5-0.7. Then IPTG was added to a final concentration of 1mM and protein production was allowed for 4 hours at 30°C.

35

Soluble and total protein fractions were isolated. The samples were analyzed using SDS-PAGE followed by Neuhoff staining and Western

analysis using the S-Tag Western Blotting detection kit (Novagen, Madison, WI). A large portion of the protein was found in the insoluble fraction, only a small amount appeared to be soluble. Clones which strongly overexpressed the homologous proteins were selected for production of the proteins in large scale cultures of 1.5 liter each.

EXAMPLE 16

In vitro antifungal assays of MS59, MS59/WL64 homologues from Californian poppy (*Eschscholtzia californica*) and Opium poppy (*Papaver somniferum*) and two homologous proteins from *Arabidopsis thaliana*

The MS59 protein produced in *E. coli* contained N-terminal trxA-, His- and S-Tags. The His-tag was used for purification of the soluble MS59 on an IMAC (immobilized metal affinity chromatography) column, charged with Ni^{2+} . Bound protein was eluted by increasing the imidazole concentration. The peak fraction from this purification contains some contaminating *E. coli* proteins.

The peak fraction of this MS59 purification was dialysed into 50 mM MES, pH 6.0, and used in an in vitro assay with *Phytophthora infestans* and *Pythium ultimum*. For the standard setup of the in vitro antifungal assay with *Phytophthora infestans* and *Pythium ultimum* see above.

As control treatment we assayed an unrelated His-tagged protein purified from the same expression host, with some *E. coli* protein background. Also a boiled MS59 control (heated 10 minutes at 100°C) was included. Approximately 40 ng of fusion protein was tested in the *Phytophthora infestans* assay, twice that amount was used for the *Pythium ultimum* inhibition assay.

Microtiter dishes were wrapped with Parafilm and incubated in the dark at room temperature. After 2-3 days the mycelium of the growing fungus in the wells was stained with lactophenol cotton blue and the extent of growth was estimated.

IMAC fractions from the soluble fraction of *E. coli* containing MS59 showed complete inhibition of *P. infestans* and *P. ultimum* at concentrations of 20-40 ng.

Table 6. Antifungal effects of MS59 from *E.coli* on *Phytophthora infestans*

Fraction	MS59 _{E. coli}	MS59 _{E. coli} boiled	His- protein E.coli	MES buffer
Growth inhibition	4	0	0	0
amount of extract	5µl	5µl	5µl	

- 5 Growth inhibition (GI) is scored visually on a linear scale of 0 (no inhibition) to 4 (complete growth inhibition).

Table 7. Antifungal effects of MS59 from *E.coli* on *Pythium ultimum*

Fraction	MS59 _{E. coli}	MS59 _{E. coli} boiled	His- protein E.coli	MES buffer
Growth inhibition	4	0	0	0
amount of extract	10µl	10µl	10µl	

10

Microscopical analysis of the wells indicate the rapid germination and subsequent growth of *Phytophthora infestans* zoospores in each of the controls. Germination is near completely inhibited in the reactions containing the MS59 protein from *E. coli*. Some spores do germinate, but hyphal tip growth appears to stop soon after initiation. After 48 hours growth of *Phytophthora infestans* mycelium is abundant in the controls, but almost undetectable in the assay containing MS59. Even after 72 hours no substantial growth is observed. Fungal hyphae appear somewhat granular and thickened in the reactions containing MS59 protein. Examples of the characteristic patterns of fungal growth in incubations with and without *E. coli*-

produced MS59 are depicted in figure 4. After 48 and 72 hours fungal growth in the control incubations is so extensive no photographic material could be gathered. Incubations in the presence of MS59 leads to complete blockage of further growth, the germination tubes observed at 24 hours do not noticeably extend further.

Likewise, in the *Pythium ultimum* inhibition assay, where mycelium fragments are used, no growth is apparent upon treatment with MS59 (see fig. 5). After 24 hours the control reactions were completely overgrown by mycelium. Only small mycelium fragments are at that stage apparent in the MS59-treated sample.

The poppy homologues were expressed in *E. coli* (pET32a) and tested for *in vitro* antifungal activity on *Phytophthora infestans* and *Pythium ultimum*. *Phytophthora infestans* spores and hyphal fragments of *Pythium ultimum* were suspended in respectively sterile water or potato dextrose broth (PDB). 400-600 spores or 200 fragments/50 µl were added to each well.

The expressed proteins were partially purified by means of IMAC column chromatography. Fractions containing the expressed proteins were buffer exchanged to 50mM MES, pH 6.0, filter sterilized, and tested for their antifungal activity, with IMAC purified *E. coli* pET32-MS59 as a positive control.

No antifungal activity was observed for both *Eschscholtzia californica* and *Papaver somniferum* MS59/WL64 homologues, not even at concentrations ten times higher than that of the positive control, leading to a 100% growth inhibition of both fungi. This could mean that these *E. coli* expressed proteins did not have the correct folding and therefore showed no biological activity.

EXAMPLE 17

Raising antibodies against denatured MS59

Antibodies were raised in rabbits against denatured MS59 (the solubilized form from the insoluble *E. coli* fraction) in PAG-slices. The antibodies showed cross-reaction with a band of about 60 kDa only in the IF of pMOG1180 (Examples 18 and 19) tobacco plants containing antifungal and glucose oxidase activity. Surprisingly, no cross-reaction with WL64 is found.

EXAMPLE 18**Tailoring a MS59 clone for expression in transgenic plants**

PCR primers were developed based on the sequence around the ATG start codon and the TGA stop codon for cloning of the open reading frame (ORF). A NcoI restriction site was introduced at the ATG start codon for fusion to a constitutive promoter by PCR using primer: 5' GCC ATG GAG ACT TCC ATT CTT ACT C 3' (SEQ ID NO:16). The second codon of the ORF was changed from caa (Q) to gag (E) as a result of the introduced NcoI restriction site.

- 10 Downstream of the TGA stop codon a BamHI restriction site was introduced by PCR using primer: 5' GCC GGA TCC TCA AGA TGA CAA AGT TGG GAT GCT 3' (SEQ ID NO:18).

- Using a PCR reaction with Pfu DNA polymerase, we amplified the entire ORF, using the PCR primers to introduce the NcoI restriction site on the startcodon ATG and the BamHI restriction enzyme recognition site just downstream of the stopcodon. The integrity of the DNA sequence was confirmed by sequencing (SEQ ID NO:19). The entire ORF was linked to a constitutive promoter which allows high level protein expression in most parts of the plant. After the ORF a 3' untranslated region of the potato proteinase inhibitor II (Thornburg et al., 1987, Proc. Natl. Acad. Sci. USA 84, 744-748), which contains sequences needed for polyadenylation (An et al., 1989, Plant Cell 1, 115-122), was introduced. The chimeric gene produced was introduced into binary vector pMOG800 (deposited at the Centraal Bureau voor Schimmelcultures, Baarn, The Netherlands, under CBS 414.93, on August 12, 1993). The resulting clone pMOG1180, which harbours the MS59 construct under control of the ocs-mas hybrid promoter (WO95/14098) was introduced in *Agrobacterium tumefaciens* strain EHA105, suitable for transformation of target crops tomato and potato, strain MOG101 for transformation of tobacco and *Arabidopsis* and MOG301 for transformation of *Brassica napus*.

EXAMPLE 19**Production and analysis of transgenic tobacco and potato plants containing the MS59 gene construct**

Using *Agrobacterium* mediated transformation system binary

5 constructs containing the MS59 gene construct as described in Example 18 were introduced into tobacco and potato. The transgenic shoots of these different plant species were regenerated into whole plants and subsequently, primary transformants were analyzed for expression of the newly introduced MS59 gene. For this analysis use was made of
10 Western blotting techniques, using antibodies against MS59 specific peptide coupled to BSA. All antisera were diluted 1:5,000. A concentration series of purified proteins (12.5, 25, 50 and 100 ng) was used to judge the expression level of the introduced proteins in the transgenic plants. Transgenic samples were homogenized in 50 mM
15 sodium acetate buffer pH 5.2 and the extracts were clarified by centrifugation. The supernatants were either directly analyzed or left overnight to precipitate on ice. Overnight precipitation was always followed by clarification (by centrifugation). The protein concentration of the supernatants obtained in either way was
20 determined using Bradford reagent (Bradford 1976, Anal. Biochem. 72, 248-254) and BSA as the standard protein. As much protein as possible (but never more than 10 µg) was loaded on a 12.5 % SDS-PAA gel (Laemmli, supra) and immunoblotted as previously described (Ponstein et al. supra).

25

Extracts from leaves of ms59-transgenic tobacco and potato plants were made by pottering leaf fragments in a buffer containing 50 mM NaAc, (pH = 5.2). After this, insoluble protein was removed by centrifugation. Total soluble protein content was measured and the
30 equivalent of 10 µg was loaded on a SDS-gel. After running the gel the proteins were transferred to blot. This blot was developed using the antiserum raised against purified MS59 (Example 17). The MS59-specific antiserum was used in a 1:5,000 dilution. Purified MS59 was also run alongside on the gel, and is included for reference.
35 A number of transformed plants selected based on their high level expression of MS59 protein and S1 progeny plants will be tested in fungal infection assays.

EXAMPLE 20**Purification of MS59 transprot ins from tobacco transgenics**

Transgenic tobacco plants were produced expressing MS59 constitutively. Levels of expression are determined using Western analysis. Extracts of the transgenic material are assayed for *in vitro* growth inhibitory activity against *Phytophthora infestans* and *Pythium ultimum*. Small scale total extracts were made from *in vitro* leaves of tobacco containing the pMOG1180 construct (mas-ocs-promotor-MS59) and of tobacco control lines. The extracts were made by grinding leaf material in 50 mM NaAc pH 5.2. The supernatant was dialysed against 50 mM MES pH 6.0 and tested for *in vitro* antifungal activity according to the methods described in the general experimental part. Some of the tobacco pMOG1180 lines showed high antifungal activity on *P. infestans* and *P. ultimum* compared to other lines or control lines.

15

EXAMPLE 21**Carbohydrate oxidase activity / Localization of MS59 in transgenic tobacco**

Equal amounts of partial purified soluble MS59 and soluble homologue fractions (*Papaver*, *Eschscholzia*, *Arabidopsis*-A11 and -B7) were tested for carbohydrate oxidase activity. Carbohydrate oxidase activity for MS59 was 0.011 ODu/min and for the homologues 0.0003-0.0012 ODu/min, a difference of a factor 10.

From the transgenic pMOG1180 tobacco lines of Example 20 that showed antifungal activity *in vitro* IF was isolated at a later stage and tested for carbohydrate oxidase activity. Also the material that was left after IF isolation (called "-IF") was tested. The same lines that showed antifungal activity have high carbohydrate oxidase activity. The activity is located in the IF.

EXAMPLE 22

Introduction of the fur genes construct containing Chi-I, Glu-I, AP24 and MS59 under control of a constitutive plant promoter, into tomato, potato, carrot, *Brassica napus* and

5

Arabidopsis

Using *Agrobacterium* mediated transformation system binary construct pMOG1145 and pMOG1180 containing the genes encoding Chi-I, Glu-I, AP24 and MS59 or pMOG1146 containing the genes encoding Chi-I, Glu-I, bPR-1 and MS59 is introduced into different crop species
10 including, tomato, potato, carrot, *Brassica napus* and *Arabidopsis*.
S1 progeny plants are tested in fungal infection assays.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

5 (i) APPLICANT:

- (A) NAME: MOGEN International nv
- (B) STREET: Einsteinweg 97
- (C) CITY: Leiden
- (E) COUNTRY: The Netherlands
- 10 (F) POSTAL CODE (ZIP): 2333 CB
- (G) TELEPHONE: 31-(0)71-5258282
- (H) TELEFAX: 31-(0)71-5221471

15 (ii) TITLE OF INVENTION: Antifungal proteins, DNA coding therefor,
and hosts incorporating same.

(iii) NUMBER OF SEQUENCES: 75

(iv) COMPUTER READABLE FORM:

- 20 (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.25 (EPO)

25 (vi) PRIOR APPLICATION DATA:

- (A) APPLICATION NUMBER: EP 96.202.466.7
- (B) FILING DATE: 04-SEP-1996

(vi) PRIOR APPLICATION DATA:

- 30 (A) APPLICATION NUMBER: EP 97.200.831.2
- (B) FILING DATE: 10-MAR-1997

(2) INFORMATION FOR SEQ ID NO: 1:

35 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

40

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

45 (iii) ANTI-SENSE: NO

(v) FRAGMENT TYPE: internal

(vi) ORIGINAL SOURCE:

- 50 (A) ORGANISM: Helianthus annuus
- (B) STRAIN: cv. zebulon

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

Ser Ile Asn Val Asp Ile Glu Gln Glu Thr Ala Trp Val Gln Ala Gly
1 5 10 15

Ala Thr Leu Gly Glu Val Tyr Tyr Arg
20 25

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(v) FRAGMENT TYPE: internal

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Helianthus annuus*

(B) STRAIN: cv. zebulon

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Asp Pro Ser Phe Pro Ile Thr Gly Glu Val Tyr Thr Pro Gly Xaa Ser
1 5 10 15

Ser Phe Pro Thr Val Leu Gln Asn Tyr
20 25

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 33 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: YES

(ix) FEATURE:

(A) NAME/KEY: misc_feature

(B) LOCATION: 1

(D) OTHER INFORMATION: /function= "primer"

TCT TTT CCT ACC GTC TTG CAA AAC TAC ATC CGA AAC CTT CGG TTC AAT 96
 Ser Phe Pro Thr Val Leu Gln Asn Tyr Ile Arg Asn Leu Arg Phe Asn
 20 25 30

5 GAA ACT ACC ACA CCA AAA CCC TTT TTA ATC ATC ACA GCC GAA CAT GTT 144
 Glu Thr Thr Thr Pro Lys Pro Phe Leu Ile Ile Thr Ala Glu His Val
 35 40 45

10 TCC CAC ATT CAG GCA GCT GTG GTT TGT GGC AAA CAA AAC CGG TTG CTA 192
 Ser His Ile Gln Ala Ala Val Val Cys Gly Lys Gln Asn Arg Leu Leu
 50 55 60

15 CTG AAA ACC AGA AGC GGT GGT CAT GAT TAT GAA GGT CTT TCC TAC CTT 240
 Leu Lys Thr Arg Ser Gly Gly His Asp Tyr Glu Gly Leu Ser Tyr Leu
 65 70 75 80

20 ACA AAC ACA AAC CAA CCC TTC TTC ATT GTG GAC ATG TTC AAT TTA AGG 288
 Thr Asn Thr Asn Gln Pro Phe Phe Ile Val Asp Met Phe Asn Leu Arg
 85 90 95

TCC ATA AAC GTA GAT ATC GAA CAA GAA ACC GCA TGG GTC CAA GCC GGC 336
 Ser Ile Asn Val Asp Ile Glu Gln Glu Thr Ala Trp Val Gln Ala Gly
 100 105 110

25 GCC ACC CTC GGA GAA GTT 354
 Ala Thr Leu Gly Glu Val
 115

30 (2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 118 amino acids

(B) TYPE: amino acid

35 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

40 Asp Pro Ser Phe Pro Ile Thr Gly Glu Val Tyr Thr Pro Gly Asn Ser
 1 5 10 15

45 Ser Phe Pro Thr Val Leu Gln Asn Tyr Ile Arg Asn Leu Arg Phe Asn
 20 25 30

Glu Thr Thr Thr Pro Lys Pro Phe Leu Ile Ile Thr Ala Glu His Val
 35 40 45

50 Ser His Ile Gln Ala Ala Val Val Cys Gly Lys Gln Asn Arg Leu Leu
 50 55 60

Leu Lys Thr Arg Ser Gly Gly His Asp Tyr Glu Gly Leu Ser Tyr Leu
 65 70 75 80

55

Thr Asn Thr Asn Gln Pro Phe Phe Ile Val Asp Met Phe Asn Leu Arg
85 90 95
Ser Ile Asn Val Asp Ile Glu Gln Glu Thr Ala Trp Val Gln Ala Gly
5 100 105 110
Ala Thr Leu Gly Glu Val
115

10 (2) INFORMATION FOR SEQ ID NO: 7:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 base pairs
 (B) TYPE: nucleic acid
15 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- 20 (iii) HYPOTHETICAL: NO
- (iii) ANTI-SENSE: NO
- (ix) FEATURE:
25 (A) NAME/KEY: misc_feature
 (B) LOCATION: 1
 (D) OTHER INFORMATION: /function= "primer"

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

CAGGCAGCTG TGGTTTGTGG C

21

35 (2) INFORMATION FOR SEQ ID NO: 8:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 base pairs
 (B) TYPE: nucleic acid
40 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- 45 (iii) ANTI-SENSE: NO
- (ix) FEATURE:
 (A) NAME/KEY: misc_feature
50 (B) LOCATION: 1
 (D) OTHER INFORMATION: /function= "primer"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

55 GTCCACAATG AAGAAGGGTT G

21

(2) INFORMATION FOR SEQ ID NO: 9:

- 5 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- 10 (ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iii) ANTI-SENSE: NO
- 15 (ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /function= "primer"

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

ACGTAGATAT CGAACAAGAA ACCGC

25

(2) INFORMATION FOR SEQ ID NO: 10:

- 25 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
30 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
35 (iii) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

40 GCTTTACTAC ACGGGCTTCC CCAG

24

(2) INFORMATION FOR SEQ ID NO: 11:

- 45 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- 50 (ii) MOLECULE TYPE: cDNA
(iii) HYPOTHETICAL: NO
(iii) ANTI-SENSE: NO
55

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

CTGGGGAAGC CCGTGTAGTA AAGC

24

5 (2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 base pairs

(B) TYPE: nucleic acid

10 (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

15 (iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

20

GGTACTCCAA CCACGGCGCT C

21

(2) INFORMATION FOR SEQ ID NO: 13:

25 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

30

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

35 (iii) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

CGGGAAGTTG CAGAAGATTG GGTTC

25

40

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

45 (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

50

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

55

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

GAGCAAGAGA AGAAGGAGAC

20

5 (2) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1784 base pairs

(B) TYPE: nucleic acid

10 (C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

15 (iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

20 (A) ORGANISM: Helianthus annuus

(B) STRAIN: Zebulon

(ix) FEATURE:

(A) NAME/KEY: CDS

25 (B) LOCATION: 21..1608

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

30	ATATCACATC TTCTTTCAAC ATG CAA ACT TCC ATT CTT ACT CTC CTT CTT	50
	Met Gln Thr Ser Ile Leu Thr Leu Leu Leu	
	1 5 10	
35	CTC TTG CTC TCA ACC CAA TCT TCT GCA ACT TCC CGT TCC ATT ACA GAT	98
	Leu Leu Leu Ser Thr Gln Ser Ser Ala Thr Ser Arg Ser Ile Thr Asp	
	15 20 25	
40	CGC TTC ATT CAA TGT TTA CAC GAC CGG GCC GAC CCT TCA TTT CCG ATA	146
	Arg Phe Ile Gln Cys Leu His Asp Arg Ala Asp Pro Ser Phe Pro Ile	
	30 35 40	
45	ACC GGA GAG GTT TAC ACT CCC GGA AAC TCA TCT TTT CCT ACC GTC TTG	194
	Thr Gly Glu Val Tyr Thr Pro Gly Asn Ser Ser Phe Pro Thr Val Leu	
	45 50 55	
50	CAA AAC TAC ATC CGA AAC CTT CGG TTC AAT GAA ACT ACC ACA CCA AAA	242
	Gln Asn Tyr Ile Arg Asn Leu Arg Phe Asn Glu Thr Thr Thr Pro Lys	
	60 65 70	
55	CCC TTT TTA ATC ATC ACA GCC GAA CAT GTT TCC CAC ATT CAG GCA GCT	290
	Pro Phe Leu Ile Ile Thr Ala Glu His Val Ser His Ile Gln Ala Ala	
	75 80 85 90	
60	GTG GTT TGT GGC AAA CAA AAC CGG TTG CTA CTG AAA ACC AGA AGC GGT	338
	Val Val Cys Gly Lys Gln Asn Arg Leu Leu Lys Thr Arg Ser Gly	
	95 100 105	

	GGT CAT GAT TAT GAA GGT CTT TCC TAC CTT ACA AAC ACA AAC CAA CCC	386
	Gly His Asp Tyr Glu Gly Leu Ser Tyr Leu Thr Asn Thr Asn Gln Pro	
	110 115 120	
5	TTC TTC ATT GTG GAC ATG TTC AAT TTA AGG TCC ATA AAC GTA GAT ATC	434
	Phe Phe Ile Val Asp Met Phe Asn Leu Arg Ser Ile Asn Val Asp Ile	
	125 130 135	
10	GAA CAA GAA ACC GCA TGG GTC CAA GCC GGT GCG ACT CTT GGT GAA GTG	482
	Glu Gln Glu Thr Ala Trp Val Gln Ala Gly Ala Thr Leu Gly Glu Val	
	140 145 150	
15	TAC TAT CGA ATA GCG GAG AAA AGT AAC AAG CAT GGT TTT CCG GCA GGG	530
	Tyr Tyr Arg Ile Ala Glu Lys Ser Asn Lys His Gly Phe Pro Ala Gly	
	155 160 165 170	
20	GTT TGT CCA ACG GTT GGC GTT GGT GGG CAT TTT AGT GGT GGT GGG TAT	578
	Val Cys Pro Thr Val Gly Val Gly Gly His Phe Ser Gly Gly Gly Tyr	
	175 180 185	
	GGT AAT TTG ATG AGA AAA TAT GGT TTG TCG GTT GAT AAT ATT GTT GAT	626
	Gly Asn Leu Met Arg Lys Tyr Gly Leu Ser Val Asp Asn Ile Val Asp	
	190 195 200	
25	GCT CAA ATA ATA GAT GTG AAT GGC AAG CTT TTG GAT CGA AAG AGT ATG	674
	Ala Gln Ile Ile Asp Val Asn Gly Lys Leu Leu Asp Arg Lys Ser Met	
	205 210 215	
30	GGT GAG GAT TTG TTT TGG GCG ATC ACC GGC GGT GGT GGT GTT AGT TTT	722
	Gly Glu Asp Leu Phe Trp Ala Ile Thr Gly Gly Gly Gly Val Ser Phe	
	220 225 230	
35	GGT GTG GTT CTA GCC TAC AAA ATC AAA CTA GTT CGT GTT CCG GAG GTT	770
	Gly Val Val Leu Ala Tyr Lys Ile Lys Leu Val Arg Val Pro Glu Val	
	235 240 245 250	
40	GTG ACC GTG TTT ACC ATT GAA AGA AGA GAG GAA CAA AAC CTC AGC ACC	818
	Val Thr Val Phe Thr Ile Glu Arg Arg Glu Glu Gln Asn Leu Ser Thr	
	255 260 265	
	ATC GCG GAA CGA TGG GTA CAA GTT GCT GAT AAG CTA GAT AGA GAT CTT	866
	Ile Ala Glu Arg Trp Val Gln Val Ala Asp Lys Leu Asp Arg Asp Leu	
	270 275 280	
45	TTC CTT CGA ATG ACC TTT AGT GTC ATA AAC GAT ACC AAC GGT GGA AAG	914
	Phe Leu Arg Met Thr Phe Ser Val Ile Asn Asp Thr Asn Gly Gly Lys	
	285 290 295	
50	ACA GTC CGT GCT ATC TTT CCA ACG TTG TAC CTT GGA AAC TCG AGG AAT	962
	Thr Val Arg Ala Ile Phe Pro Thr Leu Tyr Leu Gly Asn Ser Arg Asn	
	300 305 310	
55	CTT GTT ACA CTT TTG AAT AAA GAT TTC CCC GAG TTA GGG TTG CAA GAA	1010
	Leu Val Thr Leu Leu Asn Lys Asp Phe Pro Glu Leu Gly Leu Gln Glu	
	315 320 325 330	

TCG GAT TGT ACT GAA ATG AGT TGG GTT GAG TCT GTG CTT TAC TAC ACG 1058
 Ser Asp Cys Thr Glu Met Ser Trp Val Glu Ser Val Leu Tyr Tyr Thr
 335 340 345

5 GGC TTC CCC AGT GGT ACT CCA ACC ACG GCG CTC TTA AGC CGT ACT CCT 1106
 Gly Phe Pro Ser Gly Thr Pro Thr Thr Ala Leu Leu Ser Arg Thr Pro
 350 355 360

10 CAA AGA CTC AAC CCA TTC AAG ATC AAA TCC GAT TAT GTG CAA AAT CCT 1154
 Gln Arg Leu Asn Pro Phe Lys Ile Lys Ser Asp Tyr Val Gln Asn Pro
 365 370 375

15 ATT TCT AAA CGA CAG TTC GAG TTC ATC TTC GAA AGG CTG AAA GAA CTT 1202
 Ile Ser Lys Arg Gln Phe Glu Phe Ile Phe Glu Arg Leu Lys Glu Leu
 380 385 390

20 GAA AAC CAA ATG TTG GCT TTC AAC CCA TAT GGT GGT AGA ATG AGT GAA 1250
 Glu Asn Gln Met Leu Ala Phe Asn Pro Tyr Gly Gly Arg Met Ser Glu
 395 400 405 410

ATA TCC GAA TTC GCA AAG CCT TTC CCA CAT AGA TCG GGT AAC ATA GCG 1298
 Ile Ser Glu Phe Ala Lys Pro Phe Pro His Arg Ser Gly Asn Ile Ala
 415 420 425

25 AAA ATT CAA TAC GAA GTA AAC TGG GAG GAT CTT AGC GAT GAA GCC GAA 1346
 Lys Ile Gln Tyr Glu Val Asn Trp Glu Asp Leu Ser Asp Glu Ala Glu
 430 435 440

30 AAT CGT TAC TTG AAT TTC ACA AGG CTG ATG TAT GAT TAC ATG ACC CCA 1394
 Asn Arg Tyr Leu Asn Phe Thr Arg Leu Met Tyr Asp Tyr Met Thr Pro
 445 450 455

35 TTT GTG TCG AAA AAC CCT AGA AAA GCA TTT TTG AAC TAT AGG GAT TTG 1442
 Phe Val Ser Lys Asn Pro Arg Lys Ala Phe Leu Asn Tyr Arg Asp Leu
 460 465 470

40 GAT ATT GGT ATC AAC AGC CAT GGC AGG AAT GCT TAT ACT GAA GGA ATG 1490
 Asp Ile Gly Ile Asn Ser His Gly Arg Asn Ala Tyr Thr Glu Gly Met
 475 480 485 490

GTT TAT GGG CAC AAG TAT TTC AAA GAG ACA AAT TAC AAG AGG CTA GTA 1538
 Val Tyr Gly His Lys Tyr Phe Lys Glu Thr Asn Tyr Lys Arg Leu Val
 495 500 505

45 AGT GTG AAG ACT AAA GTT GAT CCT GAC AAC TTC TTT AGG AAT GAG CAA 1586
 Ser Val Lys Thr Lys Val Asp Pro Asp Asn Phe Phe Arg Asn Glu Gln
 510 515 520

50 AGC ATC CCA ACT TTG TCA TCT T GAAGAACGTA CATATATAAA TAAATACCTT 1638
 Ser Ile Pro Thr Leu Ser Ser
 525

TGTGCATGGT ATTTTCAGGG TGTTAAAGTG ATATTCAGAT ATTTATGATA GAATTTTGAC 1698

55 TTGTATTTTA TACAATCAAA ATTGTATGGT TCTCCGAATT TCTCTTTTTA ATTCTGAAAA 1758

ATACATATTA GTATTGTCAA AAAAAA

1784

(2) INFORMATION FOR SEQ ID NO: 16:

5

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 529 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

10

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

```

15 Met Gln Thr Ser Ile Leu Thr Leu Leu Leu Leu Leu Ser Thr Gln
    1           5           10           15

    Ser Ser Ala Thr Ser Arg Ser Ile Thr Asp Arg Phe Ile Gln Cys Leu
          20           25           30

20 His Asp Arg Ala Asp Pro Ser Phe Pro Ile Thr Gly Glu Val Tyr Thr
    35           40           45

    Pro Gly Asn Ser Ser Phe Pro Thr Val Leu Gln Asn Tyr Ile Arg Asn
25     50           55           60

    Leu Arg Phe Asn Glu Thr Thr Thr Pro Lys Pro Phe Leu Ile Ile Thr
    65           70           75           80

30 Ala Glu His Val Ser His Ile Gln Ala Ala Val Val Cys Gly Lys Gln
          85           90           95

    Asn Arg Leu Leu Leu Lys Thr Arg Ser Gly Gly His Asp Tyr Glu Gly
          100          105          110

35 Leu Ser Tyr Leu Thr Asn Thr Asn Gln Pro Phe Phe Ile Val Asp Met
    115          120          125

    Phe Asn Leu Arg Ser Ile Asn Val Asp Ile Glu Gln Glu Thr Ala Trp
40     130          135          140

    Val Gln Ala Gly Ala Thr Leu Gly Glu Val Tyr Tyr Arg Ile Ala Glu
    145          150          155          160

45 Lys Ser Asn Lys His Gly Phe Pro Ala Gly Val Cys Pro Thr Val Gly
    165          170          175

    Val Gly Gly His Phe Ser Gly Gly Gly Tyr Gly Asn Leu Met Arg Lys
          180          185          190

50 Tyr Gly Leu Ser Val Asp Asn Ile Val Asp Ala Gln Ile Ile Asp Val
    195          200          205

    Asn Gly Lys Leu Leu Asp Arg Lys Ser Met Gly Glu Asp Leu Phe Trp
55     210          215          220

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Ala Ile Thr Gly Gly Gly Gly Val Ser Phe Gly Val Val Leu Ala Tyr
 225 230 235 240
 Lys Ile Lys Leu Val Arg Val Pro Glu Val Val Thr Val Phe Thr Ile
 5 245 250 255
 Glu Arg Arg Glu Glu Gln Asn Leu Ser Thr Ile Ala Glu Arg Trp Val
 260 265 270
 10 Gln Val Ala Asp Lys Leu Asp Arg Asp Leu Phe Leu Arg Met Thr Phe
 275 280 285
 Ser Val Ile Asn Asp Thr Asn Gly Gly Lys Thr Val Arg Ala Ile Phe
 290 295 300
 15 Pro Thr Leu Tyr Leu Gly Asn Ser Arg Asn Leu Val Thr Leu Leu Asn
 305 310 315 320
 Lys Asp Phe Pro Glu Leu Gly Leu Gln Glu Ser Asp Cys Thr Glu Met
 20 325 330 335
 Ser Trp Val Glu Ser Val Leu Tyr Tyr Thr Gly Phe Pro Ser Gly Thr
 340 345 350
 25 Pro Thr Thr Ala Leu Leu Ser Arg Thr Pro Gln Arg Leu Asn Pro Phe
 355 360 365
 Lys Ile Lys Ser Asp Tyr Val Gln Asn Pro Ile Ser Lys Arg Gln Phe
 370 375 380
 30 Glu Phe Ile Phe Glu Arg Leu Lys Glu Leu Glu Asn Gln Met Leu Ala
 385 390 395 400
 Phe Asn Pro Tyr Gly Gly Arg Met Ser Glu Ile Ser Glu Phe Ala Lys
 35 405 410 415
 Pro Phe Pro His Arg Ser Gly Asn Ile Ala Lys Ile Gln Tyr Glu Val
 420 425 430
 40 ~~Asn Trp Glu Asp Leu Ser Asp Glu Ala Glu Asn Arg Tyr Leu Asn Phe~~
 435 440 445
 Thr Arg Leu Met Tyr Asp Tyr Met Thr Pro Phe Val Ser Lys Asn Pro
 450 455 460
 45 Arg Lys Ala Phe Leu Asn Tyr Arg Asp Leu Asp Ile Gly Ile Asn Ser
 465 470 475 480
 His Gly Arg Asn Ala Tyr Thr Glu Gly Met Val Tyr Gly His Lys Tyr
 50 485 490 495
 Phe Lys Glu Thr Asn Tyr Lys Arg Leu Val Ser Val Lys Thr Lys Val
 500 505 510

55

Asp Pro Asp Asn Phe Phe Arg Asn Glu Gln Ser Ile Pro Thr Leu Ser
515 520 525

Ser

5

(2) INFORMATION FOR SEQ ID NO: 17:

(i) SEQUENCE CHARACTERISTICS:

- 10 (A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

15

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

CCGCCATGGA GACTTCCATT CTTACTC

27

(2) INFORMATION FOR SEQ ID NO: 18:

25

(i) SEQUENCE CHARACTERISTICS:

- 30 (A) LENGTH: 33 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

35

(iii) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

40 GCCGGATCCT CAAGATGACA AAGTTGGGAT GCT

33

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS:

- 45 (A) LENGTH: 1590 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

55

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Helianthus annuus*

(B) STRAIN: Zebulon

5 (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..1590

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

15	ATG GAG ACT TCC ATT CTT ACT CTC CTT CTT CTC TTG CTC TCA ACC CAA	48
	Met Glu Thr Ser Ile Leu Thr Leu Leu Leu Leu Leu Ser Thr Gln	
	1 5 10 15	
15	TCT TCT GCA ACT TCC CGT TCC ATT ACA GAT CGC TTC ATT CAA TGT TTA	96
	Ser Ser Ala Thr Ser Arg Ser Ile Thr Asp Arg Phe Ile Gln Cys Leu	
	20 25 30	
20	CAC GAC CGG GCC GAC CCT TCA TTT CCG ATA ACC GGA GAG GTT TAC ACT	144
	His Asp Arg Ala Asp Pro Ser Phe Pro Ile Thr Gly Glu Val Tyr Thr	
	35 40 45	
25	CCC GGA AAC TCA TCT TTT CCT ACC GTC TTG CAA AAC TAC ATC CGA AAC	192
	Pro Gly Asn Ser Ser Phe Pro Thr Val Leu Gln Asn Tyr Ile Arg Asn	
	50 55 60	
30	CTT CGG TTC AAT GAA ACT ACC ACA CCA AAA CCC TTT TTA ATC ATC ACA	240
	Leu Arg Phe Asn Glu Thr Thr Thr Pro Lys Pro Phe Leu Ile Ile Thr	
	65 70 75 80	
35	GCC GAA CAT GTT TCC CAC ATT CAG GCA GCT GTG GTT TGT GGC AAA CAA	288
	Ala Glu His Val Ser His Ile Gln Ala Ala Val Val Cys Gly Lys Gln	
	85 90 95	
40	AAC CGG TTG CTA CTG AAA ACC AGA AGC GGT GGT CAT GAT TAT GAA GGT	336
	Asn Arg Leu Leu Leu Lys Thr Arg Ser Gly Gly His Asp Tyr Glu Gly	
	100 105 110	
40	CTT TCC TAC CTT ACA AAC ACA AAC CAA CCC TTC TTC ATT GTG GAC ATG	384
	Leu Ser Tyr Leu Thr Asn Thr Asn Gln Pro Phe Phe Ile Val Asp Met	
	115 120 125	
45	TTC AAT TTA AGG TCC ATA AAC GTA GAT ATC GAA CAA GAA ACC GCA TGG	432
	Phe Asn Leu Arg Ser Ile Asn Val Asp Ile Glu Gln Glu Thr Ala Trp	
	130 135 140	
50	GTC CAA GCC GGT GCG ACT CTT GGT GAA GTG TAC TAT CGA ATA GCG GAG	480
	Val Gln Ala Gly Ala Thr Leu Gly Glu Val Tyr Tyr Arg Ile Ala Glu	
	145 150 155 160	
55	AAA AGT AAC AAG CAT GGT TTT CCG GCA GGG GTT TGT CCA ACG GTT GGC	528
	Lys Ser Asn Lys His Gly Phe Pro Ala Gly Val Cys Pro Thr Val Gly	
	165 170 175	

	GTT	GGT	GGG	CAT	TTT	AGT	GGT	GGT	GGG	TAT	GGT	AAT	TTG	ATG	AGA	AAA	576
	Val	Gly	Gly	His	Phe	Ser	Gly	Gly	Gly	Tyr	Gly	Asn	Leu	Met	Arg	Lys	
				180					185							190	
5	TAT	GGT	TTG	TCG	GTT	GAT	AAT	ATT	GTT	GAT	GCT	CAA	ATA	ATA	GAT	GTG	624
	Tyr	Gly	Leu	Ser	Val	Asp	Asn	Ile	Val	Asp	Ala	Gln	Ile	Ile	Asp	Val	
			195					200					205				
	AAT	GGC	AAG	CTT	TTG	GAT	CGA	AAG	AGT	ATG	GGT	GAG	GAT	TTG	TTT	TGG	672
10	Asn	Gly	Lys	Leu	Leu	Asp	Arg	Lys	Ser	Met	Gly	Glu	Asp	Leu	Phe	Trp	
		210					215					220					
	GCG	ATC	ACC	GGC	GGT	GGT	GGT	GTT	AGT	TTT	GGT	GTG	GTT	CTA	GCC	TAC	720
15	Ala	Ile	Thr	Gly	Gly	Gly	Gly	Val	Ser	Phe	Gly	Val	Val	Leu	Ala	Tyr	
	225				230						235					240	
	AAA	ATC	AAA	CTA	GTT	CGT	GTT	CCG	GAG	GTT	GTG	ACC	GTG	TTT	ACC	ATT	768
	Lys	Ile	Lys	Leu	Val	Arg	Val	Pro	Glu	Val	Val	Thr	Val	Phe	Thr	Ile	
				245					250					255			
20	GAA	AGA	AGA	GAG	GAA	CAA	AAC	CTC	AGC	ACC	ATC	GCG	GAA	CGA	TGG	GTA	816
	Glu	Arg	Arg	Glu	Glu	Gln	Asn	Leu	Ser	Thr	Ile	Ala	Glu	Arg	Trp	Val	
			260					265						270			
25	CAA	GTT	GCT	GAT	AAG	CTA	GAT	AGA	GAT	CTT	TTC	CTT	CGA	ATG	ACC	TTT	864
	Gln	Val	Ala	Asp	Lys	Leu	Asp	Arg	Asp	Leu	Phe	Leu	Arg	Met	Thr	Phe	
			275					280					285				
	AGT	GTC	ATA	AAC	GAT	ACC	AAC	GGT	GGA	AAG	ACA	GTC	CGT	GCT	ATC	TTT	912
30	Ser	Val	Ile	Asn	Asp	Thr	Asn	Gly	Gly	Lys	Thr	Val	Arg	Ala	Ile	Phe	
		290						295					300				
	CCA	ACG	TTG	TAC	CTT	GGA	AAC	TCG	AGG	AAT	CTT	GTT	ACA	CTT	TTG	AAT	960
	Pro	Thr	Leu	Tyr	Leu	Gly	Asn	Ser	Arg	Asn	Leu	Val	Thr	Leu	Leu	Asn	
35	305					310					315				320		
	AAA	GAT	TTC	CCC	GAG	TTA	GGG	TTG	CAA	GAA	TCG	GAT	TGT	ACT	GAA	ATG	1008
	Lys	Asp	Phe	Pro	Glu	Leu	Gly	Leu	Gln	Ser	Asp	Cys	Thr	Glu	Met		
				325					330					335			
40	AGT	TGG	GTT	GAG	TCT	GTG	CTT	TAC	TAC	ACG	GGC	TTC	CCC	AGT	GGT	ACT	1056
	Ser	Trp	Val	Glu	Ser	Val	Leu	Tyr	Tyr	Thr	Gly	Phe	Pro	Ser	Gly	Thr	
			340						345					350			
45	CCA	ACC	ACG	GCG	CTC	TTA	AGC	CGT	ACT	CCT	CAA	AGA	CTC	AAC	CCA	TTC	1104
	Pro	Thr	Thr	Ala	Leu	Leu	Ser	Arg	Thr	Pro	Gln	Arg	Leu	Asn	Pro	Phe	
			355					360					365				
	AAG	ATC	AAA	TCC	GAT	TAT	GTG	CAA	AAT	CCT	ATT	TCT	AAA	CGA	CAG	TTC	1152
50	Lys	Ile	Lys	Ser	Asp	Tyr	Val	Gln	Asn	Pro	Ile	Ser	Lys	Arg	Gln	Phe	
		370						375					380				
	GAG	TTC	ATC	TTC	GAA	AGG	ATG	AAA	GAA	CTT	GAA	AAC	CAA	ATG	TTG	GCG	1200
	Glu	Phe	Ile	Phe	Glu	Arg	Met	Lys	Glu	Leu	Glu	Asn	Gln	Met	Leu	Ala	
55	385					390					395					400	


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TTC AAC CCA TAT GGT GGT AGA ATG AGT GAA ATA TCC GAA TTC GCA AAG 1248
Phe Asn Pro Tyr Gly Gly Arg Met Ser Glu Ile Ser Glu Phe Ala Lys
                                405                                410                                415

5  CCT TTC CCA CAT AGA TCG GGT AAC ATA GCG AAG ATT CAA TAC GAA GTA 1296
   Pro Phe Pro His Arg Ser Gly Asn Ile Ala Lys Ile Gln Tyr Glu Val
                                420                                425                                430

AAC TGG GAG GAT CTT AGC GAT GAA GCC GAA AAT CGT TAC TTG AAT TTC 1344
10 Asn Trp Glu Asp Leu Ser Asp Glu Ala Glu Asn Arg Tyr Leu Asn Phe
                                435                                440                                445

ACA AGG CTG ATG TAT GAT TAC ATG ACT CCA TTT GTG TCG AAA AAC CCT 1392
15 Thr Arg Leu Met Tyr Asp Tyr Met Thr Pro Phe Val Ser Lys Asn Pro
                                450                                455                                460

AGA GAA GCA TTT TTG AAC TAT AGG GAT TTG GAT ATT GGT ATC AAC AGC 1440
Arg Glu Ala Phe Leu Asn Tyr Arg Asp Leu Asp Ile Gly Ile Asn Ser
465                                470                                475                                480

20 CAT GGC AGG AAT GCT TAT ACT GAA GGA ATG GTT TAT GGG CAC AAA TAT 1488
   His Gly Arg Asn Ala Tyr Thr Glu Gly Met Val Tyr Gly His Lys Tyr
                                485                                490                                495

TTC AAA GAG ACA AAT TAC AAG AGG CTA GTA AGT GTG AAG ACT AAA GTT 1536
25 Phe Lys Glu Thr Asn Tyr Lys Arg Leu Val Ser Val Lys Thr Lys Val
                                500                                505                                510

GAT CCT GAC AAC TTC TTT AGG AAT GAG CAA AGC ATC CCA ACT TTG TCA 1584
30 Asp Pro Asp Asn Phe Phe Arg Asn Glu Gln Ser Ile Pro Thr Leu Ser
                                515                                520                                525

TCT TG
Ser
35 530

```

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:

40 (A) LENGTH: 529 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

```

Met Glu Thr Ser Ile Leu Thr Leu Leu Leu Leu Leu Ser Thr Gln
 1              5              10              15

50 Ser Ser Ala Thr Ser Arg Ser Ile Thr Asp Arg Phe Ile Gln Cys Leu
    20              25              30

His Asp Arg Ala Asp Pro Ser Phe Pro Ile Thr Gly Glu Val Tyr Thr
55 35              40              45

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Pro Gly Asn Ser Ser Phe Pro Thr Val Leu Gln Asn Tyr Ile Arg Asn
 50 55 60

5 Leu Arg Phe Asn Glu Thr Thr Thr Pro Lys Pro Phe Leu Ile Ile Thr
 65 70 75 80

Ala Glu His Val Ser His Ile Gln Ala Ala Val Val Cys Gly Lys Gln
 85 90 95

10 Asn Arg Leu Leu Leu Lys Thr Arg Ser Gly Gly His Asp Tyr Glu Gly
 100 105 110

15 Leu Ser Tyr Leu Thr Asn Thr Asn Gln Pro Phe Phe Ile Val Asp Met
 115 120 125

Phe Asn Leu Arg Ser Ile Asn Val Asp Ile Glu Gln Glu Thr Ala Trp
 130 135 140

20 Val Gln Ala Gly Ala Thr Leu Gly Glu Val Tyr Tyr Arg Ile Ala Glu
 145 150 155 160

Lys Ser Asn Lys His Gly Phe Pro Ala Gly Val Cys Pro Thr Val Gly
 165 170 175

25 Val Gly Gly His Phe Ser Gly Gly Gly Tyr Gly Asn Leu Met Arg Lys
 180 185 190

Tyr Gly Leu Ser Val Asp Asn Ile Val Asp Ala Gln Ile Ile Asp Val
 195 200 205

30 Asn Gly Lys Leu Leu Asp Arg Lys Ser Met Gly Glu Asp Leu Phe Trp
 210 215 220

35 Ala Ile Thr Gly Gly Gly Gly Val Ser Phe Gly Val Val Leu Ala Tyr
 225 230 235 240

Lys Ile Lys Leu Val Arg Val Pro Glu Val Val Thr Val Phe Thr Ile
 245 250 255

40 Glu Arg Arg Glu Glu Gln Asn Leu Ser Thr Ile Ala Glu Arg Trp Val
 260 265 270

45 Gln Val Ala Asp Lys Leu Asp Arg Asp Leu Phe Leu Arg Met Thr Phe
 275 280 285

Ser Val Ile Asn Asp Thr Asn Gly Gly Lys Thr Val Arg Ala Ile Phe
 290 295 300

50 Pro Thr Leu Tyr Leu Gly Asn Ser Arg Asn Leu Val Thr Leu Leu Asn
 305 310 315 320

Lys Asp Phe Pro Glu Leu Gly Leu Gln Glu Ser Asp Cys Thr Glu Met
 325 330 335

55

Ser Trp Val Glu Ser Val Leu Tyr Tyr Thr Gly Phe Pro Ser Gly Thr
 340 345 350
 Pro Thr Thr Ala Leu Leu Ser Arg Thr Pro Gln Arg Leu Asn Pro Phe
 5 355 360 365
 Lys Ile Lys Ser Asp Tyr Val Gln Asn Pro Ile Ser Lys Arg Gln Phe
 370 375 380
 10 Glu Phe Ile Phe Glu Arg Met Lys Glu Leu Glu Asn Gln Met Leu Ala
 385 390 395 400
 Phe Asn Pro Tyr Gly Gly Arg Met Ser Glu Ile Ser Glu Phe Ala Lys
 405 410 415
 15 Pro Phe Pro His Arg Ser Gly Asn Ile Ala Lys Ile Gln Tyr Glu Val
 420 425 430
 Asn Trp Glu Asp Leu Ser Asp Glu Ala Glu Asn Arg Tyr Leu Asn Phe
 20 435 440 445
 Thr Arg Leu Met Tyr Asp Tyr Met Thr Pro Phe Val Ser Lys Asn Pro
 450 455 460
 25 Arg Glu Ala Phe Leu Asn Tyr Arg Asp Leu Asp Ile Gly Ile Asn Ser
 465 470 475 480
 His Gly Arg Asn Ala Tyr Thr Glu Gly Met Val Tyr Gly His Lys Tyr
 485 490 495
 30 Phe Lys Glu Thr Asn Tyr Lys Arg Leu Val Ser Val Lys Thr Lys Val
 500 505 510
 Asp Pro Asp Asn Phe Phe Arg Asn Glu Gln Ser Ile Pro Thr Leu Ser
 35 515 520 525
 Ser

(2) INFORMATION FOR SEQ ID NO: 21:

40

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 350 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

45

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

50

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Arabidopsis thaliana*
 (B) STRAIN: ecotype Columbia

55

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 2..350

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

GAGAAACTCG GAGACTTTCA CACAATGCCT AACCTCAAAC TCCGACCCCA AACATCCCAT 60
CTCCCCCGCT ATCTTCTTCT CCGGAAATGG CTCCTACTCC TCCGTATTAC AAGCCAACAT 120
10 CCGTAACCTC CGCTTCAACA CCACCTCAAC TCCGAAACCC TTCCTCATAA TCGCCGCAAC 180
ACATGAATCC CATGTGCAAG CCGCGATTAC TTGCGGGAAA CGCCACAACC TTCAGATGAA 240
15 AATCAGAAGT GGAGGCCACG ACTACGATGG CTTGTCATAC GTTACATACT CTGGCAAACC 300
GTTCTTCGTC CTCGACATGT TTAACCTCCG TTCGGTGGAT GTCGACGTGG 350

(2) INFORMATION FOR SEQ ID NO: 22:

20

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 278 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

30

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Arabidopsis thaliana
- (B) STRAIN: ecotype Columbia

35

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 2..278

40

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

GGCATGGATC TCCGCCGGAG CGACTCTCGG AGAGGTTTAT TATCGGATTT GGGAGAAAAG 60
45 CAGAGTCCAT GGATTCCCCG CCGGAGTTTG ACCGACGGTT GGTGTTGGTG GGCATTTAAG 120
CGGCGGTGGT TACGGTAACA TGGTGAGGAA GTTTGGATTA TCTGTGGATT ACGTTGAGGA 180
50 TGCCAAGATC GTCGATGTAA ACNGTCGGGT TTTAGATCGG AAAGCAATGG GTGAGGATCT 240
GTTCTGGGCG ATTACCGGTG GAGGAGGAGG TAGCGTAC 278

(2) INFORMATION FOR SEQ ID NO: 23:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 345 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

10

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

15

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Arabidopsis thaliana
(B) STRAIN: ecotype Columbia

(ix) FEATURE:

20

- (A) NAME/KEY: CDS
(B) LOCATION: 2..345

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

25 TGGACATATT AGCGGAGGAG GATTCGGTAC AATAATGAGG AAATACGGTT TAGCGTCTGA 60
TAACGTTGTG GACGCACGTT TGATGGATGT AAATGGGAAA ACTCTTGACC GGAAAACGAT 120
GGGAGAGGAT TTGTTTGGG CGCTTAGAGG CGGTGGAGCT GCGAGTTTGG GCGTTGTCTT 180
30 GTCGTGGAAG GTTAAGCTTG CTAGGGTTCC TGAAAAGGTA ACTTGTTTCA TAAGTCAACA 240
TCCGATGGGA CCTAGCATGA ACAAGCTTGT TCATAGATGG CAATCCATAG GATCAAGANN 300
35 GCTAGACGAA GATTTATTCA TCAGAGTCAA TATTGACAAC AGTCT 345

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

- 40 (A) LENGTH: 695 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

45

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

50

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Arabidopsis thaliana
(B) STRAIN: ecotype Columbia

55

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..695

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

GTTCGTAAAA ACCTATCCTN NANGGGCNAA AGNATATCAA AGNTTGNTTA NGNAACCCAA 60
NATTTCTGAA CTGGCCNCCT TCGGTGGTAT ATGNCNAAAN CCCTTGAATC TGCGNANCCN 120
10 ATTCCGCATA GAAACGGAAC CCTCTTCAAG ATTCTCTATT TACNCGAACT GNCTAGANNG 180
AATGACAAGA CATCGAGTAG NAAAATCAAC TGGATCAAAG AGATATACAA TTACATGGCG 240
15 CCTTATGTCT CAAGCAATCC AAGACAAGCA TATGTGAACT ACAGAGATCT AGACTTCGGA 300
CAGAACAAGA ACAACGCAAA GGTAACTTC ATTGAAGCTA AAATCTGGGG ACCTAAGTAC 360
TTCAAAGGCA ATTTTGACAG ATTGGTGAAG ATTA AAAACCA AGGTTGATCC AGAGAACTTC 420
20 TTCAGGCACG AGCAGAGTAT CCCACCTATG CCCTACTAGA AGCTAGGTTT ATGAAACCAA 480
TAACATTATC AAAAATAAGR ATAAATGRTA ATTGTATACA ACATGATTCG KCTTTCTTTA 540
25 TTTTCAGACAA TGTGGACACT ACTCTAAANT AAAAWGTCNA TTTACCTTAA AAAAAAATA 600
ATCCCCNNTA ANANAAAANT GGGGGGGCCN TTTTGGGGN TCCCGGTTTT NGGACGGGGN 660
GCTTTNGGGG GGCTTGGNNT TTTTNGGN GCCCC 695
30

(2) INFORMATION FOR SEQ ID NO: 25:

(i) SEQUENCE CHARACTERISTICS:

35 (A) LENGTH: 495 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

40

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

45 (vi) ORIGINAL SOURCE:

(A) ORGANISM: Arabidopsis thaliana
(B) STRAIN: ecotype Columbia

(ix) FEATURE:

50 (A) NAME/KEY: CDS
(B) LOCATION: 2..495

55

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 25:

TCTGTTTTNA GGCAGAGCAG AGGAAGTTGT TGCTTTGCTT GGTAAGGAGT TTCCTGAATT 60
 5 NAGTTTAAAG AAGGAGAACT GTTCGGAGAT GACTTGTTT CAGTCAGCTT TATGGTGGGA 120
 TAATCGTGTT AACCCCTACTC ANATTGATCC WAAAGTGTTT CTCGATCGGA ATCTTGATAG 180
 AGCGAATTTT GGAAAGAGGA AATCGGATTA CGTTGCGAGT AAGATTCCTA GAGATGGGAT 240
 10 TAAGYCTTTT TCCAAGARGA TGMCTGACCT GGGGAAAAYC GGGCTTGTTT TTAAWCCGTA 300
 TGGTGGGAAA ATGGCGGAGG TTACGGTTAA CGCGACGCCG TTCCNCACC GAAGCAAGCT 360
 15 TTTTAAGATT CAGTACTCGG TGACTIONGCA AGAAACTCT NTCGAGATAG AGAAAGGGT 420
 TCTTGAATCA GGCTAACGTC CTTATAGGTT CATGACCGGG TTTTNAGCA AGANCCCTGG 480
 AATNCTTACT TNAAT 495

20

(2) INFORMATION FOR SEQ ID NO: 26:

(i) SEQUENCE CHARACTERISTICS:

- 25 (A) LENGTH: 204 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

30

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

35

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Arabidopsis thaliana*
 (B) STRAIN: ecotype Columbia

(ix) FEATURE:

40

- (A) NAME/KEY: CDS
 (B) LOCATION: 1..204

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:

45 AAATTAAAAC AAATCAATGT TGATATTGAA TCCAATAGTG CTTGGTTTCA ACCTGGTGCT 60
 ACGCTTGGTG AGCTTTACTA CAGAATTNCA GAGAAGAGCA AAATCCATGG ATTTCCNGCG 120
 GGTNTNTNCA CAAGCNTAGG CATAGGTGGG TATATNANAG GCGGTGGATA CGGTACCTTG 180
 50 ATGAGGAAGT ATGGTCTTNC GGGA 204

(2) INFORMATION FOR SEQ ID NO: 27:

55

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 491 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

5

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

10

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Arabidopsis thaliana

(B) STRAIN: ecotype Columbia

15

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 2..491

20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

GAGATTTCTC GAGCAAGATA CTCCACTGAT GATCTTTGAG CCATTGGGTG GGAAAATCAG 60
 CAAGATTTCA GAAACAGAAT CTCCATATCC ACACAGAAGA GGTAATCTGT ATAATATACA 120
 25 GTACATGGTG AAATGGAAAG TGAATGANGT CGAGGAGATG AACAAACATG TCAGGTGGAT 180
 GAGATCGTTA CACGATTACA TGA TCTCCGTA TGTTTCTAAA TCGCCGAGAG GAGCTTATTT 240
 30 GANTTACAGA GATCTTGATT TGGGCTCGAC CAAAGGGATT AACACGGGTT TCGGAGATGC 300
 AAGGAAATGG NNGGGTGAGN CTTTTTTTCAA AGGTAATTTT CAAGGGGTTA GGTTTTGGTT 360
 AAAGGGGAGG TTTNNCCCAN CAAATTTTTT TTCAGGANCC GGCCANGNTT TTCCCCCCCC 420
 35 TTTTTTTTNGG NCCCCAATCN AAANCCCCGT TTTAAAAGGG GGGCCATTTC NTTTTTTNCA 480
 NNTTAAAAGG G 491

40

(2) INFORMATION FOR SEQ ID NO: 28:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 407 base pairs

(B) TYPE: nucleic acid

45

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

50

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

55

(A) ORGANISM: Arabidopsis thaliana

(B) STRAIN: ecotype Columbia

(ix) FEATURE:

(A) NAME/KEY: CDS

5 (B) LOCATION: 3..407

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:

10 ATTTGTTTCGT GAGGTTAACT TTGACTTTAG TCAACGGTAC GAAGCCTGGT GAGAATACGG 60
 TTTTAGCGAC TTTCATTGGG ATGTATTTAG GCCGGTCGGA TAAGCTGTTG ACCGTNATGA 120
 ACCGGGATTT CCCGGAGTTG AAGCTGAAGA AAACCGATTN TACCGAGATG AGATGGATCG 180
 15 ATTCGGTTCT GTTTTGGGAC GATTATCCGG TTGGTACACC GACTTCTGTG CTAATAAATC 240
 CGCTAGTCGC AAAAAAGTTG TTCATGAAAC GAAAATCGGA CTACGTGAAG CGTCTNATTT 300
 TCGAGAACCC GATCTCNNGT TTGATACTCA AGAAATTTGT AGAGGTTNNG AAAGTTAAAA 360
 20 TNAATTTGGA TCCGCATTNN GGNANNNATG GTGAAACCCC NNGTTNT 407

(2) INFORMATION FOR SEQ ID NO: 29:

25 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 360 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

30

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

35 (iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Arabidopsis thaliana

(B) STRAIN: ecotype Columbia

40

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 3..360

45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

ACGGCGTCGT ATTGGCCTAC AAAATAAAACC TTGTTGAAGT CCCAGAAAAC GTCACCGTTT 60
 TCAGAATCTC CCGGACGTTA GAACAAAATG CGACGGATAT CATTACCGG TGGCAACAAG 120
 50 TTGCACCGAA GCTTCCCGAC GAGCTTTTCA TAAGANCAGT CATTGACGTA NAAACGGCAC 180
 TGTTCATNN CTCAAAGAC CGTCAGACAA CATTATAGC AATGTTTCTA GGAGACACGN 240
 55 CAACTCTACT GTCGATATTA AACCGGAGAT TCCAGAATT GGGTTTGGTC CGGTCTGACT 300

GTACCGNAAC AAGCNNTTGG ATCCAATCTG TGCTATTTT GGGACAAATA TCCCAGGTTG 360

(2) INFORMATION FOR SEQ ID NO: 30:

- 5 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 427 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- 10 (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- 15 (iii) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: Arabidopsis thaliana
 (B) STRAIN: ecotype Columbia
- 20 (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 3..427
- 25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:

TCTTCACTGT CACCAAAACG TTAGAACAAG ACGCAAGATT GAAGACTATT TCTAAGTGGC 60
AACAAATTTT ATCCAAGATT ATTGAAGAGA TACACATCCG AGTGGTACTC AGAGCAGCTG 120
30 GAAATGATGG AAACAAGACT GTGACAATGA CCTACCTAGG TCAGTTTCTT GGCGAGAAAG 180
GCACCTTGCT GAAGGTTATG GAGAAGGCTT TTCCAGAACT AGGGTTAACT CAAAAGGATT 240
35 GTACTGAAAT GAGCTGGATT GAAGCCGCCC TTTTCCATGG TGGRTTTCCT ACAGGKTCTC 300
CTATTGAAAT TTTGCTTMAG CTCAAGTCGC CTYTAGGAAA AGRTTWCTTC AAAGCAACGK 360
CGGATTTCGT TAAAGAACCT WTTCTGTGA TAGGGCTCAA AGGAATATTC AAAAGATTGA 420
40 TTGAAGG 427

(2) INFORMATION FOR SEQ ID NO: 31:

- 45 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 437 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- 50 (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- 55 (iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Arabidopsis thaliana*
 (B) STRAIN: ecotype Columbia

5 (ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 1..437

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 31:

10 GTTGTACTAT CATNGAAGAT TAAGTTAGTC GATGTTCCGT CCACGGTCAC CGNGTTTAAA 60
 GTCCAGAAAC ATNAGGAGAA AGAGGCCGTT AGGNTCATCA ACAAGTGGCA GTATGTTGCG 120
 15 GATAAGGTCC CTGAAGATCT TTTCATCAGC GCAACGTTGG NGAGATCAAA CGGAAACTCT 180
 GTGCAGGCTT TGTTTACTGG ACTCTATCTT GGNCCGGTGA ATAATNTCTT GGCCTTGATG 240
 GAAGAAAAGT TTCCAGANTT AGGTCTTGAT ATCCAAGNCT GCACAGAGAT GAGTTGGGCT 300
 20 GAATCTGCAC TCTGGTNTNC TGNTTTCNCT AAAGGAGAGN CTCCTTGGGT GTTCNCGCG 360
 GATCGGNAGC GGNCAATTTN TGGNCTTTCA AGGGGAAAGN CGGCTTTTTN CAAGAACCCG 420
 25 NTACCCGGGG TTCAATT 437

(2) INFORMATION FOR SEQ ID NO: 32:

(i) SEQUENCE CHARACTERISTICS:

- 30 (A) LENGTH: 441 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

40

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Arabidopsis thaliana*

(ix) FEATURE:

- 45 (A) NAME/KEY: CDS
 (B) LOCATION: 1..441

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 32:

50 GCGGACCCTA TAGATCANNA TGTGCTACTG ANAGAAGAGG AAGCCAAGAA CAAGCCGGAG 60
 ACAGATAAAT ATCTGAAATG GGNCGATANC GTTACGAAT TTATGACNCC ATATGTTTCG 120
 AAATCTCCAA GAGGAGCTTA TGTCAATTTT AAGGATATGG ATTTGGGTAT GTATCTTGGA 180
 55

AAGAAGAAGA CAAAGTACGA GGAAGGAAAG AGTTGGGGAG TGAAGTATTT CAAGAACAAT 240
 TTCGAGAGAT TGGTGAGAGT GAAGACTAGG GTTGATCCAA CAGATTTCTT CTGCGATGAA 300
 5 CAGAGCATTCT CTCTGGTGAA CAAAGTTACC TGAAGATATC ATTTGAAGTT TTTTATTAGT 360
 CCCTTTTCTC TGTGAAATCA TCTGTGCGTG TTGAATATTA TCGTCAAGT GTGTAACCTA 420
 TGTGTGTGAT TGTGAATTGT G 441

10

(2) INFORMATION FOR SEQ ID NO: 33:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 502 base pairs
 15 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
 20 (iii) HYPOTHETICAL: NO
 (iii) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 25 (A) ORGANISM: Arabidopsis thaliana
 (B) STRAIN: ecotype Columbia
- (ix) FEATURE:
 30 (A) NAME/KEY: CDS
 (B) LOCATION: 2..502

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 33:

35 CTGGCTTAAC ACAACGTCGT TTTGGGCCAA TTACCCGGCG GGTACACCCA AGAGCATCCT 60
 TCTAGATAGG CCTCCGACGA ATTCAGTGTC ATTTAAGAGT AAATCGGATT TTGTCAAAAA 120
 ACCAATACCC AAAAAAGGTT TAGAGAAGCT TTGGAAGACA ATGTTTAAAT TCAACAGTAG 180
 40 CGTCTCGTTG CAATTCAACC CTTACGGTGG AGTGATGGAC CGGATTCCGG CAACGGCCAC 240
 CGCTTTTCCT CATCGGAAAG GAAACTTGTT CAAGGTTCAA TACNCTACGA TGTGGTTTGA 300
 45 CGCAAACGCC ACACAGAGTA GCCNGGCTAT GATGAATGAG CTTTTTGAGG TGGCGGGACC 360
 GTACGTGNGT CAAGTAAACC CGAGANANGG CTTCTTTTAA NTTTCAGAGNC CATCGNTNTT 420
 NGGAGCAANN CCAAGTGGGG GGGNCCAACC GGGGGNTNAA ANCNNAGNTC TTNGGGGGCC 480
 50 CAGAATTTCC TTNGGGGAAT TT 502

(2) INFORMATION FOR SEQ ID NO: 34:

55

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 400 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Arabidopsis thaliana
(B) STRAIN: ecotype Columbia

(ix) FEATURE:

- (A) NAME/KEY: CDS
(B) LOCATION: 2..400

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 34:

NGGGAATTGC NCGAGGNAAG TTGTACCCAA TTCTGGACC ACCATTGGTT TCCCAAGAAN 60
CCCGAGACAA CCGTTTTTCA ATNACCGTGA TGTGATTTG GGTATTAATT CTCATAATGG 120
TAAAATCAGT AGTTATGTGG AAGGTAAACG TTACGGGAAG AAGTATTTTCG CAGGTAATTT 180
CGAGAGATTG GTGAAGATTA AGACGAGAGT TGATAGTGGT AATTTCTTTA GGAACGAACA 240
GAGTATTCCT GTGTTACCAT AAGTGTATTT ATTTGATTAT TGGTTAGTGA AATTTGTTGT 300
TGTATAATGA TTATATGTCG TATTTTTTATT TATTATTAGT AATTTATAAA GTTTGATATT 360
AAATACAAAT AGTATAATAA GATAGTTTCT TTTAGTAAAA 400

(2) INFORMATION FOR SEQ ID NO: 35:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 383 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Arabidopsis thaliana
(B) STRAIN: ecotype Columbia

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 2..383

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 35:

CAACTCTAAT GGAACACCT ACTTCGATCG AATGTCGATG GGGGAAGAGC TTTTCTGGGC 60
 GGTTCGAGGA GGTGGAGCCG CGAGTTTCGG CATCGTGATG GGATACAAA TCCGGTTGGT 120
 10 TCCGGTTCCG GAGAAAGTTA CGGTTTTTAG CGTCGGAAAA ACCGTCGGAG AAGGAGCCGT 180
 TGATCTTATA ATGAAGTGGC AGAACTTCTC TCATAGTACG GNTCGGAATT TTTTTGTGAA 240
 15 GCTGANTTTT GANTTTTAGTC AACGGTGCAA AGCCGGGTGA AAAAAAGGTT TTAGNGNCTT 300
 TCANTTTGGN TGNAANCTTG GGGGTTTTAT NAGAACGGTT AACCGGGATT NANCCCGNGT 360
 TTTCCCGGGG TAAAACCTT NGG 383

20

(2) INFORMATION FOR SEQ ID NO: 36:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 354 base pairs

25

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

30

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

35

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Arabidopsis thaliana

(B) STRAIN: ecotype Columbia

(ix) FEATURE:

40

(A) NAME/KEY: CDS

(B) LOCATION: 1..354

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 36:

45 ATCAATGTTT TACTAAACG TACACGAGCA TCGTTGGCTT TCAAGGCTAA ATCTGATTTT 60
 NTTCAAGAAC CGATNCCTAA AACCGCGATT TCGAAGCTTT GGAGACGGTT GCAAGAACCG 120
 GAAGCAGAGC ATGCTCAGCT AATTTCACN CCATTGGTG GTAAAATGAG TNAGATTGCA 180
 50 GATTACGAAA CACCATTTC GCATAGGAAG GGAATATAT ATNAGATTCA GTACTTGAAT 240
 TACTGGAGAG GAGACGTGAA AGAGAAGTAT ATTGAGATNG GTGGAGGAGA GTTTACGGTT 300
 55 GNTATNAGTA AGTTTTTTGG CGAAGTNTNC CNAGAGNGN CTTNNTNTAA ACCT 354

(2) INFORMATION FOR SEQ ID NO: 37:

- (i) SEQUENCE CHARACTERISTICS:
5 (A) LENGTH: 403 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
10 (iii) HYPOTHETICAL: NO
(iii) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
15 (A) ORGANISM: Arabidopsis thaliana
(B) STRAIN: ecotype Columbia
- (ix) FEATURE:
20 (A) NAME/KEY: CDS
(B) LOCATION: 2..403

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 37:

25 TTTTCTAGTA CACTAATAAT CAAATGGAAT GAGAAATGAA GCCACAAAAG TATCTGCAAT 60
CAAAATATCC TGCTATCTCC ATCTCAAGCT CTCAATAGTA TCCTCTCCGA AAGTGAAATC 120
AACATTTCAA ACTCTATTTT TGGTGGAAT CGATAGACTG ATTCCTCTGA TGAACCAGAA 180
30 GTTTCGGGAA CTCGGCTTAC GATCTCAAGA CTGTTCGGAA ATGAGCTGGA TCGAATCGAT 240
AATGTTCTTC AACTGGAGAT CAGGACAGCC GTTAGAGATT TTGCTCAACA GAGACCTAAG 300
35 GATTCGAGGA TCAGTATTTT AAAGCAAAGT CAGGATTATG GTTCAAAAAC CCGTTCCTGA 360
AAACGTTTTT CGAAGAGGTA TCCAAGGGGT TTCTCGAGCA AGT 403

(2) INFORMATION FOR SEQ ID NO: 38:

40

- (i) SEQUENCE CHARACTERISTICS:
45 (A) LENGTH: 260 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
50 (iii) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
55 (A) ORGANISM: Arabidopsis thaliana
(B) STRAIN: ecotype Columbia

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..260

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 38:

GAGATGAGTT GGATTAANTC TGTACTCTGG TTTGCTGATT TCCCTAAAGG AGAATCTCTT 60
NGTGTCTCA CGAATCGTAA GCGTACATCT CTATCTTTNA AAGGCAAAGA TGATTTTATC 120
10 CAAGAACCGA TACCCGAGGC TGCAATTNAA GAGATATGGA GCGGATTAGA AGCCCCCNAG 180
GCTCGGCTTG GAAAGATCAT ATTA ACTCCA TTTGGTGGGA AAATNAGTGA AATGGCAGAG 240
15 TACGTANCAC CATTCCCACA 260

(2) INFORMATION FOR SEQ ID NO: 39:

(i) SEQUENCE CHARACTERISTICS:

- 20 (A) LENGTH: 605 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

25 (ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

30

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Arabidopsis thaliana
- (B) STRAIN: ecotype Columbia

35

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 2..605

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 39:

40

CTCTTGATA TTCGCTGCAA GGATGGGAAA TTCAAACCA CTCCCTACAA TTTTTTGTAT 60
TATAGTTTCA GTCTTGATTT TTTAATTCTA TTGCATAACA CCAACTTCTT CATCAGCCTC 120
45 CATCCAAGAT CAATTCATAA ACTGTGTCAA AAGAAACACA CATGTTTCTT TTCCACTCGA 180
GAAAACGTTA TTCACCCCTG CGAAAAACGT CTCTTTGTTC AACCAAGTCC TTGANTCGAC 240
GGCTCAAAAT CTCCAGTTCT TGGCAAAATC CATGCCTAAA CCGGGRTTCA TATTCAGACC 300
50 GATTCACCAG TCTCAAGTCC AAGSTTCCAT CATTTGTTCA AMGRAACTCG GGNTTCATTT 360
TNGTGTTTGA NGTGGCGGTC ACGATTTTCG AGGCCTTTGT NTTTTATGTTT CACGGTTTGA 420
55 AAAAACCGTT TATATTACTC GGCCTGTCAA ANTTGNANNC AAAATCANAT GTTGGATATT 480

GNATTCCAAA TAGGTNCTTG GGGTNAACCT GGTGGCTANC GTTTGGTGAG CTTTTACTTT 540
 CAAGAATTTG CANGNGGANG TGCAAAGATT CCATGGGATT TCCCGGGGGG TTTNTTGCAC 600
 5 AATGT 605

(2) INFORMATION FOR SEQ ID NO: 40:

- 10 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 464 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- 15 (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- 20 (iii) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: *Arabidopsis thaliana*
 (B) STRAIN: ecotype Columbia
- 25 (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 2..464

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 40:

AACACAAAAC TCTTCCATTT GGCTTCTCTC TTGCATATTC GTTGCAAGGA TGGGAAATTC 60
 AAAACCACTC CCTACAATTN CTTGTATTAT CGTTTCAGTC TTGTATTTTN NATTCTATTG 120
 35 CATAACACCA ACTTCTTCAT CAGCCTCCAT CCAAGNTCAA TTCATAAACT GTGTCAAAAG 180
 GAACACACAT GTTTCTTTTC CACTCGAGNA AACGGTATTC ACTCCTGCGG AAAACGGCTC 240
 40 TTTTATTCAA CGGGTCCNTG AATCGACGGG TCAAAATCTC CAGTTCTTGG NAAAATCCAT 300
 GNCTAAACCG GGGTTCATAT TCAGGCCGGT TCACCAGTCT CAAGTCCAAG NTTCCATCAT 360
 TTGTTCAAAG GAACTCGGGA TTCATTTCCG CGNTAGAAGT GGCGGGCANN GGTTCGGGG 420
 45 CCTGTCTNTT GNTTANGGN AGGAAAACCG GTTNTATTNC TCGG 464

(2) INFORMATION FOR SEQ ID NO: 41:

- 50 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 386 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- 55

(ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

5 (iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Arabidopsis thaliana

(B) STRAIN: ecotype Columbia

10 (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..386

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 41:

TCGGGAGCCC ANGNTAAATT ANNTGAAAAT GGGGNCGNAT ANCCGTTTAC NGAATTTTAT 60

GACNCCCAAT ATGTTTCGAA ATCTCAAAGA NNGGGANCTT ATGTCAATTT CAAGGATATG 120

20 GATTGCGGTA TGTATCTTGG AAAGNAGAAG ACAAAGTACG AGGAAGGAAA GAGTTGGGGA 180

GTGAAGTATT TCAAGAACAA TTTCGAGAGA TTGGTGAGAG TGAAGACTAG GGTGATCCN 240

25 ACAGATTTTCN TCTGCGATGA ACAGAGCATT CCTCTGGTGN ACAAAGTTAC CTGAAGATAT 300

CATTTGAAGT TTTTATTAG TCCCTTTTCT CTGTGAAATC ATCTGTGCGT GTTGAATANT 360

ATGCGTCAAG TGTGTAACCTT ATGTGT 386

30 (2) INFORMATION FOR SEQ ID NO: 42:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 377 base pairs

35 (B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

40 (iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

45 (vi) ORIGINAL SOURCE:

(A) ORGANISM: Arabidopsis thaliana

(B) STRAIN: ecotype Columbia

(ix) FEATURE:

50 (A) NAME/KEY: CDS

(B) LOCATION: 1..377

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 42:

55 TACCATAGGG AGGTGGTGNA AGATTTTGTA TGTAGNCTTA GGGGAAGGCG AGTAGTATGG 60

TGGTGGTGGG GAGCTGTAAA CGTATGGTGG TGGTGGAGAT TTGTATGTGG GCTGGTTAAC 120
TTCATTGAAG CTAAAATCTG GGGACCTAAG TACTTCAAAG GCAATTTTGA CAGATTGGTG 180
5 AAGATTAAAA CCAAGGTTGA TCCAGAGAAC TTCTTCAGGC ACGAGCAGAG TATCCCACCT 240
ATGCCCTACT AGAAGCTAGG TTCATGAAAC CAATAACATT ATCAAAAATA AGAATAAATG 300
ATAATTGTAT ACAACATGAT TCGTCTTTCT TTATTTTCTAGA CAATGTGGAC ACTACTCTAA 360
10 ATAAAATGTC ATTTACC 377

(2) INFORMATION FOR SEQ ID NO: 43:

- 15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 377 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
20 (ii) MOLECULE TYPE: cDNA to mRNA
(iii) HYPOTHETICAL: NO
25 (iii) ANTI-SENSE: NO
(vi) ORIGINAL SOURCE:
(A) ORGANISM: Arabidopsis thaliana
(B) STRAIN: ecotype Columbia
30 (ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 1..377
35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 43:

TACCATAGGG AGGTGGTGNA AGATTTTGTG TGTAGNCTTA GGGGAAGGCG AGTAGTATGG 60
TGGTGGTGGG GAGCTGTAAA CGTATGGTGG TGGTGGAGAT TTGTATGTGG GCTGGTTAAC 120
40 TTCATTGAAG CTAAAATCTG GGGACCTAAG TACTTCAAAG GCAATTTTGA CAGATTGGTG 180
AAGATTAAAA CCAAGGTTGA TCCAGAGAAC TTCTTCAGGC ACGAGCAGAG TATCCCACCT 240
45 ATGCCCTACT AGAAGCTAGG TTCATGAAAC CAATAACATT ATCAAAAATA AGAATAAATG 300
ATAATTGTAT ACAACATGAT TCGTCTTTCT TTATTTTCTAGA CAATGTGGAC ACTACTCTAA 360
ATAAAAATGTC ATTTACC 377
50

(2) INFORMATION FOR SEQ ID NO: 44:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 346 base pairs
55 (B) TYPE: nucleic acid

(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

10 (vi) ORIGINAL SOURCE:

(A) ORGANISM: Arabidopsis thaliana

(B) STRAIN: ecotype Columbia

(ix) FEATURE:

15 (A) NAME/KEY: CDS

(B) LOCATION: 2..346

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 44:

20 GAGCTGTGGA TATGGTCACA AATGGCAATC GGTGGTCCG AAAACTGATC CGAATCTTTT 60
TATGAGAATN TTGATTCAAC CAGTGACGAG GAAGAAGGTA AAGACTGTGA GAGCTTCTNT 120
GGTTGCCCTN TTTTNAGGCN AGACAGATGA AGTTTTTGCT TTCCTTAGTA AGGAGTTTCC 180
25 TGAATTGGGT TTAAAGAAGG AGAATTNTTC GGAGATGACT TGGTTTCANT CTGCTTTATG 240
GTGGGACAAT CGTCTTAATG CTACTCAGGT TGATCCTAAA GTNTTCTTGT ATCGGAATCT 300
30 CGATACCTCG AGTTTCGGTA AGAGGAAATC GGATTACGTC GCGACT 346

(2) INFORMATION FOR SEQ ID NO: 45:

(i) SEQUENCE CHARACTERISTICS:

35 (A) LENGTH: 261 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

40 (ii) MOLECULE TYPE: cDNA to mRNA

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

45

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Arabidopsis thaliana

(B) STRAIN: ecotype Columbia

50 (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 2..261

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 45:

55

ATGGGGTGAG ACTTATTTCA AAGGTAATTT CAAGAGATTA GGTTTGGTTA AAGGGAAGNT 60
 TGATCCAACA AATTTCTTCA GGAACGAACA GAGTATTCCT CCTCTGTTTT GAGTCCTCAA 120
 5 TACAAAACCA GATATAAAAG ATGTCATTTT ATTTTTTCAA TTATAATAGA TAATGTAAGT 180
 TTCTGCTACA ATTGTAAAAG TGAGATGTAC CCAATACGGT TTAAGCGGAC CGAGAATAGT 240
 CAATTCAAAG ACCAAATTCT G 261

10

(2) INFORMATION FOR SEQ ID NO: 46:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 478 base pairs
 15 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

 (ii) MOLECULE TYPE: cDNA to mRNA
 20 (iii) HYPOTHETICAL: NO
 (iii) ANTI-SENSE: NO

 25 (vi) ORIGINAL SOURCE:
 (A) ORGANISM: *Arabidopsis thaliana*
 (B) STRAIN: ecotype Columbia

 (ix) FEATURE:
 30 (A) NAME/KEY: CDS
 (B) LOCATION: 1..478

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 46:

35 GCTCAAAGGA CTAACCATGA AACTTCCTC AAGTGTCTCT CTCACCGANT CAACGAGGAC 60
 GACTCAAGAN TTATACACAC ATCAAAAGAT CCTTCGTATT TNTCAATCTT GATTTCTTCC 120
 ATACAAAATC CAAGTTTCTC TGTTCGAA ACACCTAAAC CGGTTTCAAT CATCACTCCG 180
 40 GTTCAAGCCA CCGATGTTCA ATCTACGNTT AAATNCGCAC GGNCTTCACG GGTATACACA 240
 ATCAGGGCTA GGAGTGGTNG TCATGACTAC GGAGGTTTAT CTTTACATTG GCTTAAAAAN 300
 45 CANNCCGTTT GTTNNTCATT GATTTNNAGA AATCTTCCGG GCTTATTTAA CATNTAAGAT 360
 GTTTGATAAN CCGGNNCCNG TTTGGGGTTC AAATCCCGGT GGCTTACAAA NTNNGGGGGA 420
 ATTGTNCCTA TGAGGTTTGG AAAATTAANG CAAAATNTTT TGGGCCTTCC CGGCCGGT 478

50

(2) INFORMATION FOR SEQ ID NO: 47:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 579 base pairs
 55 (B) TYPE: nucleic acid

(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

5

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

10 (vi) ORIGINAL SOURCE:

(A) ORGANISM: *Arabidopsis thaliana*

(B) STRAIN: ecotype Columbia

(ix) FEATURE:

15

(A) NAME/KEY: CDS

(B) LOCATION: 2..579

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 47:

20 GGCCGTTAGG ATCATCAAGA AATGGCAATA TGCTGCAGAT AAGGTTCTTG ATGATCTTTT 60
CATTAGGACA ACATTGGAGA GATCAAACAA GAACGCAGTA CACGCTTTGT TCACTGGACT 120
ATATATTGGT CCGGTGAACA ATCTATTGGC GTTGATGGAA GAAAAGTTTC CGGAAGTAGG 180
25 TCTTGAGAAA GAAGGTTGTG AAGAGATGAG TTGGATTGAG TCTGTACTCT GGTTCGCTGA 240
TTTCCCTAAA GGAGAATCTC TTGGTGTCTT CACGAATCGT GAGCGTACAT CTCTATCTTT 300
30 CAAAGGCAAA GATGATTTTG TCCAAGAACC GATACCCGAG GCTGCAATTC AAGAGATATG 360
GAGGCGATTA GAAGCCCCCG AGGCTCGGCT TGGAAAGATC ATATTAATCT CATTGCGGTG 420
NGGNAAAATG AGTGAAATGG CAGAGNCCGA ACCACCAATT CCCACANNCG AGGGAGGGGA 480
35 ACCCCTNTGN GGNTCAGAAT GTGGTTCCTG GNNNNNAAGN GGGNGCCAGN ACCAANCCGG 540
GNCNGTAAAN CNTGNAATGG GCCNAACCCG TNCCGGATT 579

40 (2) INFORMATION FOR SEQ ID NO: 48:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 252 base pairs

(B) TYPE: nucleic acid

45

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

50

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

55

(A) ORGANISM: *Oryza sativa*

(B) STRAIN: Nipponbare, subsp. japonica
 (D) DEVELOPMENTAL STAGE: etiolated shoot (8 days old)

(ix) FEATURE:

5 (A) NAME/KEY: CDS
 (B) LOCATION: 3..252

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 48:

10 TGTCTGGAA GGTCCGCCTC GTGCAGGTTN CGACGACGGT GACGGTGTTC GTCGTCGGGA 60
 GGAACGTCGA CCAGGGCGCC GCNGACGTCG TCGCCAGATG GCAAGACGTC GCGCCGAGCC 120
 TCCCTCCCGA GCTCACCATA CGGGTGATCG TNCGAGGGCA GCGCGCCACG TTCCAGTCGC 180
 15 TGTACCTCGG CTCGTGCGCC GACCTGGTGC CGACGATGAG CAGCATGTTC CCGGAGCTCG 240
 GGATGACGAT TG 252

20 (2) INFORMATION FOR SEQ ID NO: 49:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 amino acids
 (B) TYPE: amino acid
 25 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

30 (iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Lactuca sativa
 (B) STRAIN: lollo bionda
 35

(ix) FEATURE:

(A) NAME/KEY: Modified-site
 (B) LOCATION: 12
 (D) OTHER INFORMATION: /label= Ambiguous
 40 /note= "Xaa = Cys or Ser"

(ix) FEATURE:

(A) NAME/KEY: Modified-site
 (B) LOCATION: 20..21
 45 (D) OTHER INFORMATION: /label= ambiguous
 /note= "Xaa-Xaa probably is Ser-Phe"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 49:

50 Thr Ser Thr Ser Ile Ile Asp Arg Phe Thr Gln Xaa Leu Asn Asn Arg
 1 5 10 15
 Ala Asp Pro Xaa Xaa
 20
 55

(2) INFORMATION FOR SEQ ID NO: 50:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 24 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

10

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- 15 (A) ORGANISM: Lactuca sativa
 (B) STRAIN: lollo bionda

(ix) FEATURE:

- 20 (A) NAME/KEY: Modified-site
 (B) LOCATION: 1
 (D) OTHER INFORMATION: /label= ambiguous
 /note= "Xaa = probably Ser"

(ix) FEATURE:

- 25 (A) NAME/KEY: Modified-site
 (B) LOCATION: 3
 (D) OTHER INFORMATION: /label= unknown

(ix) FEATURE:

- 30 (A) NAME/KEY: Modified-site
 (B) LOCATION: 5
 (D) OTHER INFORMATION: /label= ambiguous
 /note= "Xaa = probably Ser"

(ix) FEATURE:

- 35 (A) NAME/KEY: Modified-site
 (B) LOCATION: 12
 (D) OTHER INFORMATION: /label= ambiguous
 /note= "Xaa = probably Trp"

40 (ix) FEATURE:

- (A) NAME/KEY: Modified-site
 (B) LOCATION: 24
 (D) OTHER INFORMATION: /label= ambiguous
 /note= "Xaa = probably Tyr"

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 50:

Xaa Ile Xaa Val Xaa Ile Glu Asp Glu Thr Ala Xaa Val Gln Ala Gly
 1 5 10 15
 Ala Thr Leu Gly Glu Val Tyr Xaa
 20

55

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..405

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 53:

```

ACT TCT ACT TCT ATT ATT GAT AGG TTT ACT CAA TGT CTA AAC AAC CGA      48
Thr Ser Thr Ser Ile Ile Asp Arg Phe Thr Gln Cys Leu Asn Asn Arg
  1             5             10             15

10 GCT GAC CCT TCT TTC CCG CTC AGT GGA CAA CTT TAC ACT CCC GAT AAC      96
Ala Asp Pro Ser Phe Pro Leu Ser Gly Gln Leu Tyr Thr Pro Asp Asn
             20             25             30

15 TCC TCT TTT CCA TCC GTC TTG CAA GCT TAC ATC CGG AAC CTC CGA TTC      144
Ser Ser Phe Pro Ser Val Leu Gln Ala Tyr Ile Arg Asn Leu Arg Phe
             35             40             45

20 AAT GAA TCC ACG ACT CCC AAA CCC ATC TTA ATC ATC ACC GCC TTA CAC      192
Asn Glu Ser Thr Thr Pro Lys Pro Ile Leu Ile Ile Thr Ala Leu His
             50             55             60

25 CCT TCA CAC ATT CAA GCA GCT GTT GTG TGC GCC AAA ACA CAC CGC CTG      240
Pro Ser His Ile Gln Ala Ala Val Val Cys Ala Lys Thr His Arg Leu
             65             70             75             80

CTA ATG AAA ACC AGA AGC GGA GGC CAT GAT TAT GAG GGG CTT TCC TAT      288
Leu Met Lys Thr Arg Ser Gly Gly His Asp Tyr Glu Gly Leu Ser Tyr
             85             90             95

30 GTG ACC AAT TCG AAC CAA CCC TTT TTT GTT GTT GAC ATG TTC AAC TTA      336
Val Thr Asn Ser Asn Gln Pro Phe Phe Val Val Asp Met Phe Asn Leu
             100            105            110

35 CGC TCC ATA AAC GTG AGT ATT GAA GAT GAA ACT GCA TGG GTC CAA GCC      384
Arg Ser Ile Asn Val Ser Ile Glu Asp Glu Thr Ala Trp Val Gln Ala
             115            120            125

GGC GCC ACC CTC GGA GAA GTT
40 Gly Ala Thr Leu Gly Glu Val
             130            135

```

(2) INFORMATION FOR SEQ ID NO: 54:

45 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 135 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 54:

```

Thr Ser Thr Ser Ile Ile Asp Arg Phe Thr Gln Cys Leu Asn Asn Arg
55  1             5             10             15

```

Ala Asp Pro Ser Phe Pro Leu Ser Gly Gln Leu Tyr Thr Pro Asp Asn
 20 25 30

Ser Ser Phe Pro Ser Val Leu Gln Ala Tyr Ile Arg Asn Leu Arg Phe
 5 35 40 45

Asn Glu Ser Thr Thr Pro Lys Pro Ile Leu Ile Ile Thr Ala Leu His
 50 55 60

10 Pro Ser His Ile Gln Ala Ala Val Val Cys Ala Lys Thr His Arg Leu
 65 70 75 80

Leu Met Lys Thr Arg Ser Gly Gly His Asp Tyr Glu Gly Leu Ser Tyr
 85 90 95

15 Val Thr Asn Ser Asn Gln Pro Phe Phe Val Val Asp Met Phe Asn Leu
 100 105 110

Arg Ser Ile Asn Val Ser Ile Glu Asp Glu Thr Ala Trp Val Gln Ala
 20 115 120 125

Gly Ala Thr Leu Gly Glu Val
 130 135

25 (2) INFORMATION FOR SEQ ID NO: 55:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 25 base pairs
 (B) TYPE: nucleic acid
 30 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

35 (iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 55:

CACGTTTATG GAGCGTAAAGT TGAAC

25

40

(2) INFORMATION FOR SEQ ID NO: 56:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 base pairs
 45 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

50

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 56:

55 CACCCTTCAC ACATTCAAGC AGC

23

(2) INFORMATION FOR SEQ ID NO: 57:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1981 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (iii) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: Lactuca sativa
 (B) STRAIN: lollo bionda
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 7..1626
- (ix) FEATURE:
 (A) NAME/KEY: unsure
 (B) LOCATION: replace(372, "g")
- (ix) FEATURE:
 (A) NAME/KEY: unsure
 (B) LOCATION: replace(379, "g")
- (ix) FEATURE:
 (A) NAME/KEY: unsure
 (B) LOCATION: replace(786, "t")
- (ix) FEATURE:
 (A) NAME/KEY: unsure
 (B) LOCATION: replace(1105..1106, "ga")
 (D) OTHER INFORMATION: /note= "also possible "gg" and "aa"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 57:

```

45  ACAAAA ATG GCA ATT ACC TAT TCT TTC AAC TTC AAA TCT TAT ATT TTT      48
    Met Ala Ile Thr Tyr Ser Phe Asn Phe Lys Ser Tyr Ile Phe
        1             5             10

    CCT CTC CTC CTT GTC TTG CTC TCT ACC CAT TCA TCA GCG ACT TCA ACT      96
    Pro Leu Leu Leu Val Leu Leu Ser Thr His Ser Ser Ala Thr Ser Thr
50  15             20             25             30

    TCC ATT ATA GAT CGC TTC ACC CAA TGT CTA AAC AAC CGA GCT GAC CCT      144
    Ser Ile Ile Asp Arg Phe Thr Gln Cys Leu Asn Asn Arg Ala Asp Pro
        35             40             45
55

```

	TCT	TTC	CCG	CTC	AGT	GGA	CAA	CTT	TAC	ACT	CCC	GAT	AAC	TCC	TCT	TTT	192
	Ser	Phe	Pro	Leu	Ser	Gly	Gln	Leu	Tyr	Thr	Pro	Asp	Asn	Ser	Ser	Phe	
				50					55					60			
5	CCA	TCC	GTC	TTG	CAA	GCT	TAC	ATC	CGG	AAC	CTC	CGA	TTC	AAT	GAA	TCC	240
	Pro	Ser	Val	Leu	Gln	Ala	Tyr	Ile	Arg	Asn	Leu	Arg	Phe	Asn	Glu	Ser	
			65					70					75				
	ACG	ACT	CCC	AAA	CCC	ATC	TTA	ATC	ATC	ACC	GCC	TTA	CAC	CCT	TCA	CAC	288
10	Thr	Thr	Pro	Lys	Pro	Ile	Leu	Ile	Ile	Thr	Ala	Leu	His	Pro	Ser	His	
		80					85					90					
	ATT	CAA	GCA	GCT	GTT	GTG	TGC	GCC	AAA	ACA	CAC	CGC	CTG	CTA	ATG	AAA	336
	Ile	Gln	Ala	Ala	Val	Val	Cys	Ala	Lys	Thr	His	Arg	Leu	Leu	Met	Lys	
15	95					100					105					110	
	ACC	AGA	AGC	GGA	GGC	CAT	GAT	TAT	GAG	GGG	CTT	TCC	TAT	GTG	ACC	AAT	384
	Thr	Arg	Ser	Gly	Gly	His	Asp	Tyr	Glu	Gly	Leu	Ser	Tyr	Val	Thr	Asn	
					115					120					125		
20	TCG	AAC	CAA	CCC	TTT	TTT	GTT	GTT	GAC	ATG	TTC	AAC	TTA	CGC	TCC	ATA	432
	Ser	Asn	Gln	Pro	Phe	Phe	Val	Val	Asp	Met	Phe	Asn	Leu	Arg	Ser	Ile	
				130					135					140			
25	AAC	GTG	AGT	ATT	GAA	GAT	GAA	ACT	GCA	TGG	GTC	CAA	GCT	GGT	GCG	ACT	480
	Asn	Val	Ser	Ile	Glu	Asp	Glu	Thr	Ala	Trp	Val	Gln	Ala	Gly	Ala	Thr	
			145					150					155				
	CTT	GGT	GAA	GTC	TAC	TAC	CGA	ATA	GCA	GAG	AAA	AGC	AAC	AGT	CAT	GCT	528
30	Leu	Gly	Glu	Val	Tyr	Tyr	Arg	Ile	Ala	Glu	Lys	Ser	Asn	Ser	His	Ala	
		160					165					170					
	TTT	CCG	GCT	GGC	GTT	TGC	CCT	ACT	GTT	GGA	GTT	GGT	GGC	CAT	TTT	AGT	576
	Phe	Pro	Ala	Gly	Val	Cys	Pro	Thr	Val	Gly	Val	Gly	Gly	His	Phe	Ser	
35	175					180					185					190	
	GGT	GGT	GGT	TAT	GGT	AAC	TTG	ATG	GGA	AAA	TAC	GGC	CTT	TCT	GTT	GAC	624
	Gly	Gly	Gly	Tyr	Gly	Asn	Leu	Met	Gly	Lys	Tyr	Gly	Leu	Ser	Val	Asp	
					195					200					205		
40	AAT	ATT	GTC	GAT	GCT	CAG	TTA	ATC	GAT	GTG	AAT	GGT	AAA	CTT	CTG	AAT	672
	Asn	Ile	Val	Asp	Ala	Gln	Leu	Ile	Asp	Val	Asn	Gly	Lys	Leu	Leu	Asn	
				210					215					220			
45	CGG	AAA	TCA	ATG	GGT	GAA	GAT	CTT	TTT	TGG	GCC	ATC	ACA	GGT	GGT	GGT	720
	Arg	Lys	Ser	Met	Gly	Glu	Asp	Leu	Phe	Trp	Ala	Ile	Thr	Gly	Gly	Gly	
			225					230					235				
	GGT	GTC	AGC	TTT	GGT	GTG	GTT	GTA	GCG	TAC	AAG	ATC	AAA	CTG	GTT	CGT	768
50	Gly	Val	Ser	Phe	Gly	Val	Val	Ala	Tyr	Lys	Ile	Lys	Leu	Val	Arg		
		240					245				250						
	GTT	CCT	ACC	ACT	GTG	ACC	GTT	TTT	AAC	GTA	CAA	AGA	ACA	TCC	GAG	CAG	816
	Val	Pro	Thr	Thr	Val	Thr	Val	Phe	Asn	Val	Gln	Arg	Thr	Ser	Glu	Gln	
55	255					260					265					270	

	AAC CTA AGC ACC ATA GCC CAC CGA TGG ATA CAA GTT GCG GAT AAG CTC	864
	Asn Leu Ser Thr Ile Ala His Arg Trp Ile Gln Val Ala Asp Lys Leu	
	275 280 285	
5	GAT AAT GAC CTT TTC CTT CGA ATG ACC TTT AAC GTG ATA AAC AAC ACA	912
	Asp Asn Asp Leu Phe Leu Arg Met Thr Phe Asn Val Ile Asn Asn Thr	
	290 295 300	
10	AAT GGC GAA AAG ACG ATA CGT GGT TTG TTT CCA ACA CTG TAC CTC GGA	960
	Asn Gly Glu Lys Thr Ile Arg Gly Leu Phe Pro Thr Leu Tyr Leu Gly	
	305 310 315	
15	AAC TCT ACC GCT CTT GTT GCC CTC CTG AAC AAG GAT TTC CCT GAA TTA	1008
	Asn Ser Thr Ala Leu Val Ala Leu Leu Asn Lys Asp Phe Pro Glu Leu	
	320 325 330	
20	GGT GTA GAA ATT TCA GAT TGT ATT GAA ATG AGT TGG ATC GAG TCT GTT	1056
	Gly Val Glu Ile Ser Asp Cys Ile Glu Met Ser Trp Ile Glu Ser Val	
	335 340 345 350	
	CTT TTC TAC ACA AAC TTC CCC ATT GGT ACT CCG ACC ACT GCT CTT CTA	1104
	Leu Phe Tyr Thr Asn Phe Pro Ile Gly Thr Pro Thr Thr Ala Leu Leu	
	355 360 365	
25	AGC CGT ACA CCT CAA AGA CTA AAC CCA TTC AAA ATC AAA TCT GAT TAC	1152
	Ser Arg Thr Pro Gln Arg Leu Asn Pro Phe Lys Ile Lys Ser Asp Tyr	
	370 375 380	
30	GTA AAA AAC ACT ATT TCC AAA CAG GGA TTC GAA TCC ATA TTT GAA AGG	1200
	Val Lys Asn Thr Ile Ser Lys Gln Gly Phe Glu Ser Ile Phe Glu Arg	
	385 390 395	
35	ATG AAA GAA CTC GAA AAC CAA ATG CTA GCT TTC AAC CCT TAT GGT GGA	1248
	Met Lys Glu Leu Glu Asn Gln Met Leu Ala Phe Asn Pro Tyr Gly Gly	
	400 405 410	
40	AGA ATG AGC GAA ATT TCC GAA TTT GCA AAG CCT TTT CCC CAT CGA TCA	1296
	Arg Met Ser Glu Ile Ser Glu Phe Ala Lys Pro Phe Pro His Arg Ser	
	415 420 425 430	
	GGG AAT ATA GCG AAG ATC CAA TAC GAA GTA AAC TGG GAT GAA CTT GGC	1344
	Gly Asn Ile Ala Lys Ile Gln Tyr Glu Val Asn Trp Asp Glu Leu Gly	
	435 440 445	
45	GTT GAA GCA GCC AAT CGG TAC TTG AAC TTC ACA AGG GTG ATG TAT GAT	1392
	Val Glu Ala Ala Asn Arg Tyr Leu Asn Phe Thr Arg Val Met Tyr Asp	
	450 455 460	
50	TAT ATG ACT CCG TTT GTT TCT AAG AAC CCC AGG GAA GCA TTT CTG AAC	1440
	Tyr Met Thr Pro Phe Val Ser Lys Asn Pro Arg Glu Ala Phe Leu Asn	
	465 470 475	
55	TAC AGG GAT TTA GAT ATT GGT GTC AAC AGT CAT GGC AAG AAT GCT TAC	1488
	Tyr Arg Asp Leu Asp Ile Gly Val Asn Ser His Gly Lys Asn Ala Tyr	
	480 485 490	

GGT GAA GGA ATG GTT TAT GGG CAC AAG TAT TTC AAA GAG ACG AAT TAT 1536
 Gly Glu Gly Met Val Tyr Gly His Lys Tyr Phe Lys Glu Thr Asn Tyr
 495 500 505 510

5 AAG AGG CTA ACG ATG GTG AAG ACG AGG GTT GAT CCT AGC AAT TTT TTT 1584
 Lys Arg Leu Thr Met Val Lys Thr Arg Val Asp Pro Ser Asn Phe Phe
 515 520 525

AGG AAT GAG CAA AGT ATC CCA ACT TTG TCA TCT TCA TGG AAG 1626
 10 Arg Asn Glu Gln Ser Ile Pro Thr Leu Ser Ser Ser Trp Lys
 530 535 540

TAAATTCTAA ATTCACCTTGT GAAATTGAAT AAAAGTATGG CTTTTTCAAG GTCATGGTAT 1686

15 CCAGATTCAG ATGATATTGA TATAATTTTG ACTTGTATTT ATACAAACAA AATTATATTA 1746
 TATTTTTCTG AATTTAGATT TTCCATTCTT TGGAAAAATA TACGAACATT GATGTTGATA 1806
 TTTTAAAGAA TTATAGATTT TGAACATTGT GAACAATGAA TAAACCGAGG ACTTCCCTTG 1866
 20 GGTTTTTTTT ATAAGTATGT AATAGCATGT CTTTAATCAA GATAACCGAT CATTGGATGC 1926
 AATTATTAT TATAACCTT ATTTAAAAA AAAAAAAAAA AAAAAAAAAA AAAAA 1981

25 (2) INFORMATION FOR SEQ ID NO: 58:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 540 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 58:

35 Met Ala Ile Thr Tyr Ser Phe Asn Phe Lys Ser Tyr Ile Phe Pro Leu
 1 5 10 15

Leu Leu Val Leu Leu Ser Thr His Ser Ser Ala Thr Ser Thr Ser Ile
 40 20 25 30

Ile Asp Arg Phe Thr Gln Cys Leu Asn Asn Arg Ala Asp Pro Ser Phe
 35 40 45

45 Pro Leu Ser Gly Gln Leu Tyr Thr Pro Asp Asn Ser Ser Phe Pro Ser
 50 55 60

Val Leu Gln Ala Tyr Ile Arg Asn Leu Arg Phe Asn Glu Ser Thr Thr
 65 70 75 80

50 Pro Lys Pro Ile Leu Ile Ile Thr Ala Leu His Pro Ser His Ile Gln
 85 90 95

Ala Ala Val Val Cys Ala Lys Thr His Arg Leu Leu Met Lys Thr Arg
 55 100 105 110

Ser Gly Gly His Asp Tyr Glu Gly Leu Ser Tyr Val Thr Asn Ser Asn
 115 120 125
 5 Gln Pro Phe Phe Val Val Asp Met Phe Asn Leu Arg Ser Ile Asn Val
 130 135 140
 Ser Ile Glu Asp Glu Thr Ala Trp Val Gln Ala Gly Ala Thr Leu Gly
 145 150 155 160
 10 Glu Val Tyr Tyr Arg Ile Ala Glu Lys Ser Asn Ser His Ala Phe Pro
 165 170 175
 Ala Gly Val Cys Pro Thr Val Gly Val Gly Gly His Phe Ser Gly Gly
 15 180 185 190
 Gly Tyr Gly Asn Leu Met Gly Lys Tyr Gly Leu Ser Val Asp Asn Ile
 195 200 205
 20 Val Asp Ala Gln Leu Ile Asp Val Asn Gly Lys Leu Leu Asn Arg Lys
 210 215 220
 Ser Met Gly Glu Asp Leu Phe Trp Ala Ile Thr Gly Gly Gly Gly Val
 225 230 235 240
 25 Ser Phe Gly Val Val Val Ala Tyr Lys Ile Lys Leu Val Arg Val Pro
 245 250 255
 Thr Thr Val Thr Val Phe Asn Val Gln Arg Thr Ser Glu Gln Asn Leu
 30 260 265 270
 Ser Thr Ile Ala His Arg Trp Ile Gln Val Ala Asp Lys Leu Asp Asn
 275 280 285
 35 Asp Leu Phe Leu Arg Met Thr Phe Asn Val Ile Asn Asn Thr Asn Gly
 290 295 300
 Glu Lys Thr Ile Arg Gly Leu Phe Pro Thr Leu Tyr Leu Gly Asn Ser
 305 310 315 320
 40 Thr Ala Leu Val Ala Leu Leu Asn Lys Asp Phe Pro Glu Leu Gly Val
 325 330 335
 Glu Ile Ser Asp Cys Ile Glu Met Ser Trp Ile Glu Ser Val Leu Phe
 45 340 345 350
 Tyr Thr Asn Phe Pro Ile Gly Thr Pro Thr Thr Ala Leu Leu Ser Arg
 355 360 365
 50 Thr Pro Gln Arg Leu Asn Pro Phe Lys Ile Lys Ser Asp Tyr Val Lys
 370 375 380
 Asn Thr Ile Ser Lys Gln Gly Phe Glu Ser Ile Phe Glu Arg Met Lys
 385 390 395 400
 55

	Glu	Leu	Glu	Asn	Gln	Met	Leu	Ala	Phe	Asn	Pro	Tyr	Gly	Gly	Arg	Met	
						405											415
5	Ser	Glu	Ile	Ser	Glu	Phe	Ala	Lys	Pro	Phe	Pro	His	Arg	Ser	Gly	Asn	
				420					425					430			
	Ile	Ala	Lys	Ile	Gln	Tyr	Glu	Val	Asn	Trp	Asp	Glu	Leu	Gly	Val	Glu	
			435					440					445				
10	Ala	Ala	Asn	Arg	Tyr	Leu	Asn	Phe	Thr	Arg	Val	Met	Tyr	Asp	Tyr	Met	
		450					455					460					
	Thr	Pro	Phe	Val	Ser	Lys	Asn	Pro	Arg	Glu	Ala	Phe	Leu	Asn	Tyr	Arg	
	465					470					475					480	
15	Asp	Leu	Asp	Ile	Gly	Val	Asn	Ser	His	Gly	Lys	Asn	Ala	Tyr	Gly	Glu	
					485					490					495		
	Gly	Met	Val	Tyr	Gly	His	Lys	Tyr	Phe	Lys	Glu	Thr	Asn	Tyr	Lys	Arg	
20				500					505					510			
	Leu	Thr	Met	Val	Lys	Thr	Arg	Val	Asp	Pro	Ser	Asn	Phe	Phe	Arg	Asn	
			515					520					525				
25	Glu	Gln	Ser	Ile	Pro	Thr	Leu	Ser	Ser	Ser	Trp	Lys					
		530					535					540					

(2) INFORMATION FOR SEQ ID NO: 59:

30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

40 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 59:

GGTAATGATC TCCTTTCTTG TTTGACC

27

(2) INFORMATION FOR SEQ ID NO: 60:

45 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 41 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 50 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

55

(iii) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 60:

5 AGAGCGGCCG CTATATTACA ACTTCTCCAC CATCACTCCT C 41

(2) INFORMATION FOR SEQ ID NO: 61:

(i) SEQUENCE CHARACTERISTICS:

10 (A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

15 (ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 61:

20 GGTGATGTTA ATGATAATCT CCTC 24

(2) INFORMATION FOR SEQ ID NO: 62:

(i) SEQUENCE CHARACTERISTICS:

25 (A) LENGTH: 40 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

30 (ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 62:

AGAGCGGCCG CTACAATTCC TTCAACATGT AAATTCCTC 40

(2) INFORMATION FOR SEQ ID NO: 63:

40 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
45 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 63:

55 ACTTCCCGTA GAAACTCGGA GACTTTCACA CAATGC 36

(2) INFORMATION FOR SEQ ID NO: 64:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

10

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 64:

15 TCCATCCAAG ATCAATTCAT AAACGTGTGC

30

(2) INFORMATION FOR SEQ ID NO: 65:

(i) SEQUENCE CHARACTERISTICS:

- 20 (A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

25

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 65:

30

AGAGCGGCCG CTTTCATGAA CCTAGCTTCT AGTAGG

36

(2) INFORMATION FOR SEQ ID NO: 66:

(i) SEQUENCE CHARACTERISTICS:

- 35 (A) LENGTH: 37 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

40

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 66:

45

AGAGCGGCCG CGAAATGGCC CCCCTTTTAA AACGGGG

37

(2) INFORMATION FOR SEQ ID NO: 67:

(i) SEQUENCE CHARACTERISTICS:

- 50 (A) LENGTH: 40 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
55 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 67:

AGAGCGGCCG CAAATGATAT CTTCAGGTAA CTTTGTTCAC

40

(2) INFORMATION FOR SEQ ID NO: 68:

10

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 43 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

15

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

20

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 68:

AGAGCGGCCG CATAATCAAA TAAATACACT TATGGTAACA CAG

43

25 (2) INFORMATION FOR SEQ ID NO: 69:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 38 base pairs

(B) TYPE: nucleic acid

30

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

35

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 69:

AGAGCGGCCG CTGGTTTTGT ATTGAGGACT CAAAACAG

38

40

(2) INFORMATION FOR SEQ ID NO: 70:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1757 base pairs

45

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

50

(iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

55

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Arabidopsis thaliana*

(B) STRAIN: Colombia

5 (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: join(1..570, 801..1754)

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 70:

	ACT TCC CGT AGA AAC TCG GAG ACT TTC ACA CAA TGC CTA ACC TCA AAC	48
	Thr Ser Arg Arg Asn Ser Glu Thr Phe Thr Gln Cys Leu Thr Ser Asn	
	1 5 10 15	
15	TCC GAC CCC AAA CAT CCC ATC TCC CCC GCT ATC TTC TTC TCC GGA AAT	96
	Ser Asp Pro Lys His Pro Ile Ser Pro Ala Ile Phe Phe Ser Gly Asn	
	20 25 30	
20	GGC TCC TAC TCC TCC GTA TTA CAA GCC AAC ATC CGT AAC CTC CGC TTC	144
	Gly Ser Tyr Ser Ser Val Leu Gln Ala Asn Ile Arg Asn Leu Arg Phe	
	35 40 45	
	AAC ACC ACC TCA ACT CCG AAA CCC TTC CTC ATA ATC GCC GCA ACA CAT	192
25	Asn Thr Thr Ser Thr Pro Lys Pro Phe Leu Ile Ile Ala Ala Thr His	
	50 55 60	
	GAA TCC CAT GTG CAA GCC GCG ATT ACT TGC GGG AAA CGC CAC AAC CTT	240
	Glu Ser His Val Gln Ala Ala Ile Thr Cys Gly Lys Arg His Asn Leu	
30	65 70 75 80	
	CAG ATG AAA ATC AGA AGT GGA GGC CAC GAC TAC GAT GGC TTG TCA TAC	288
	Gln Met Lys Ile Arg Ser Gly Gly His Asp Tyr Asp Gly Leu Ser Tyr	
	85 90 95	
35	GTT ACA TAC TCT GGC AAA CCG TTC TTC GTC CTC GAC ATG TTT AAC CTC	336
	Val Thr Tyr Ser Gly Lys Pro Phe Phe Val Leu Asp Met Phe Asn Leu	
	100 105 110	
40	CGT TCG GTG GAT GTC GAT GTG GCA AGT AAG ACC GCG TGG GTC CAA ACC	384
	Arg Ser Val Asp Val Asp Val Ala Ser Lys Thr Ala Trp Val Gln Thr	
	115 120 125	
	GGT GCC ATA CTC GGA GAA GTT TAT TAC TAT ATA TGG GAG AAG AGC AAA	432
45	Gly Ala Ile Leu Gly Glu Val Tyr Tyr Tyr Ile Trp Glu Lys Ser Lys	
	130 135 140	
	ACC CTA GCT TAT CCC GCC GGA ATT TGT CCC ACG GTT GGT GTC GGT GGC	480
	Thr Leu Ala Tyr Pro Ala Gly Ile Cys Pro Thr Val Gly Val Gly Gly	
50	145 150 155 160	
	CAT ATC AGT GGT GGA GGT TAC GGT AAC ATG ATG AGA AAA TAC GGT CTC	528
	His Ile Ser Gly Gly Gly Tyr Gly Asn Met Met Arg Lys Tyr Gly Leu	
	165 170 175	
55		

ACC GTA GAT AAT ACC ATC GAT GCA AGA ATG GTC GAC GTT AAT 570
 Thr Val Asp Asn Thr Ile Asp Ala Arg Met Val Asp Val Asn
 180 185 190

5 GGTATAATTG ATATCTCTAT TTTATATACT AATTAAATTT TATAGTGTGG ATCGGATAGT 630
 GATTTTGGTC CATCAATTAA AAACCTGGTG AACATAAAAT TAACCAAGCA ATCAATTTAG 690
 ACAAGCAACA TAATCATATA TATTTTTCTT ACATTTGTAT GTACCTGAAT ATTTATATTT 750

10 ATGTTTATAT GTTCTCACTA TATTTTCACT TTTGTATTTG AAAATTTTGA GGA AAA 806
 Gly Lys

ATT TTG GAT AGA AAA TTG ATG GGA GAA GAT CTC TAC TGG GCA ATA AAC 854
 15 Ile Leu Asp Arg Lys Leu Met Gly Glu Asp Leu Tyr Trp Ala Ile Asn
 195 200 205

GGA GGA GGA GGA GGG AGC TAC GGC GTC GTA TTG GCC TAC AAA ATA AAC 902
 Gly Gly Gly Gly Gly Ser Tyr Gly Val Val Leu Ala Tyr Lys Ile Asn
 20 210 215 220

CTT GTT GAA GTC CCA GAA AAC GTC ACC GTT TTC AGA ATC TCC CGG ACG 950
 Leu Val Glu Val Pro Glu Asn Val Thr Val Phe Arg Ile Ser Arg Thr
 225 230 235 240

25 TTA GAA CAA AAT GCG ACG GAT ATC ATT CAC CGG TGG CAA CAA GTT GCA 998
 Leu Glu Gln Asn Ala Thr Asp Ile Ile His Arg Trp Gln Gln Val Ala
 245 250 255

30 CCG AAG CTT CCC GAC GAG CTT TTC ATA AGA ACA GTC ATT GAC GTA GTA 1046
 Pro Lys Leu Pro Asp Glu Leu Phe Ile Arg Thr Val Ile Asp Val Val
 260 265 270

AAC GGC ACT GTT TCA TCT CAA AAG ACC GTC AGG ACA ACA TTC ATA GCA 1094
 35 Asn Gly Thr Val Ser Ser Gln Lys Thr Val Arg Thr Thr Phe Ile Ala
 275 280 285

ATG TTT CTA GGA GAC ACG ACA ACT CTA CTG TCG ATA TTA AAC CGG AGA 1142
 Met Phe Leu Gly Asp Thr Thr Thr Leu Leu Ser Ile Leu Asn Arg Arg
 40 290 295 300

TTC CCA GAA TTG GGT TTG GTC CGG TCT GAC TGT ACC GAA ACA AGC TGG 1190
 Phe Pro Glu Leu Gly Leu Val Arg Ser Asp Cys Thr Glu Thr Ser Trp
 305 310 315 320

45 ATC CAA TCT GTG CTA TTC TGG ACA AAT ATC CAA GTT GGT TCG TCG GAG 1238
 Ile Gln Ser Val Leu Phe Trp Thr Asn Ile Gln Val Gly Ser Ser Glu
 325 330 335

50 ACA CTT CTA CTC CAA AGG AAT CAA CCC GTG AAC TAC CTC AAG AGG AAA 1286
 Thr Leu Leu Leu Gln Arg Asn Gln Pro Val Asn Tyr Leu Lys Arg Lys
 340 345 350

TCA GAT TAC GTA CGT GAA CCG ATT TCA AGA ACC GGT TTA GAG TCA ATT 1334
 Ser Asp Tyr Val Arg Glu Pro Ile Ser Arg Thr Gly Leu Glu Ser Ile
 355 360 365

5 TGG AAG AAA ATG ATC GAG CTT GAA ATT CCG ACA ATG GCT TTC AAT CCA 1382
 Trp Lys Lys Met Ile Glu Leu Glu Ile Pro Thr Met Ala Phe Asn Pro
 370 375 380

10 TAC GGT GGT GAG ATG GGG AGG ATA TCA TTA CGG GTG ACT CCG TTC CCA 1430
 Tyr Gly Gly Glu Met Gly Arg Ile Ser Leu Arg Val Thr Pro Phe Pro
 385 390 395 400

15 TAC AGA GCC GGT AAT CTC TGG AAG ATT CAG TAC GGT GCG AAT TGG AGA 1478
 Tyr Arg Ala Gly Asn Leu Trp Lys Ile Gln Tyr Gly Ala Asn Trp Arg
 405 410 415

20 GAT GAG ACT TTA ACC GAC CGG TAC ATG GAA TTG ACG AGG AAG TTG TAC 1526
 Asp Glu Thr Leu Thr Asp Arg Tyr Met Glu Leu Thr Arg Lys Leu Tyr
 420 425 430

CAA TTC ATG ACA CCA TTT GTT TCC AAG AAT CCG AGA CAA TCG TTT TTC 1574
 Gln Phe Met Thr Pro Phe Val Ser Lys Asn Pro Arg Gln Ser Phe Phe
 435 440 445

25 AAT AAC CGT GAT GTT GAT TTG GGT ATT AAT TCT CAT AAT GGT AAA ATC 1622
 Asn Asn Arg Asp Val Asp Leu Gly Ile Asn Ser His Asn Gly Lys Ile
 450 455 460

30 AGT AGT TAT GTG GAA GGT AAA CGT TAC GGG AAG AAG TAT TTC GCA GGT 1670
 Ser Ser Tyr Val Glu Gly Lys Arg Tyr Gly Lys Lys Tyr Phe Ala Gly
 465 470 475 480

35 AAT TTC GAG AGA TTG GTG AAG ATT AAG ACG AGA GTT GAT AGT GGT AAT 1718
 Asn Phe Glu Arg Leu Val Lys Ile Lys Thr Arg Val Asp Ser Gly Asn
 485 490 495

TTC TTT AGG AAC GAA CAC AGT ATT CCT GTG TTA CCA TAA 1757
 Phe Phe Arg Asn Glu His Ser Ile Pro Val Leu Pro
 500 505

40

(2) INFORMATION FOR SEQ ID NO: 71:

(i) SEQUENCE CHARACTERISTICS:

- 45 (A) LENGTH: 508 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 71:

Thr Ser Arg Arg Asn Ser Glu Thr Phe Thr Gln Cys Leu Thr Ser Asn
 1 5 10 15
 55

Ser Asp Pro Lys His Pro Ile Ser Pro Ala Ile Phe Phe Ser Gly Asn
 20 25 30
 Gly Ser Tyr Ser Ser Val Leu Gln Ala Asn Ile Arg Asn Leu Arg Phe
 5 35 40 45
 Asn Thr Thr Ser Thr Pro Lys Pro Phe Leu Ile Ile Ala Ala Thr His
 50 55 60
 10 Glu Ser His Val Gln Ala Ala Ile Thr Cys Gly Lys Arg His Asn Leu
 65 70 75 80
 Gln Met Lys Ile Arg Ser Gly Gly His Asp Tyr Asp Gly Leu Ser Tyr
 85 90 95
 15 Val Thr Tyr Ser Gly Lys Pro Phe Phe Val Leu Asp Met Phe Asn Leu
 100 105 110
 Arg Ser Val Asp Val Asp Val Ala Ser Lys Thr Ala Trp Val Gln Thr
 115 120 125
 Gly Ala Ile Leu Gly Glu Val Tyr Tyr Tyr Ile Trp Glu Lys Ser Lys
 130 135 140
 25 Thr Leu Ala Tyr Pro Ala Gly Ile Cys Pro Thr Val Gly Val Gly Gly
 145 150 155 160
 His Ile Ser Gly Gly Gly Tyr Gly Asn Met Met Arg Lys Tyr Gly Leu
 165 170 175
 30 Thr Val Asp Asn Thr Ile Asp Ala Arg Met Val Asp Val Asn Gly Lys
 180 185 190
 Ile Leu Asp Arg Lys Leu Met Gly Glu Asp Leu Tyr Trp Ala Ile Asn
 195 200 205
 Gly Gly Gly Gly Gly Ser Tyr Gly Val Val Leu Ala Tyr Lys Ile Asn
 210 215 220
 40 Leu Val Glu Val Pro Glu Asn Val Thr Val Phe Arg Ile Ser Arg Thr
 225 230 235 240
 Leu Glu Gln Asn Ala Thr Asp Ile Ile His Arg Trp Gln Gln Val Ala
 245 250 255
 45 Pro Lys Leu Pro Asp Glu Leu Phe Ile Arg Thr Val Ile Asp Val Val
 260 265 270
 Asn Gly Thr Val Ser Ser Gln Lys Thr Val Arg Thr Thr Phe Ile Ala
 275 280 285
 50 Met Phe Leu Gly Asp Thr Thr Thr Leu Leu Ser Ile Leu Asn Arg Arg
 290 295 300
 55

Phe Pro Glu Leu Gly Leu Val Arg Ser Asp Cys Thr Glu Thr Ser Trp
 305 310 315 320
 5 Ile Gln Ser Val Leu Phe Trp Thr Asn Ile Gln Val Gly Ser Ser Glu
 325 330 335
 Thr Leu Leu Leu Gln Arg Asn Gln Pro Val Asn Tyr Leu Lys Arg Lys
 340 345 350
 10 Ser Asp Tyr Val Arg Glu Pro Ile Ser Arg Thr Gly Leu Glu Ser Ile
 355 360 365
 Trp Lys Lys Met Ile Glu Leu Glu Ile Pro Thr Met Ala Phe Asn Pro
 370 375 380
 15 Tyr Gly Gly Glu Met Gly Arg Ile Ser Leu Arg Val Thr Pro Phe Pro
 385 390 395 400
 Tyr Arg Ala Gly Asn Leu Trp Lys Ile Gln Tyr Gly Ala Asn Trp Arg
 405 410 415
 20 Asp Glu Thr Leu Thr Asp Arg Tyr Met Glu Leu Thr Arg Lys Leu Tyr
 420 425 430
 Gln Phe Met Thr Pro Phe Val Ser Lys Asn Pro Arg Gln Ser Phe Phe
 435 440 445
 25 Asn Asn Arg Asp Val Asp Leu Gly Ile Asn Ser His Asn Gly Lys Ile
 450 455 460
 30 Ser Ser Tyr Val Glu Gly Lys Arg Tyr Gly Lys Lys Tyr Phe Ala Gly
 465 470 475 480
 Asn Phe Glu Arg Leu Val Lys Ile Lys Thr Arg Val Asp Ser Gly Asn
 485 490 495
 35 Phe Phe Arg Asn Glu His Ser Ile Pro Val Leu Pro
 500 505

40 (2) INFORMATION FOR SEQ ID NO: 72:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1527 base pairs
 (B) TYPE: nucleic acid
 45 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

50 (iii) HYPOTHETICAL: NO

(iii) ANTI-SENSE: NO

55

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Arabidopsis thaliana*

(B) STRAIN: Colombia

5 (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..1524

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 72:

ACT TCC CGT AGA AAC TCG GAG ACT TTC ACA CAA TGC CTA ACC TCA AAC	48
Thr Ser Arg Arg Asn Ser Glu Thr Phe Thr Gln Cys Leu Thr Ser Asn	
1 5 10 15	
15 TCC GAC CCC AAA CAT CCC ATC TCC CCC GCT ATC TTC TTC TCC GGA AAT	96
Ser Asp Pro Lys His Pro Ile Ser Pro Ala Ile Phe Phe Ser Gly Asn	
20 25 30	
20 GGC TCC TAC TCC TCC GTA TTA CAA GCC AAC ATC CGT AAC CTC CGC TTC	144
Gly Ser Tyr Ser Ser Val Leu Gln Ala Asn Ile Arg Asn Leu Arg Phe	
35 40 45	
AAC ACC ACC TCA ACT CCG AAA CCC TTC CTC ATA ATC GCC GCA ACA CAT	192
Asn Thr Thr Ser Thr Pro Lys Pro Phe Leu Ile Ile Ala Ala Thr His	
50 55 60	
GAA TCC CAT GTG CAA GCC GCG ATT ACT TGC GGG AAA CGC CAC AAC CTT	240
Glu Ser His Val Gln Ala Ala Ile Thr Cys Gly Lys Arg His Asn Leu	
30 65 70 75 80	
CAG ATG AAA ATC AGA AGT GGA GGC CAC GAC TAC GAT GGC TTG TCA TAC	288
Gln Met Lys Ile Arg Ser Gly Gly His Asp Tyr Asp Gly Leu Ser Tyr	
85 90 95	
35 GTT ACA TAC TCT GGC AAA CCG TTC TTC GTC CTC GAC ATG TTT AAC CTC	336
Val Thr Tyr Ser Gly Lys Pro Phe Phe Val Leu Asp Met Phe Asn Leu	
100 105 110	
40 CGT TCG GTG GAT GTC GAC GTG GCA AGT AAG ACC GCG TGG GTC CAA ACC	384
Arg Ser Val Asp Val Asp Val Ala Ser Lys Thr Ala Trp Val Gln Thr	
115 120 125	
GGT GCC ATA CTC GGA GAA GTT TAT TAC TAT ATA TGG GAG AAG AGC AAA	432
Gly Ala Ile Leu Gly Glu Val Tyr Tyr Tyr Ile Trp Glu Lys Ser Lys	
130 135 140	
ACC CTA GCT TAT CCC GCC GGA ATT TGT CCC ACG GTT GGT GTC GGT GGC	480
Thr Leu Ala Tyr Pro Ala Gly Ile Cys Pro Thr Val Gly Val Gly Gly	
50 145 150 155 160	
CAT ATC AGT GGT GGA GGT TAC GGT AAC ATG ATG AGA AAA TAC GGT CTC	528
His Ile Ser Gly Gly Gly Tyr Gly Asn Met Met Arg Lys Tyr Gly Leu	
165 170 175	
55	

	ACC	GTA	GAT	AAT	ACC	ATC	GAT	GCA	AGA	ATG	GTC	GAC	GTA	AAT	GGA	AAA	576
	Thr	Val	Asp	Asn	Thr	Ile	Asp	Ala	Arg	Met	Val	Asp	Val	Asn	Gly	Lys	
				180					185						190		
5	ATT	TTG	GAT	AGA	AAA	TTG	ATG	GGA	GAA	GAT	CTC	TAC	TGG	GCA	ATA	AAC	624
	Ile	Leu	Asp	Arg	Lys	Leu	Met	Gly	Glu	Asp	Leu	Tyr	Trp	Ala	Ile	Asn	
			195					200					205				
10	GGA	GGA	GGA	GGA	GGG	AGC	TAC	GGC	GTC	GTA	TTG	GCC	TAC	AAA	ATA	AAC	672
	Gly	Gly	Gly	Gly	Gly	Ser	Tyr	Gly	Val	Val	Leu	Ala	Tyr	Lys	Ile	Asn	
		210					215					220					
15	CTT	GTT	GAA	GTC	CCA	GAA	AAC	GTC	ACC	GTT	TTC	AGA	ATC	TCC	CGG	ACG	720
	Leu	Val	Glu	Val	Pro	Glu	Asn	Val	Thr	Val	Phe	Arg	Ile	Ser	Arg	Thr	
	225					230					235					240	
20	TTA	GAA	CAA	AAT	GCG	ACG	GAT	ATC	ATT	CAC	CGG	TGG	CAA	CAA	GTT	GCA	768
	Leu	Glu	Gln	Asn	Ala	Thr	Asp	Ile	Ile	His	Arg	Trp	Gln	Gln	Val	Ala	
				245						250					255		
	CCG	AAG	CTT	CCC	GAC	GAG	CTT	TTC	ATA	AGA	ACA	GTC	ATT	GAC	GTA	GTA	816
	Pro	Lys	Leu	Pro	Asp	Glu	Leu	Phe	Ile	Arg	Thr	Val	Ile	Asp	Val	Val	
				260					265					270			
25	AAC	GGC	ACT	GTT	TCA	TCT	CAA	AAG	ACC	GTC	AGG	ACA	ACA	TTC	ATA	GCA	864
	Asn	Gly	Thr	Val	Ser	Ser	Gln	Lys	Thr	Val	Arg	Thr	Thr	Phe	Ile	Ala	
			275					280					285				
30	ATG	TTT	CTA	GGA	GAC	ACG	ACA	ACT	CTA	CTG	TCG	ATA	TTA	AAC	CGG	AGA	912
	Met	Phe	Leu	Gly	Asp	Thr	Thr	Thr	Leu	Leu	Ser	Ile	Leu	Asn	Arg	Arg	
		290					295					300					
35	TTC	CCA	GAA	TTG	GGT	TTG	GTC	CGG	TCT	GAC	TGT	ACC	GAA	ACA	AGC	TGG	960
	Phe	Pro	Glu	Leu	Gly	Leu	Val	Arg	Ser	Asp	Cys	Thr	Glu	Thr	Ser	Trp	
	305					310					315					320	
40	ATC	CAA	TCT	GTG	CTA	TTC	TGG	ACA	AAT	ATC	CAA	GTT	GGT	TCG	TCG	GAG	1008
	Ile	Gln	Ser	Val	Leu	Phe	Trp	Thr	Asn	Ile	Gln	Val	Gly	Ser	Ser	Glu	
				325					330						335		
	ACA	CTT	CTA	CTC	CAA	AGG	AAT	CAA	CCC	GTG	AAC	TAC	CTC	AAG	AGG	AAA	1056
	Thr	Leu	Leu	Leu	Gln	Arg	Asn	Gln	Pro	Val	Asn	Tyr	Leu	Lys	Arg	Lys	
				340					345					350			
45	TCA	GAT	TAC	GTA	CGT	GAA	CCG	ATT	TCA	AGA	ACC	GGT	TTA	GAG	TCA	ATT	1104
	Ser	Asp	Tyr	Val	Arg	Glu	Pro	Ile	Ser	Arg	Thr	Gly	Leu	Glu	Ser	Ile	
			355					360					365				
50	TGG	AAG	AAA	ATG	ATC	GAG	CTT	GAA	ATT	CCG	ACA	ATG	GCT	TTC	AAT	CCA	1152
	Trp	Lys	Lys	Met	Ile	Glu	Leu	Glu	Ile	Pro	Thr	Met	Ala	Phe	Asn	Pro	
		370					375					380					
55	TAC	GGT	GGT	GAG	ATG	GGG	AGG	ATA	TCA	TCT	ACG	GTG	ACT	CCG	TTC	CCA	1200
	Tyr	Gly	Gly	Glu	Met	Gly	Arg	Ile	Ser	Ser	Thr	Val	Thr	Pro	Phe	Pro	
	385					390					395					400	

	TAC	AGA	GCC	GGT	AAT	CTC	TGG	AAG	ATT	CAG	TAC	GGT	GCG	AAT	TGG	AGA	1248
	Tyr	Arg	Ala	Gly	Asn	Leu	Trp	Lys	Ile	Gln	Tyr	Gly	Ala	Asn	Trp	Arg	
					405					410					415		
5	GAT	GAG	ACT	TTA	ACC	GAC	CGG	TAC	ATG	GAA	TTG	ACG	AGG	AAG	TTG	TAC	1296
	Asp	Glu	Thr	Leu	Thr	Asp	Arg	Tyr	Met	Glu	Leu	Thr	Arg	Lys	Leu	Tyr	
				420					425					430			
	CAA	TTC	ATG	ACA	CCA	TTT	GTT	TCC	AAG	AAT	CCG	AGA	CAA	TCG	TTT	TTC	1344
10	Gln	Phe	Met	Thr	Pro	Phe	Val	Ser	Lys	Asn	Pro	Arg	Gln	Ser	Phe	Phe	
			435					440					445				
	AAT	TAC	CGT	GAT	GTT	GAT	TTG	GGT	ATT	AAT	TCT	CAT	AAT	GGT	AAA	ATC	1392
15	Asn	Tyr	Arg	Asp	Val	Asp	Leu	Gly	Ile	Asn	Ser	His	Asn	Gly	Lys	Ile	
		450					455					460					
	AGT	AGT	TAT	GTG	GAA	GGT	AAA	CGT	TAC	GGG	AAG	AAG	TAT	TTC	GCA	GGT	1440
	Ser	Ser	Tyr	Val	Glu	Gly	Lys	Arg	Tyr	Gly	Lys	Lys	Tyr	Phe	Ala	Gly	
	465					470				475						480	
20	AAT	TTC	GAG	AGA	TTG	GTG	AAG	ATT	AAG	ACG	AGA	GTT	GAT	AGT	GGT	AAT	1488
	Asn	Phe	Glu	Arg	Leu	Val	Lys	Ile	Lys	Thr	Arg	Val	Asp	Ser	Gly	Asn	
					485					490					495		
25	TTC	TTT	AGG	AAC	GAA	CAG	AGT	ATT	CCT	GTG	TTA	CCA	TAA				1527
	Phe	Phe	Arg	Asn	Glu	Gln	Ser	Ile	Pro	Val	Leu	Pro					
				500					505								

30 (2) INFORMATION FOR SEQ ID NO: 73:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 508 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 73:

[illegible]

Gln Met Lys Ile Arg Ser Gly Gly His Asp Tyr Asp Gly Leu Ser Tyr
 85 90 95
 Val Thr Tyr Ser Gly Lys Pro Phe Phe Val Leu Asp Met Phe Asn Leu
 5 100 105 110
 Arg Ser Val Asp Val Asp Val Ala Ser Lys Thr Ala Trp Val Gln Thr
 115 120 125
 10 Gly Ala Ile Leu Gly Glu Val Tyr Tyr Tyr Ile Trp Glu Lys Ser Lys
 130 135 140
 Thr Leu Ala Tyr Pro Ala Gly Ile Cys Pro Thr Val Gly Val Gly Gly
 145 150 155 160
 15 His Ile Ser Gly Gly Gly Tyr Gly Asn Met Met Arg Lys Tyr Gly Leu
 165 170 175
 Thr Val Asp Asn Thr Ile Asp Ala Arg Met Val Asp Val Asn Gly Lys
 20 180 185 190
 Ile Leu Asp Arg Lys Leu Met Gly Glu Asp Leu Tyr Trp Ala Ile Asn
 195 200 205
 25 Gly Gly Gly Gly Gly Ser Tyr Gly Val Val Leu Ala Tyr Lys Ile Asn
 210 215 220
 Leu Val Glu Val Pro Glu Asn Val Thr Val Phe Arg Ile Ser Arg Thr
 225 230 235 240
 30 Leu Glu Gln Asn Ala Thr Asp Ile Ile His Arg Trp Gln Gln Val Ala
 245 250 255
 Pro Lys Leu Pro Asp Glu Leu Phe Ile Arg Thr Val Ile Asp Val Val
 35 260 265 270
 Asn Gly Thr Val Ser Ser Gln Lys Thr Val Arg Thr Thr Phe Ile Ala
 275 280 285
 40 Met Phe Leu Gly Asp Thr Thr Thr Leu Leu Ser Ile Leu Asn Arg Arg
 290 295 300
 Phe Pro Glu Leu Gly Leu Val Arg Ser Asp Cys Thr Glu Thr Ser Trp
 305 310 315 320
 45 Ile Gln Ser Val Leu Phe Trp Thr Asn Ile Gln Val Gly Ser Ser Glu
 325 330 335
 Thr Leu Leu Leu Gln Arg Asn Gln Pro Val Asn Tyr Leu Lys Arg Lys
 50 340 345 350
 Ser Asp Tyr Val Arg Glu Pro Ile Ser Arg Thr Gly Leu Glu Ser Ile
 355 360 365

55

Trp Lys Lys Met Ile Glu Leu Glu Ile Pro Thr Met Ala Phe Asn Pro
 370 375 380
 Tyr Gly Gly Glu Met Gly Arg Ile Ser Ser Thr Val Thr Pro Phe Pro
 5 385 390 395 400
 Tyr Arg Ala Gly Asn Leu Trp Lys Ile Gln Tyr Gly Ala Asn Trp Arg
 405 410 415
 10 Asp Glu Thr Leu Thr Asp Arg Tyr Met Glu Leu Thr Arg Lys Leu Tyr
 420 425 430
 Gln Phe Met Thr Pro Phe Val Ser Lys Asn Pro Arg Gln Ser Phe Phe
 435 440 445
 15 Asn Tyr Arg Asp Val Asp Leu Gly Ile Asn Ser His Asn Gly Lys Ile
 450 455 460
 Ser Ser Tyr Val Glu Gly Lys Arg Tyr Gly Lys Lys Tyr Phe Ala Gly
 20 465 470 475 480
 Asn Phe Glu Arg Leu Val Lys Ile Lys Thr Arg Val Asp Ser Gly Asn
 485 490 495
 25 Phe Phe Arg Asn Glu Gln Ser Ile Pro Val Leu Pro
 500 505

(2) INFORMATION FOR SEQ ID NO: 74:

- 30 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1530 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
 35 (ii) MOLECULE TYPE: cDNA to mRNA
 (iii) HYPOTHETICAL: NO
 40 (iii) ANTI-SENSE: NO
 (vi) ORIGINAL SOURCE:
 (A) ORGANISM: *Arabidopsis thaliana*
 (B) STRAIN: Colombia
 45 (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 1..1527
 50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 74:

TCC ATC CAA GAT CAA TTC ATA AAC TGT GTC AAA AGA AAC ACA CAT GTT 48
 Ser Ile Gln Asp Gln Phe Ile Asn Cys Val Lys Arg Asn Thr His Val
 55 1 5 10 15

	TCT	TTT	CCA	CTC	GAG	AAA	ACG	TTA	TTC	ACC	CCT	GCG	AAA	AAC	GTC	TCT	96
	Ser	Phe	Pro	Leu	Glu	Lys	Thr	Leu	Phe	Thr	Pro	Ala	Lys	Asn	Val	Ser	
				20					25						30		
5	TTG	TTC	AAC	CAA	GTC	CTT	GAA	TCG	ACG	GCT	CAA	AAT	CTC	CAG	TTC	TTG	144
	Leu	Phe	Asn	Gln	Val	Leu	Glu	Ser	Thr	Ala	Gln	Asn	Leu	Gln	Phe	Leu	
			35					40						45			
	GCA	AAA	TCC	ATG	CCT	AAA	CCG	GGA	TTC	ATA	TTC	AGA	CCG	ATT	CAC	CAG	192
10	Ala	Lys	Ser	Met	Pro	Lys	Pro	Gly	Phe	Ile	Phe	Arg	Pro	Ile	His	Gln	
		50					55					60					
	TCT	CAA	GTC	CAA	GCT	TCC	ATC	ATT	TGT	TCA	AAG	AAA	CTC	GGA	ATT	CAT	240
15	Ser	Gln	Val	Gln	Ala	Ser	Ile	Ile	Cys	Ser	Lys	Lys	Leu	Gly	Ile	His	
	65					70				75						80	
	TTT	CGT	GTT	AGA	AGT	GGC	GGT	CAC	GAT	TTC	GAG	GCC	TTG	TCT	TAT	GTT	288
	Phe	Arg	Val	Arg	Ser	Gly	Gly	His	Asp	Phe	Glu	Ala	Leu	Ser	Tyr	Val	
				85					90						95		
20	TCA	CGG	ATT	GAA	AAA	CCG	TTT	ATA	TTA	CTC	GAC	CTG	TCA	AAA	TTG	AAA	336
	Ser	Arg	Ile	Glu	Lys	Pro	Phe	Ile	Leu	Leu	Asp	Leu	Ser	Lys	Leu	Lys	
				100					105					110			
25	CAA	ATC	AAT	GTT	GAT	ATT	GAA	TCC	AAT	AGT	GCT	TGG	GTT	CAA	CCT	GGT	384
	Gln	Ile	Asn	Val	Asp	Ile	Glu	Ser	Asn	Ser	Ala	Trp	Val	Gln	Pro	Gly	
			115					120					125				
	GCT	ACG	CTT	GGT	GAG	CTT	TAC	TAC	AGA	ATT	GCA	GAG	AAG	AGC	AAG	ATC	432
30	Ala	Thr	Leu	Gly	Glu	Leu	Tyr	Tyr	Arg	Ile	Ala	Glu	Lys	Ser	Lys	Ile	
		130					135					140					
	CAT	GGA	TTT	CCC	GCG	GGT	TTG	TGC	ACA	AGT	GTA	GGC	ATA	GGT	GGG	TAT	480
35	His	Gly	Phe	Pro	Ala	Gly	Leu	Cys	Thr	Ser	Val	Gly	Ile	Gly	Gly	Tyr	
	145					150					155					160	
	ATG	ACA	GGC	GGT	GGA	TAC	GGT	ACC	TTG	ATG	AGG	AAG	TAT	GGT	CTT	GCG	528
	Met	Thr	Gly	Gly	Gly	Tyr	Gly	Thr	Leu	Met	Arg	Lys	Tyr	Gly	Leu	Ala	
				165					170						175		
40	GGA	GAT	AAT	GTT	CTA	GAC	GTA	AAG	ATG	GTT	GAT	GCA	AAT	GGT	AAA	TTA	576
	Gly	Asp	Asn	Val	Leu	Asp	Val	Lys	Met	Val	Asp	Ala	Asn	Gly	Lys	Leu	
				180				185						190			
45	CTC	GAC	AGA	GCC	GCG	ATG	GGT	GAG	GAC	CTA	TTT	TGG	GCG	ATT	AGA	GGA	624
	Leu	Asp	Arg	Ala	Ala	Met	Gly	Glu	Asp	Leu	Phe	Trp	Ala	Ile	Arg	Gly	
			195					200					205				
	GGC	GGT	GGA	GCG	AGT	TTC	GGG	ATA	GTT	CTA	GCA	TGG	AAG	ATC	AAG	CTT	672
50	Gly	Gly	Gly	Ala	Ser	Phe	Gly	Ile	Val	Leu	Ala	Trp	Lys	Ile	Lys	Leu	
		210					215					220					
	GTT	CCT	GTT	CCT	AAG	ACT	GTT	ACC	GTC	TTC	ACT	GTC	ACC	AAA	ACG	TTA	720
55	Val	Pro	Val	Pro	Lys	Thr	Val	Thr	Val	Phe	Thr	Val	Thr	Lys	Thr	Leu	
	225					230					235					240	

	GAA CAA GAC GCA AGA TTG AAG ACT ATT TCT AAG TGG CAA CAA ATT TCA	768
	Glu Gln Asp Ala Arg Leu Lys Thr Ile Ser Lys Trp Gln Gln Ile Ser	
	245 250 255	
5	TCC AAG ATT ATT GAA GAG ATA CAC ATC CGA GTG GTA CTC AGA GCA GCT	816
	Ser Lys Ile Ile Glu Glu Ile His Ile Arg Val Val Leu Arg Ala Ala	
	260 265 270	
10	GGA AAT GAT GGA AAC AAG ACT GTG ACA ATG ACC TAC CTA GGT CAG TTT	864
	Gly Asn Asp Gly Asn Lys Thr Val Thr Met Thr Tyr Leu Gly Gln Phe	
	275 280 285	
15	CTT GGC GAG AAA GGC ACC TTG CTG AAG GTT ATG GAG AAG GCT TTT CCA	912
	Leu Gly Glu Lys Gly Thr Leu Leu Lys Val Met Glu Lys Ala Phe Pro	
	290 295 300	
20	GAA CTA GGG TTA ACT CAA AAG GAT TGT ACT GAA ATG AGC TGG ATT GAA	960
	Glu Leu Gly Leu Thr Gln Lys Asp Cys Thr Glu Met Ser Trp Ile Glu	
	305 310 315 320	
25	GCC GCC CTT TTC CAT GGT GGA TTT CCA ACA GGT TCT CCT ATT GAA ATT	1008
	Ala Ala Leu Phe His Gly Gly Phe Pro Thr Gly Ser Pro Ile Glu Ile	
	325 330 335	
30	TTG CTT CAG CTC AAG TCG CCT CTA GGA AAA GAT TAC TTC AAA GCA ACG	1056
	Leu Leu Gln Leu Lys Ser Pro Leu Gly Lys Asp Tyr Phe Lys Ala Thr	
	340 345 350	
35	TCG GAT TTC GTT AAA GAA CCT ATT CCT GTG ATA GGC TTC AAA GGA ATA	1104
	Ser Asp Phe Val Lys Glu Pro Ile Pro Val Ile Gly Phe Lys Gly Ile	
	355 360 365	
40	TTC AAA AGA TTG ATT GAA GGA AAC ACA ACA TTT CTG AAC TGG ACT CCT	1152
	Phe Lys Arg Leu Ile Glu Gly Asn Thr Thr Phe Leu Asn Trp Thr Pro	
	370 375 380	
45	TAC GGT GGT ATG ATG TCG AAA ATC CCT GAA TCT GCG ATC CCA TTT CCG	1200
	Tyr Gly Gly Met Met Ser Lys Ile Pro Glu Ser Ala Ile Pro Phe Pro	
	385 390 395 400	
50	CAT AGA AAC GGA ACC CTC TTC AAG ATT CTC TAT TAC GCG AAC TGG CTA	1248
	His Arg Asn Gly Thr Leu Phe Lys Ile Leu Tyr Tyr Ala Asn Trp Leu	
	405 410 415	
55	GAG AAT GAC AAG ACA TCG AGT AGA AAA ATC AAC TGG ATC AAA GAG ATA	1296
	Glu Asn Asp Lys Thr Ser Ser Arg Lys Ile Asn Trp Ile Lys Glu Ile	
	420 425 430	
60	TAC AAT TAC ATG GCG CCT TAT GTC TCA AGC AAT CCA AGA CAA GCA TAT	1344
	Tyr Asn Tyr Met Ala Pro Tyr Val Ser Ser Asn Pro Arg Gln Ala Tyr	
	435 440 445	
65	GTG AAC TAC AGA GAT CTA GAC TTC GGA CAG AAC AAG AAC AAC GCA AAG	1392
	Val Asn Tyr Arg Asp Leu Asp Phe Gly Gln Asn Lys Asn Asn Ala Lys	
	450 455 460	

GTT AAC TTC ATT GAA GCT AAA ATC TGG GGA CCT AAG TAC TTC AAA GGC 1440
Val Asn Phe Ile Glu Ala Lys Ile Trp Gly Pro Lys Tyr Phe Lys Gly
465 470 475 480

5 AAT TTT GAC AGA TTG GTG AAG ATT AAA ACC AAG GTT GAT CCA GAG AAC 1488
Asn Phe Asp Arg Leu Val Lys Ile Lys Thr Lys Val Asp Pro Glu Asn
485 490 495

TTC TTC AGG CAC GAG CAG AGT ATC CCA CCT ATG CCC TAC TAG 1530
10 Phe Phe Arg His Glu Gln Ser Ile Pro Pro Met Pro Tyr
500 505

(2) INFORMATION FOR SEQ ID NO: 75:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 509 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 75:

Ser Ile Gln Asp Gln Phe Ile Asn Cys Val Lys Arg Asn Thr His Val
25 1 5 10 15

Ser Phe Pro Leu Glu Lys Thr Leu Phe Thr Pro Ala Lys Asn Val Ser
20 25 30

Leu Phe Asn Gln Val Leu Glu Ser Thr Ala Gln Asn Leu Gln Phe Leu
30 35 40 45

Ala Lys Ser Met Pro Lys Pro Gly Phe Ile Phe Arg Pro Ile His Gln
50 55 60

35 Ser Gln Val Gln Ala Ser Ile Ile Cys Ser Lys Lys Leu Gly Ile His
65 70 75 80

Phe Arg Val Arg Ser Gly Gly His Asp Phe Glu Ala Leu Ser Tyr Val
40 85 90 95

Ser Arg Ile Glu Lys Pro Phe Ile Leu Leu Asp Leu Ser Lys Leu Lys
100 105 110

45 Gln Ile Asn Val Asp Ile Glu Ser Asn Ser Ala Trp Val Gln Pro Gly
115 120 125

Ala Thr Leu Gly Glu Leu Tyr Tyr Arg Ile Ala Glu Lys Ser Lys Ile
130 135 140

50 His Gly Phe Pro Ala Gly Leu Cys Thr Ser Val Gly Ile Gly Gly Tyr
145 150 155 160

Met Thr Gly Gly Gly Tyr Gly Thr Leu Met Arg Lys Tyr Gly Leu Ala
55 165 170 175

Gly Asp Asn Val Leu Asp Val Lys Met Val Asp Ala Asn Gly Lys Leu
 180 185 190
 5 Leu Asp Arg Ala Ala Met Gly Glu Asp Leu Phe Trp Ala Ile Arg Gly
 195 200 205
 Gly Gly Gly Ala Ser Phe Gly Ile Val Leu Ala Trp Lys Ile Lys Leu
 210 215 220
 10 Val Pro Val Pro Lys Thr Val Thr Val Phe Thr Val Thr Lys Thr Leu
 225 230 235 240
 Glu Gln Asp Ala Arg Leu Lys Thr Ile Ser Lys Trp Gln Gln Ile Ser
 15 245 250 255
 Ser Lys Ile Ile Glu Glu Ile His Ile Arg Val Val Leu Arg Ala Ala
 260 265 270
 20 Gly Asn Asp Gly Asn Lys Thr Val Thr Met Thr Tyr Leu Gly Gln Phe
 275 280 285
 Leu Gly Glu Lys Gly Thr Leu Leu Lys Val Met Glu Lys Ala Phe Pro
 290 295 300
 25 Glu Leu Gly Leu Thr Gln Lys Asp Cys Thr Glu Met Ser Trp Ile Glu
 305 310 315 320
 Ala Ala Leu Phe His Gly Gly Phe Pro Thr Gly Ser Pro Ile Glu Ile
 30 325 330 335
 Leu Leu Gln Leu Lys Ser Pro Leu Gly Lys Asp Tyr Phe Lys Ala Thr
 340 345 350
 35 Ser Asp Phe Val Lys Glu Pro Ile Pro Val Ile Gly Phe Lys Gly Ile
 355 360 365
 Phe Lys Arg Leu Ile Glu Gly Asn Thr Thr Phe Leu Asn Trp Thr Pro
 370 375 380
 40 Tyr Gly Gly Met Met Ser Lys Ile Pro Glu Ser Ala Ile Pro Phe Pro
 385 390 395 400
 His Arg Asn Gly Thr Leu Phe Lys Ile Leu Tyr Tyr Ala Asn Trp Leu
 45 405 410 415
 Glu Asn Asp Lys Thr Ser Ser Arg Lys Ile Asn Trp Ile Lys Glu Ile
 420 425 430
 50 Tyr Asn Tyr Met Ala Pro Tyr Val Ser Ser Asn Pro Arg Gln Ala Tyr
 435 440 445
 Val Asn Tyr Arg Asp Leu Asp Phe Gly Gln Asn Lys Asn Asn Ala Lys
 450 455 460
 55

Val Asn Phe Ile Glu Ala Lys Ile Trp Gly Pro Lys Tyr Phe Lys Gly
465 470 475 480

5 Asn Phe Asp Arg Leu Val Lys Ile Lys Thr Lys Val Asp Pro Glu Asn
485 490 495

Phe Phe Arg His Glu Gln Ser Ile Pro Pro Met Pro Tyr
500 505

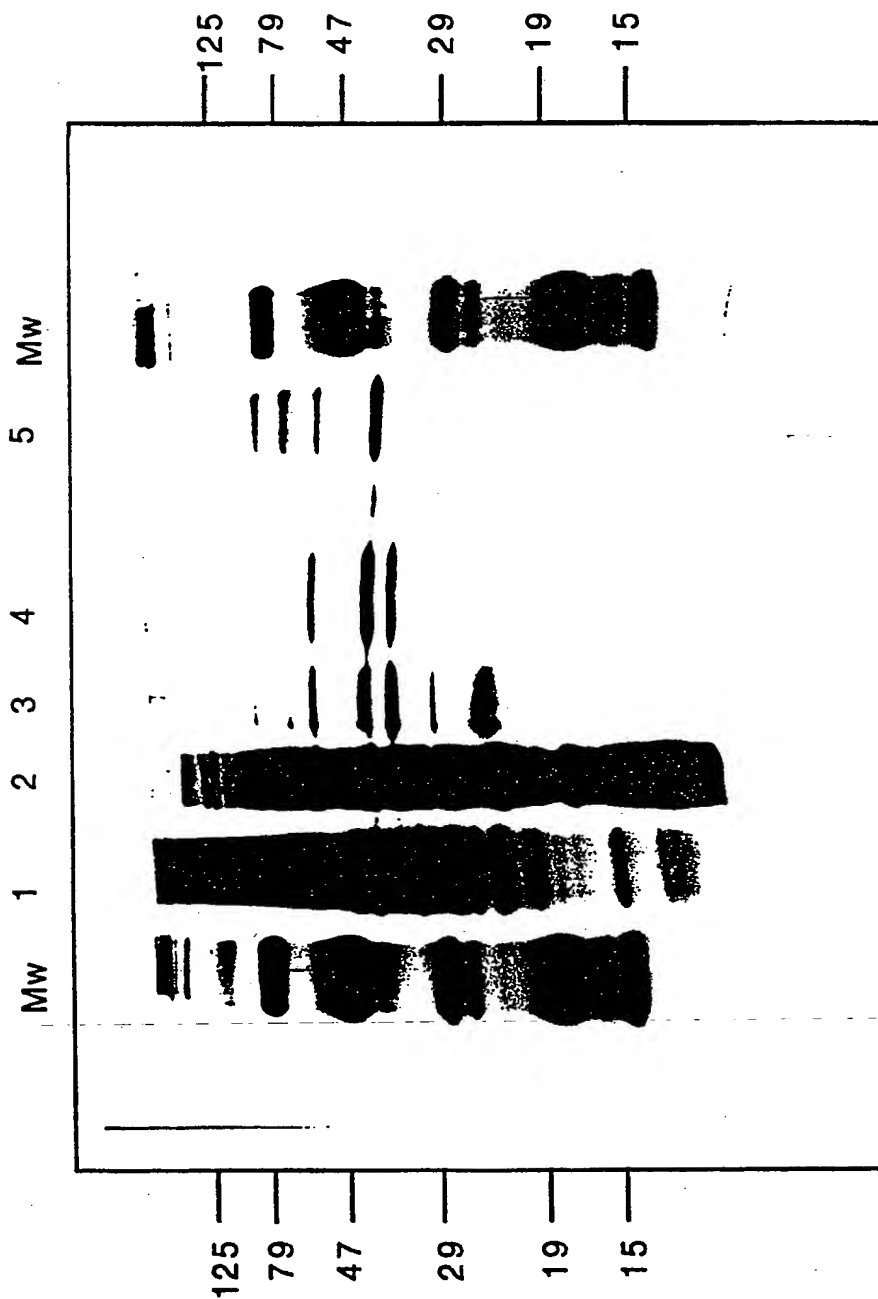


Fig. 1

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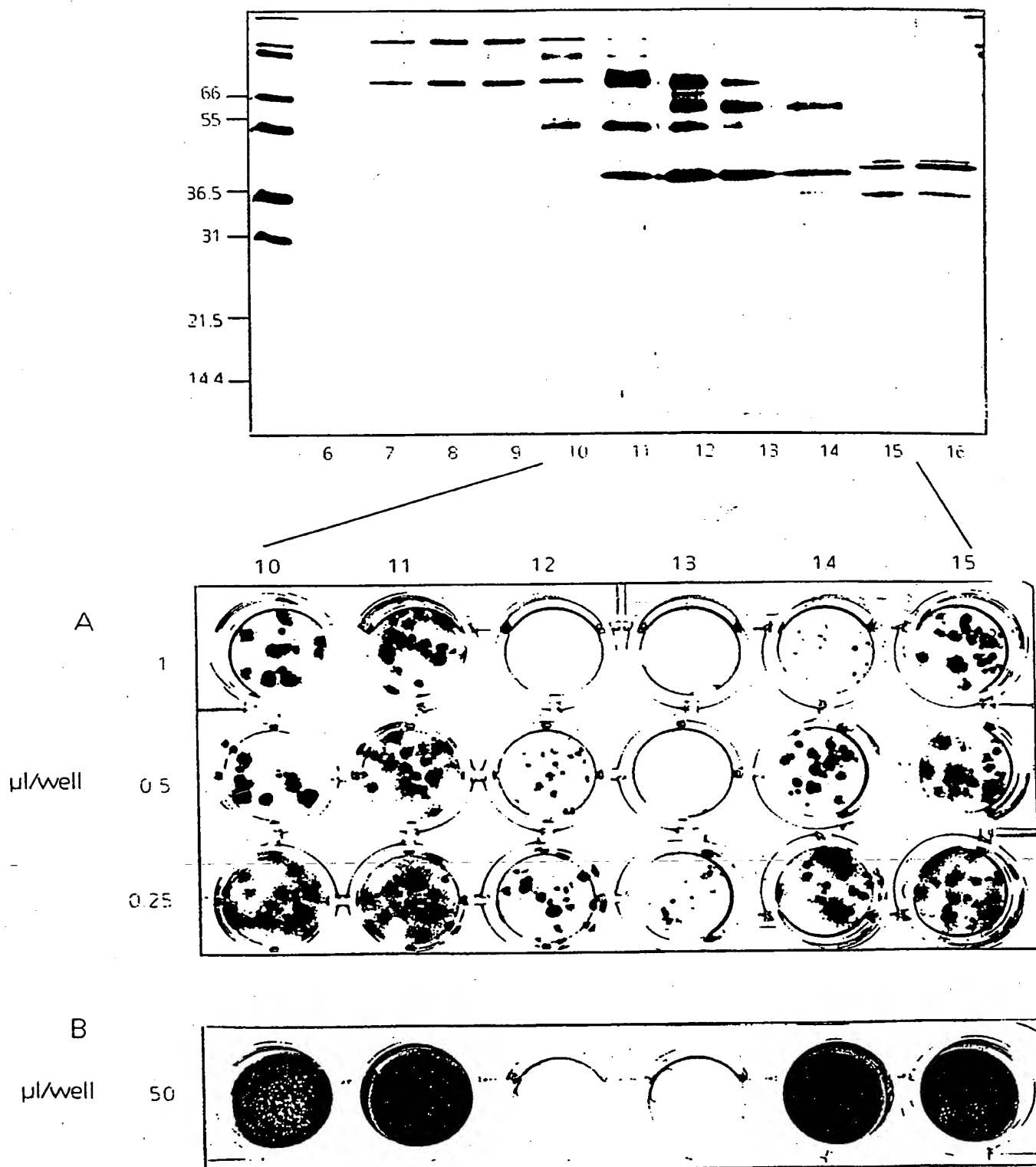


Fig. 2

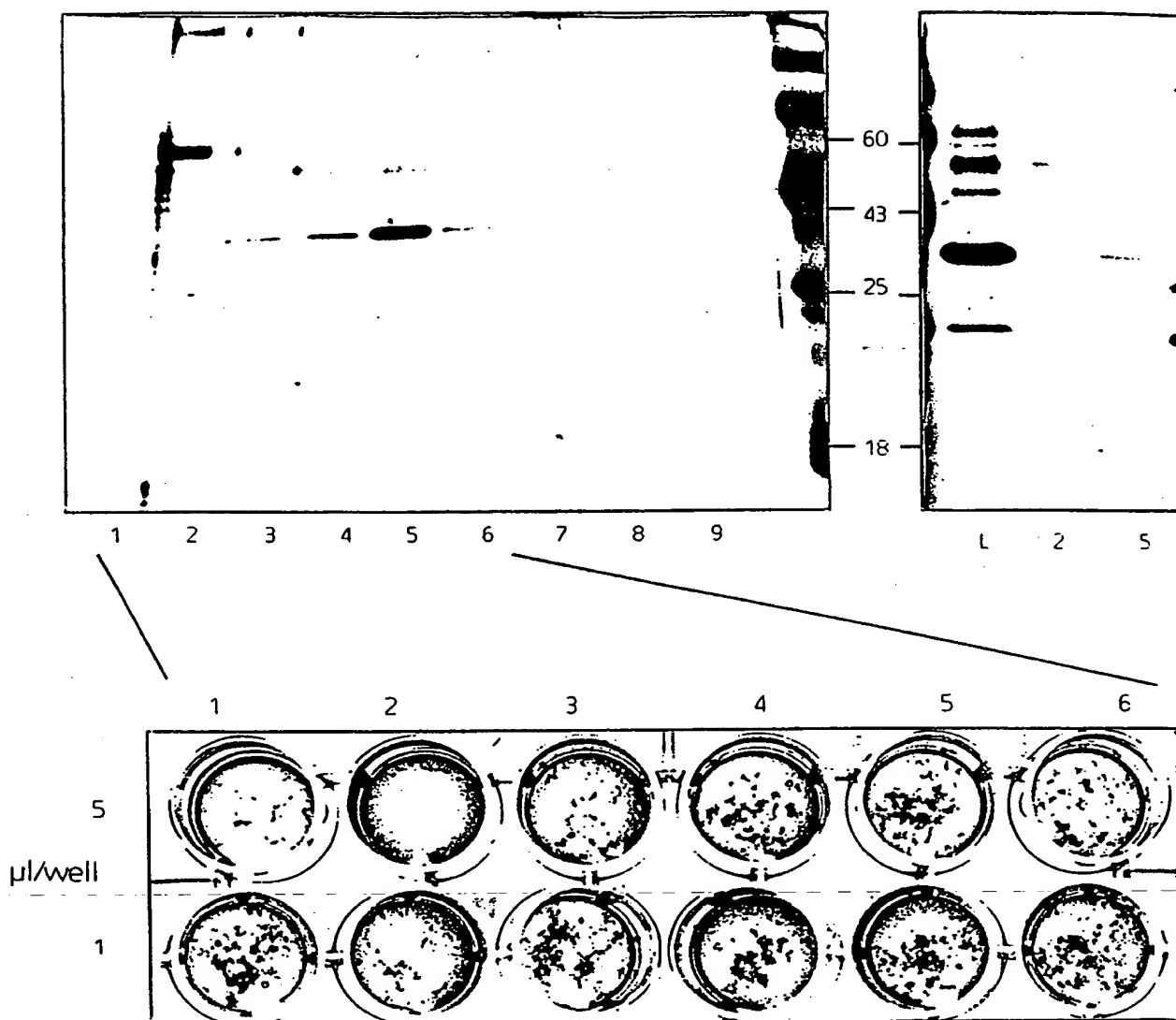
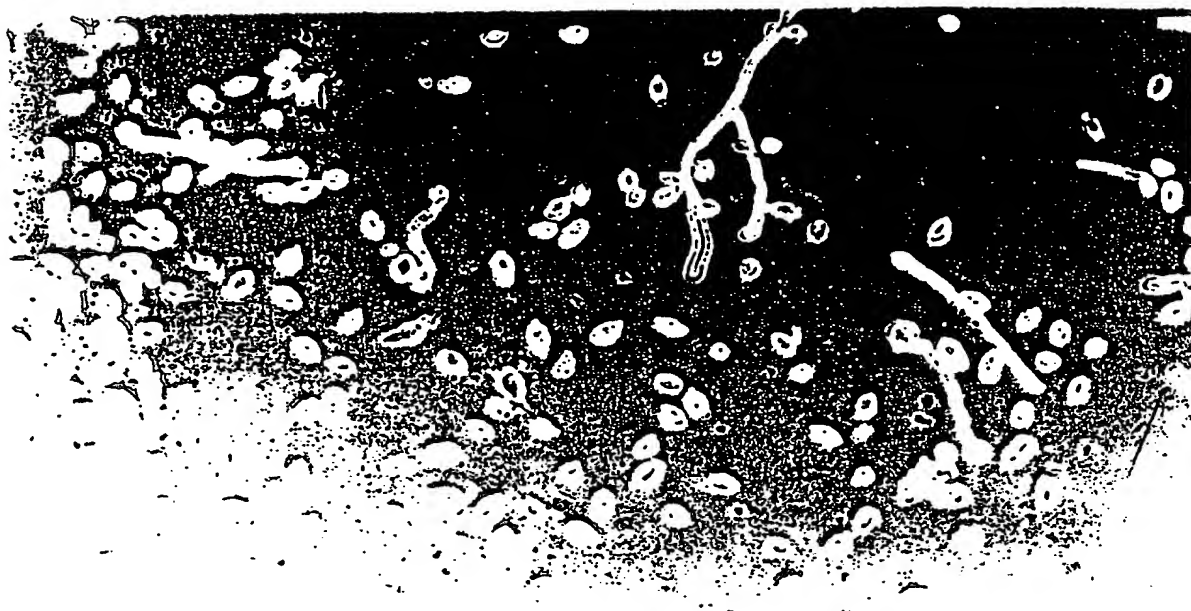
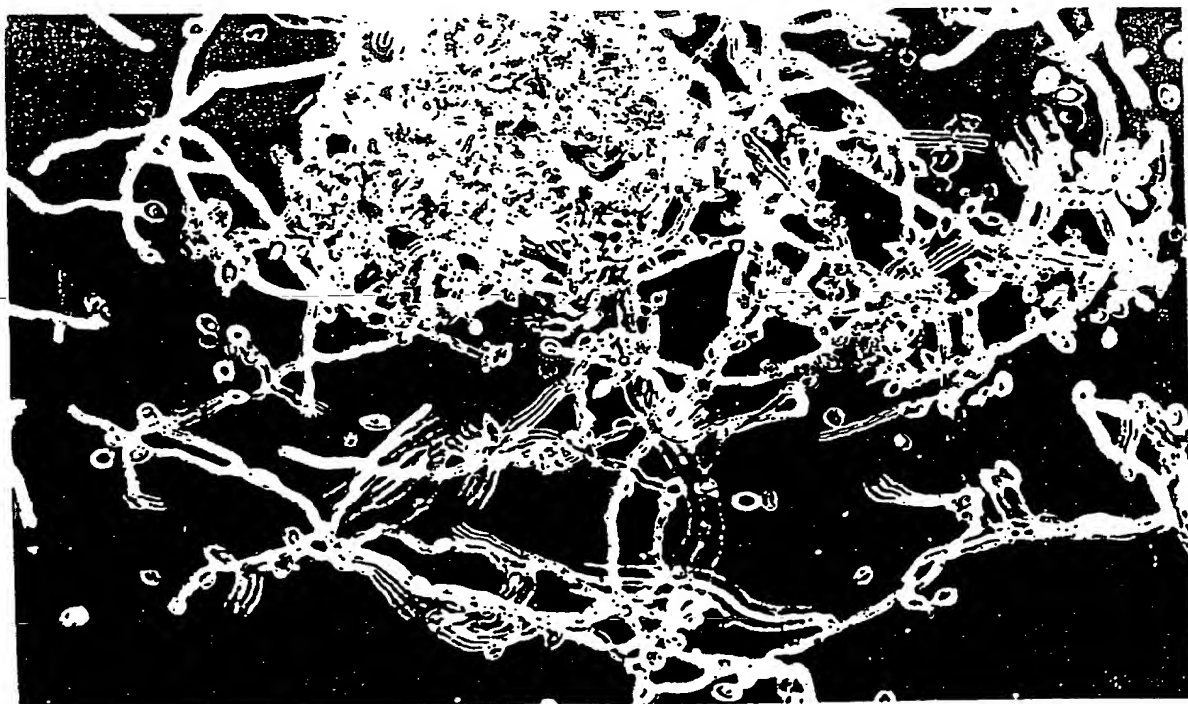


Fig. 3



RIGHT

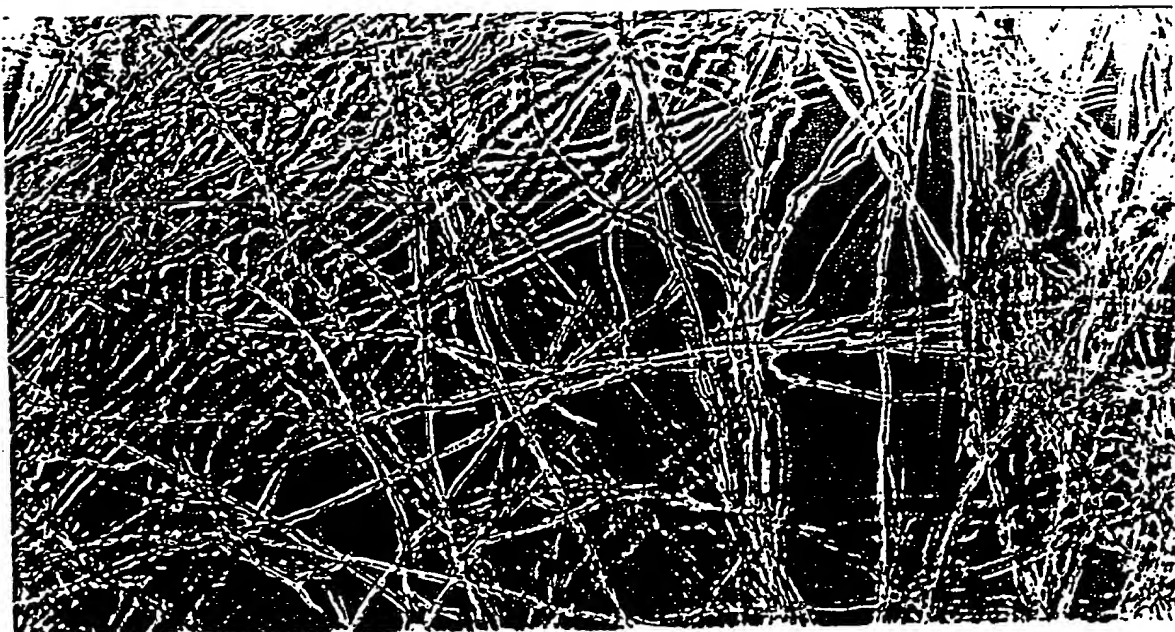


LEFT

Fig. 4



RIGHT



LEFT

Fig. 5

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6 A

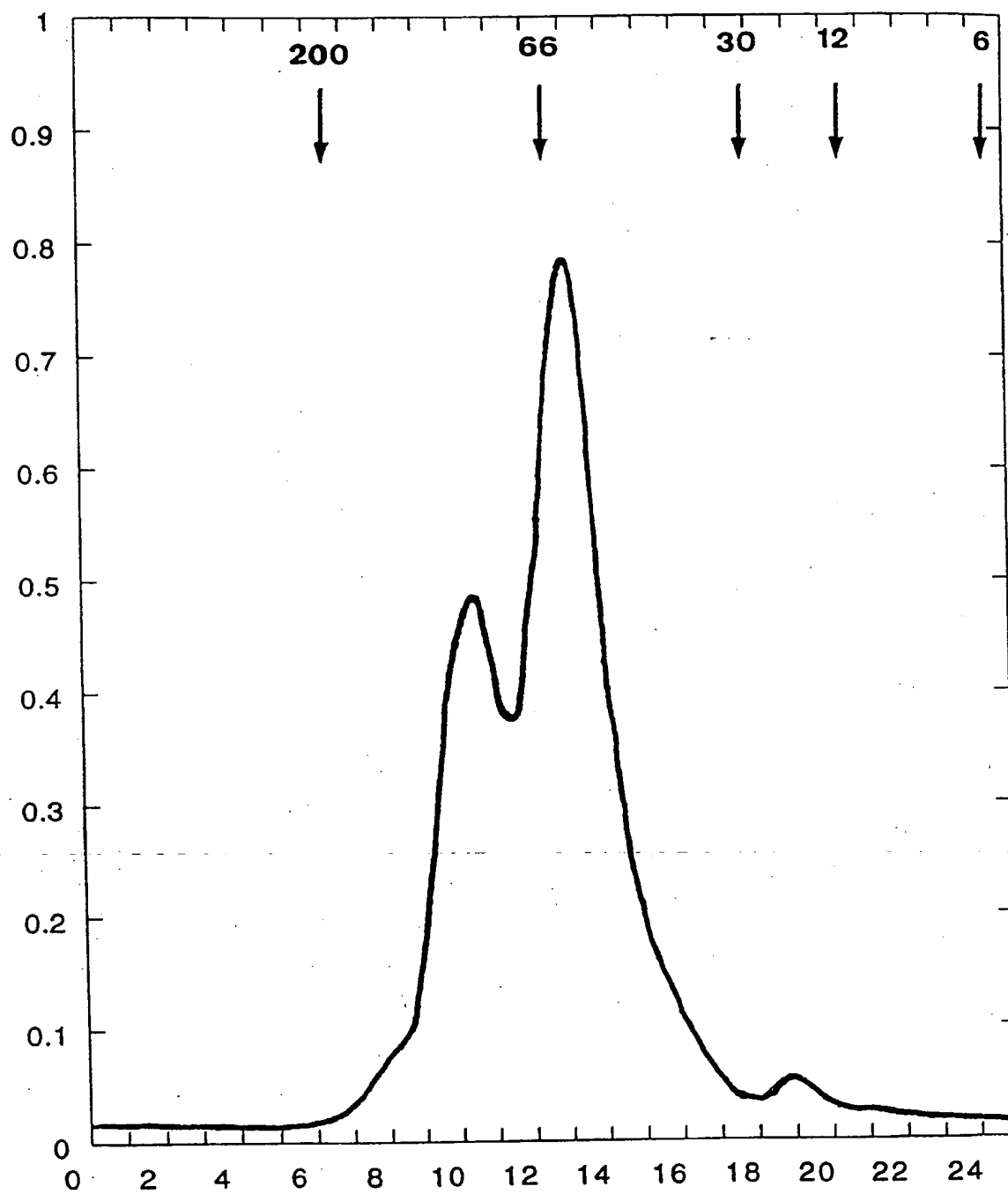


Fig. 6-1

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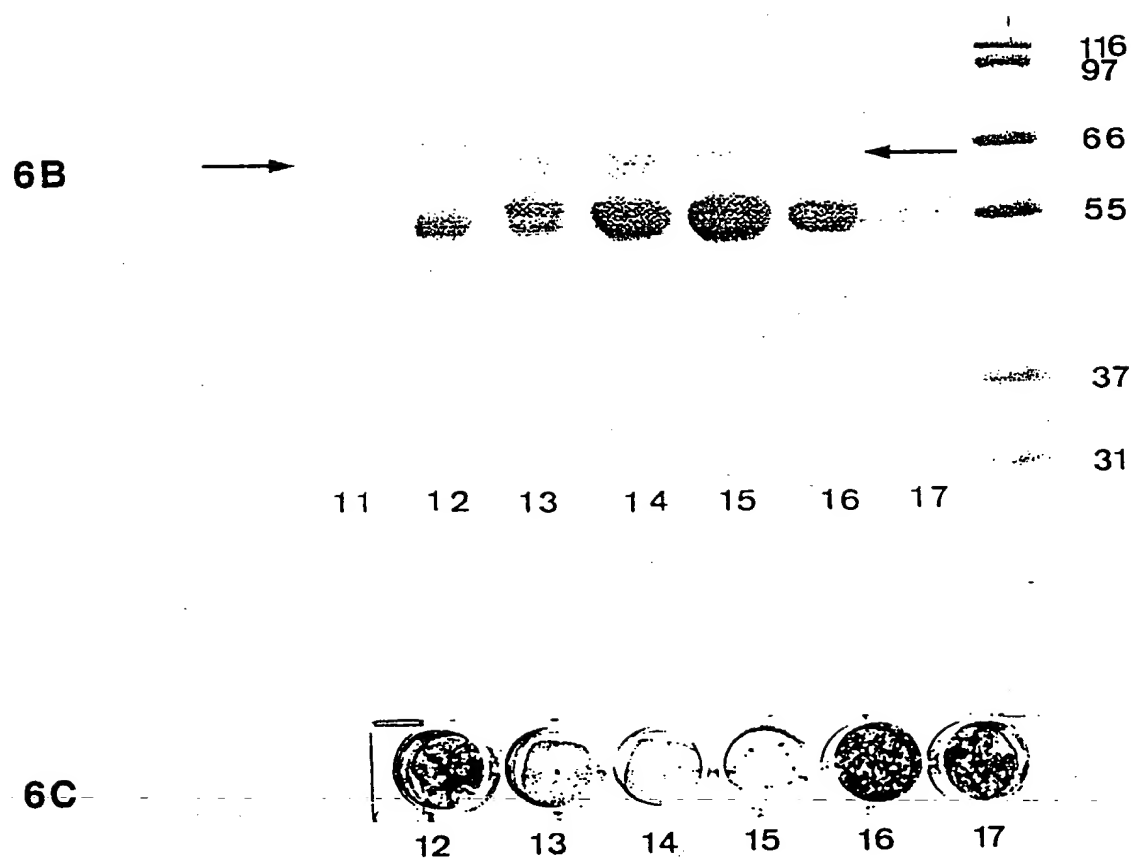


Fig. 6-2

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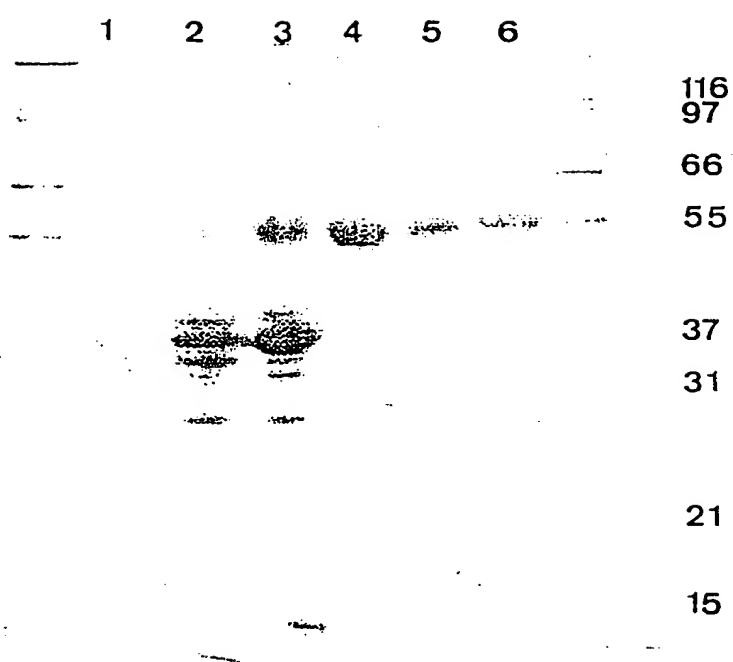


Fig. 7

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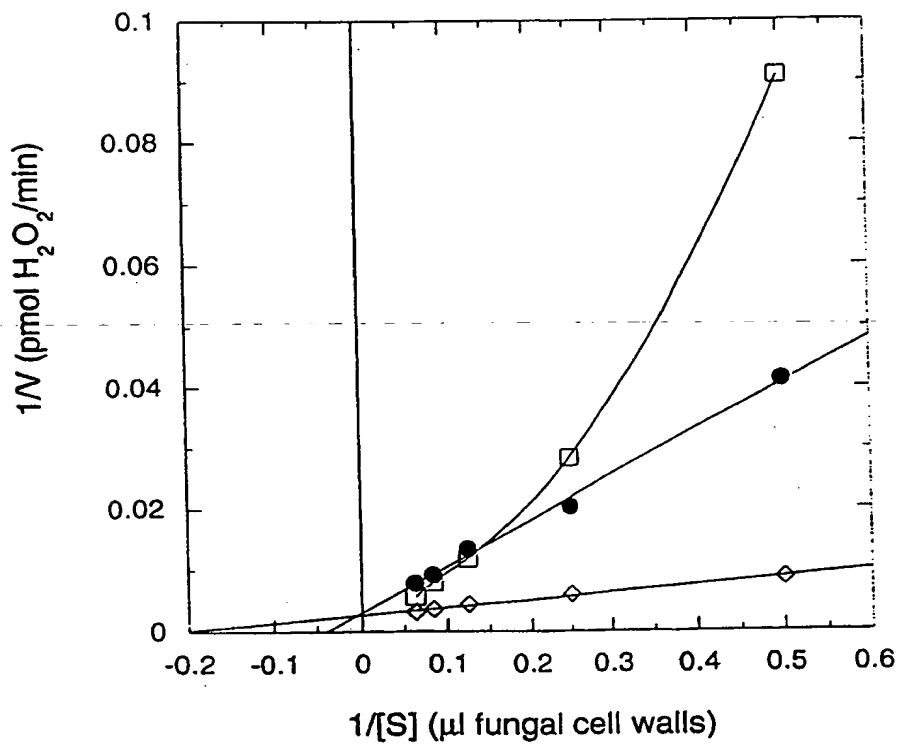
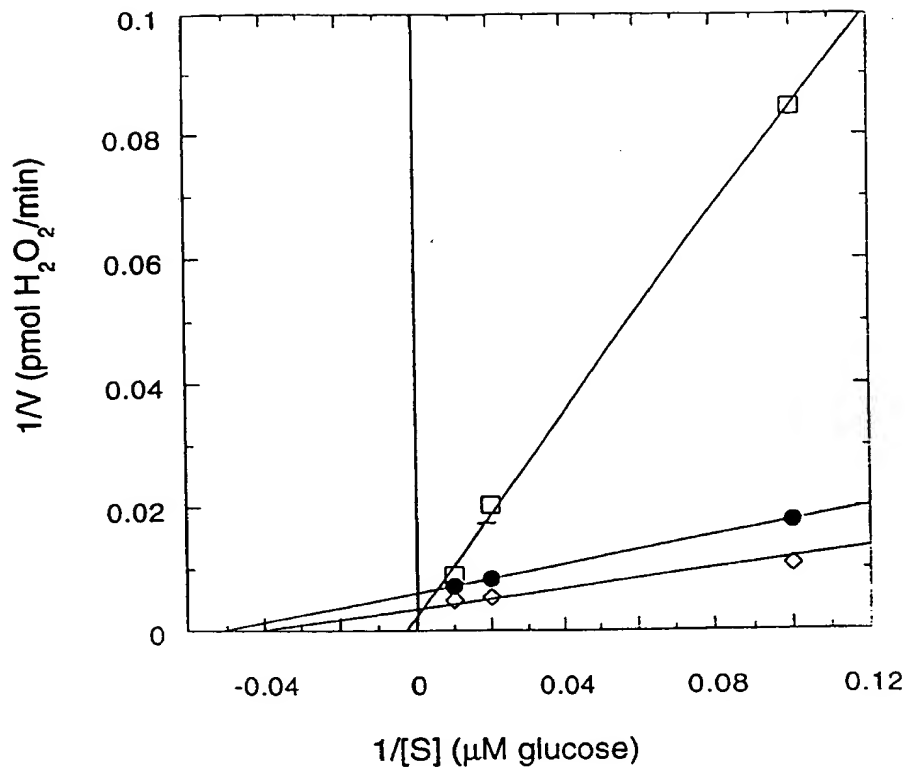


Fig. 8

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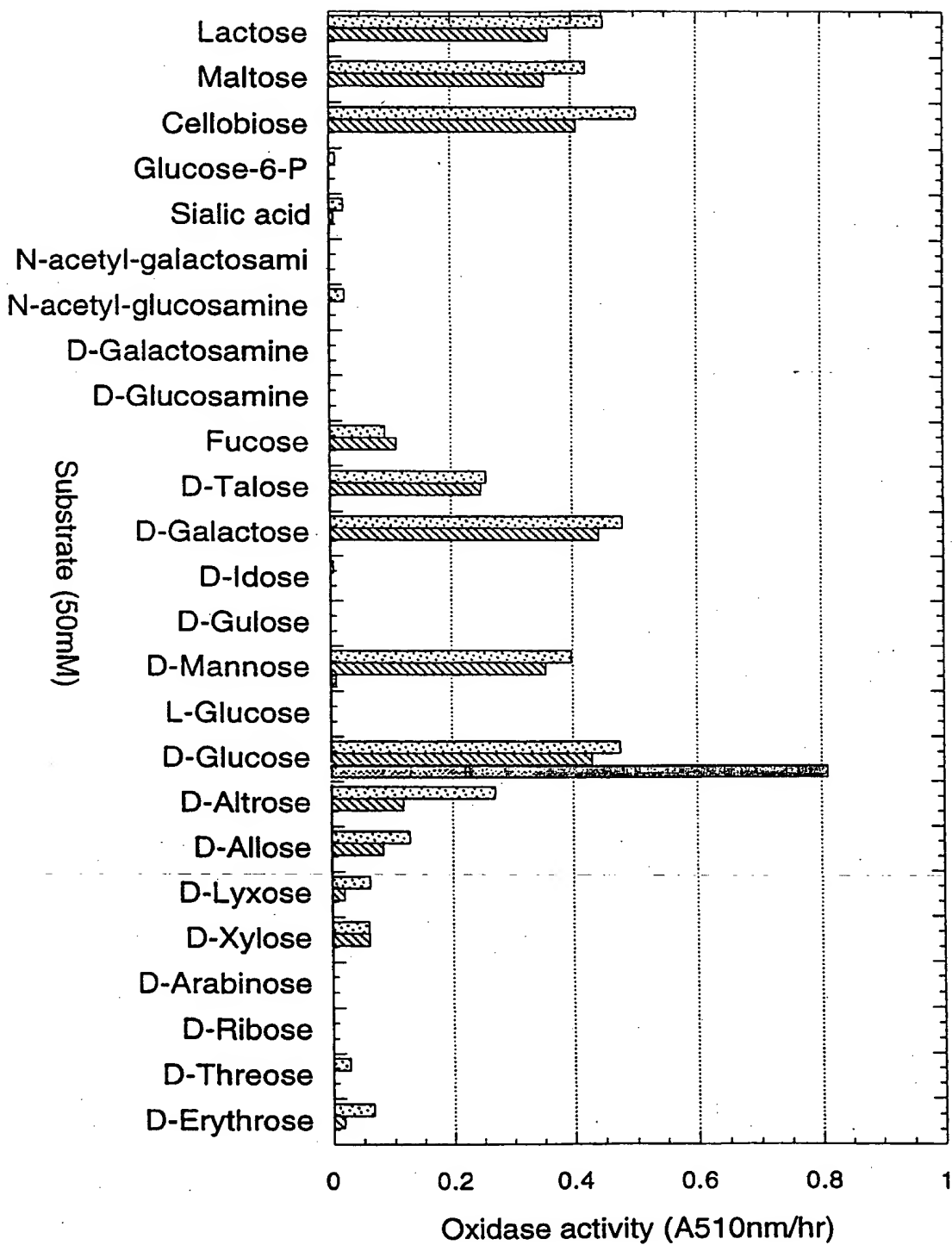


Fig. 9

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MS59	1	-----MQTS	ILTL	LLLL	LLST	QSSATS	RSITD	-R	FIQCLH	DRADPS	SFP
WL64	1	MAITYSFNFKSY	IFPL	LLVL	LLST	THSSATS	TSIID	-R	FQCLNN	RADPS	SFP
At26	1	-----	-----	-----	-----	TSRRNSE	-T	FQCLTS	NSDP	KHP	
At27	1	-----	-----	-----	-----	SIQD	-Q	FINCVR	NTHV	SFP	
EcBBE	1	-----MENKTP	IFFS	LSIF	LSLL	NCA	LG	--ND	--LL	SLCTF	NGVRNE
PsBBE	1	-----MMCRSLT	LRFF	FLFIVL	-LQ	TCVRGGDV	N	DNLL	SSCL	NSHG	VHNF
MS59	42	ITGEVYTPGN	--SS	FPTV	LQNY	IRNL	RFN	ET	TPKP	FLI	ITAEHVSHIQ
WL64	50	LSGQLYTPDN	--SS	FPSV	LQAY	IRNL	RFN	ET	TPKP	ILI	ITALHPSHIQ
At26	23	ISPAIFFSGN	--GS	YSSV	LQAN	IRNL	RFN	ET	TPKP	FLI	IAATHESHVQ
At27	20	LEKTLFTPAKNV	SLFN	QVLE	STAQ	NLQ	FLAK	SM	PKPG	GFI	FRPIHQSQVQ
EcBBE	40	--TVFSADSD	--SD	ENRF	LHLS	IQNP	LE	QNSLI	SKPS	AIIL	PGSKEELS
PsBBE	44	--TTLS	TD	TN	--SD	YFKLL	HA	SMQ	NPLE	AKPT	VSKPSFIVMPGSKEELS
MS59	90	AVVCGKQNR	LLK	TRSG	GH	DYEG	LSYL	TNT	NQ	PFF	IVDMFNLR
WL64	98	AVVCAKTHR	LLM	KTRSG	GH	DYEG	LSYV	TNS	NQ	PFF	IVDMFNLR
At26	71	AITCGKRHN	LQM	KIRSG	GH	DYD	GLSY	VTY	SGK	PFF	IVDMFNLR
At27	70	SIICS	KKLGI	HFR	VRSG	GH	DE	EALSY	VSR	IEK	PEILL
EcBBE	86	TIRCIRKGS	WTI	RLRSG	GH	SYEG	LSYT	SD	T--	PEILL	IDL
PsBBE	90	TVHCCTRES	WTI	RLRSG	GH	SYEG	LSYT	AD	T--	PEVIV	DMN
MS59	140	QETAWVQA	GATL	GEV	YYRIA	EKS	NKH	GFP	PAGV	CPTV	GVGGH
WL64	148	DETAWVQA	GATL	GEV	YYRIA	EKS	NSH	AEP	PAGV	CPTV	GVGGH
At26	121	SKTAWVQT	GAIL	GEV	YYI	WEKS	KT	LAY	PAGI	CPTV	GVGGH
At27	120	SNSAWVQP	GATL	GEL	YYRIA	EKS	KI	HGF	PAGL	C	TEVG
EcBBE	134	SETAWVES	SG	STL	GEL	YYA	IT	ESS	SKL	GFT	AGW
PsBBE	138	SETAWVES	SG	ATL	GEL	YYA	IA	QST	DT	L	GFT
MS59	190	MRKYGLS	VDNI	VD	DAQ	IIDV	NGK	L	LD	RK	SM
WL64	198	MGKYGLS	VDNI	VD	DAQ	IIDV	NGK	L	LN	RK	SM
At26	171	MRKYGLT	VDNT	I	DAR	MV	DV	NGK	I	LD	RK
At27	170	MRKYGLA	GDNV	L	DV	KM	V	D	ANGK	L	LD
EcBBE	184	SRKYGLA	ADNV	V	DAI	L	I	D	ANGA	I	LD
PsBBE	188	SRKYGLA	ADNV	V	DAI	L	I	D	SNGA	I	LD
MS59	240	YKIKLVR	VEV	TV	FT	IER	RE	EON	LS	-T	IAER
WL64	248	YKIKLVR	VEV	TV	FT	IER	RE	EON	LS	-T	IAER
At26	221	YKINLVE	VP	ENV	TV	FR	I	SRT	LE	Q	NAT
At27	220	WKIKLVP	V	PKT	TV	FT	TV	KT	LE	Q	DAR
EcBBE	234	WKIKL	LPV	PEK	V	TV	FR	V	T	K	NVA
PsBBE	238	WKIKL	LPV	PEK	L	TV	FR	V	T	K	NV
MS59	289	SVIND	TNG	-G	KTV	RAI	F	P	T	L	YLG
WL64	297	NVINNT	TNG	-E	KT	I	R	G	L	F	P
At26	270	DVNGT	VSS	Q	KT	V	R	T	F	E	I
At27	270	RAAGND	GN	-K	T	V	T	M	T	Y	L
EcBBE	280	SVLGG	ADE	--K	Q	V	W	L	T	M	L
PsBBE	284	SVLGG	VNG	--N	D	A	W	L	M	F	L

Fig. 10-1

12/12

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MS59      338  WVESVLYYTGFPSTGTPTALLSRTTP-QRLNPFKIKSDYVONPISKROFEF
WL64      346  WIESVLFYTNFPIGTETTALLSRTTP-QRLNPFKIKSDYVKNTISKQGEES
At26      320  WIQSVLFWTNIQVGSSETLLQORN--QPVNYLKRKSDYVREPISTRTGLES
At27      318  WIEAALFHGGFPTGSEIEILLQLKSP LGKDYFKATSD FVKEPIPVIGFKG
EcBBE     328  WGESFAYLAGLET---VSQ LNNRFLKFDERAFKTKVDLTKEPLPSKAFYG
PsBBE     332  WGESMAFLSGLDT---ISE LNNRFLKFDERAFKTKVDFTKVSVP LNVERH

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MS59      1  FIFERLKELENQMLAFNPYGGRMSEISEFAKPPPHRS GNIAKIQYEVNWE
WL64      1  SIFERMKELENQMLAFNPYGGRMSEISEFAKPPPHRS GNIAKIQYEVNWD
At26      1  SIWKKMIELEIPTMAFNPYGGEMGRISSTVTPFPYRAGNLWKIQYGANWR
At27      1  GIEKRLIEGNTTFLNWTPTYGGMMSKIPESAIPEPHRNGTLFKILYANWL
EcBBE     1  GLLERLSKEPNGFIALNGFGGQMSKISSDFTPPPHRS GTRLMVEYIVAWN
PsBBE     1  HALEMGLSEQPGGFIALNGFGGKMSEISTDFTPPPHRKGT KLMFEMI IAWN

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MS59      51  DLSDEAENRYLNFTRLMYDYMTPEVSKNPRKAFLNYRDL DIG- INSHG--
WL64      51  ELGVEAANRYLNFTRVMYDYMTPEVSKNPREAFLNYRDL DIG- VNSHG--
At26      51  DET--LTD RYMELTRKLYQFMTPEVSKNPROSFENYRDVDLG- INSHNG-
At27      51  END-KTSSRKINWIKEIYNYMAPYVSSNPROAYVNYRDL DFG- QNKNN--
EcBBE     51  QSEQKKKTEFLDWLEKVEYEFMKPEVSKNPRLG YVNHI DLDLGG IDWGNKT
PsBBE     51  QDEESKI GEFSEWLAKFYDYLEPEVSKPEPRVG YVNHI DLDLGG IDWRNKS

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MS59      98  -RNAVTEGMV-YGHKYFKETNYKRLVSVKTKVDPDNFFRNEQSIPTLSS-
WL64      98  -KNAYGEGMV-YGHKYFKETNYKRLTMVKTRVDPDNFFRNEQSIPTLSSS
At26      97  KISSYVEGKR-YGKKYFAG-NFERLVKIKTRVDSGNFFRNEQSIPTLSP--
At27      97  AKVNFIEAKI-WGPKYFKG-NEDRLVKIKTKVDPENFFRNEQSIPTLPPY-
EcBBE     101  VVNNNAIEISRWSGESYFLS-NYERLIRAKTLIDPNNVFNHPQSIPP MANF
PsBBE     101  STTNAVEIARNWGERYFSS-NYERLVKAKTLIDPNNVFNHPQSIPPMMKF

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MS59      -----
WL64      146  WK-----
At26      -----
At27      -----
EcBBE     150  D--YLEKTLGSDGGEVVI
PsBBE     150  EEIYMLKEL-----

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Fig. 10-2

BUDAPEST TREATY ON THE INTERNATIONAL
RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

Mogen International N.V.
Einsteinweg 97
2333 CB LEIDEN
Nederland

name and address of depositor

RECEIPT IN THE CASE OF AN ORIGINAL DEPOSIT
issued pursuant to Rule 7.1 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified at the bottom of this page

I. IDENTIFICATION OF THE MICROORGANISM

Identification reference given by the
DEPOSITOR:

E. coli DH5 alpha strain / the plasmid
pMOG800

Accession number given by the
INTERNATIONAL DEPOSITARY AUTHORITY:

CBS 414.93

II. SCIENTIFIC DESCRIPTION AND/OR PROPOSED TAXONOMIC DESIGNATION

The microorganism identified under I above was accompanied by:



a scientific description



a proposed taxonomic designation

(mark with a cross where applicable)

III. RECEIPT AND ACCEPTANCE

This International Depositary accepts the microorganism identified under I above, which was
received by it on Thursday, 12 August 1993 (date of the original deposit)¹

IV. RECEIPT OF REQUEST FOR CONVERSION

The microorganism identified under I above was received by this International Depositary
Authority on not applicable (date of the original deposit) and a
request to convert the original deposit to a deposit under the Budapest Treaty was received by
it on not applicable (date of receipt of request for conversion)

V. INTERNATIONAL DEPOSITARY AUTHORITY

Name: Centraalbureau voor Schimmelcultures

Address: Oosterstraat 1
P.O. Box 273
3740 AG BAARN
The Netherlands

Signature(s) of person(s) having the power to
represent the International Depositary
Authority or of authorized official(s):



drs F.M. van Asma

Date: Friday, 13 August 1993

¹ Where Rule 6.4(d) applies, such date is the date on which the status of international
depositary authority was acquired.

BUDAPEST TREATY ON THE INTERNATIONAL
RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

Mogen International N.V.
Einsteinweg 97
2333 CB LEIDEN
Nederland

*name and address of the party to whom the
viability statement is issued*

VIABILITY STATEMENT

issued pursuant to Rule 10.2 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified on the following page

I. DEPOSITOR	II. IDENTIFICATION OF THE MICROORGANISM
Name: Mogen International N.V. Address: Einsteinweg 97 2333 CB LEIDEN Nederland	Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: CBS 414.93 Date of the deposit or of the transfer: ¹ Thursday, 12 August 1993
III. VIABILITY STATEMENT	
The viability of the microorganism identified under II above was tested on Friday, 13 August 1993 ² . On that date, the said microorganism was <input checked="checked" type="checkbox"/> ³ viable <input type="checkbox"/> ³ no longer viable	

¹ Indicate the date of the original deposit or, where a new deposit or a transfer has been made, the most recent relevant date (date of the new deposit or date of the transfer).

² In the cases referred to in Rule 10.2(a)(ii) and (iii), refer to the most recent viability test.

³ Mark with a cross the applicable box.

IV. CONDITIONS UNDER WHICH THE VIABILITY HAS BEEN PERFORMED⁴

V. INTERNATIONAL DEPOSITARY AUTHORITY

Name: Centraalbureau voor Schimmelcultures

Address: Oosterstraat 1
P.O. Box 273
3740 AG BAARN
The NetherlandsSignature(s) of person(s) having the power to
represent the International Depositary
Authority or of authorized official(s):

drs F.M. van Asma

Date: Friday, 13 August 1993

⁴ Fill in if the information has been requested and if the results of the test were negative.